

Magnetic Luminex[®] Performance Assay

TGF- β Base Kit

Catalog Number LTGM00

For the quantitative determination of transforming growth factor beta 1, 2, and 3 (TGF- β 1, TGF- β 2, and TGF- β 3) concentrations in cell culture supernates, serum, plasma, urine, and human milk.

This package insert must be read in its entirety before using this product.
For research use only. Not for use in diagnostic procedures.

TABLE OF CONTENTS

SECTION	PAGE
INTRODUCTION	1
PRINCIPLE OF THE ASSAY.....	2
LIMITATIONS OF THE PROCEDURE	2
TECHNICAL HINTS.....	2
MATERIALS PROVIDED & STORAGE CONDITIONS	3
PRECAUTIONS.....	4
ADDITIONAL REAGENTS REQUIRED	4
OTHER SUPPLIES REQUIRED	4
SAMPLE COLLECTION AND STORAGE	5
ACTIVATION/NEUTRALIZATION REAGENT PREPARATION	6
SAMPLE ACTIVATION PROCEDURE	6
SAMPLE PREPARATION.....	6
REAGENT PREPARATION.....	7
DILUTED MICROPARTICLE COCKTAIL PREPARATION.....	8
DILUTED BIOTIN ANTIBODY COCKTAIL PREPARATION.....	8
STREPTAVIDIN-PE PREPARATION	8
INSTRUMENT SETTINGS	9
ASSAY PROCEDURE	10
CALCULATION OF RESULTS.....	11
CALIBRATION	11
SPECIFICITY.....	12
REFERENCES	13
PLATE LAYOUT	14

MANUFACTURED AND DISTRIBUTED BY:

USA & Canada | R&D Systems, Inc.

614 McKinley Place NE, Minneapolis, MN 55413, USA
TEL: (800) 343-7475 (612) 379-2956 FAX: (612) 656-4400
E-MAIL: info@RnDSystems.com

DISTRIBUTED BY:

UK & Europe | R&D Systems Europe, Ltd.

19 Barton Lane, Abingdon Science Park, Abingdon OX14 3NB, UK
TEL: +44 (0)1235 529449 FAX: +44 (0)1235 533420
E-MAIL: info@RnDSystems.co.uk

China | R&D Systems China Co., Ltd.

24A1 Hua Min Empire Plaza, 726 West Yan An Road, Shanghai PRC 200050
TEL: +86 (21) 52380373 FAX: +86 (21) 52371001
E-MAIL: info@RnDSystemsChina.com.cn

INTRODUCTION

Transforming Growth Factor Beta 1, 2, and 3 (TGF- β 1, TGF- β 2, and TGF- β 3) are highly pleiotropic cytokines that are secreted by virtually all cell types. TGF- β molecules are proposed to act as cellular switches that regulate processes such as immune function, proliferation, and epithelial-mesenchymal transition (1-4). Targeted deletions of these genes in mice show that each TGF- β isoform has some non-redundant functions: TGF- β 1 is involved in hematopoiesis and endothelial differentiation; TGF- β 2 affects development of cardiac, lung, craniofacial, limb, eye, ear, and urogenital systems; and TGF- β 3 influences palatogenesis and pulmonary development (2). TGF- β ligands are initially synthesized as precursor proteins that undergo proteolytic cleavage. The mature segments form active ligand dimers via a disulfide-rich core consisting of the characteristic "cysteine knot" (1-4). TGF- β signaling begins with binding to a complex of the accessory receptor betaglycan (also known as TGF- β RIII) and a type II serine/threonine kinase receptor termed TGF- β RII. This receptor then phosphorylates and activates a type I serine/threonine kinase receptor, either ALK-1 or TGF- β RI (also called ALK-5). The activated type I receptor phosphorylates and activates Smad proteins that regulate transcription (3, 5, 6). Use of other signaling pathways that are Smad-independent allows for distinct actions observed in response to TGF- β in different contexts (5).

This kit can be used to assess the levels of three TGF- β molecules. Due to high cross-species homology, human, mouse, rat, and porcine samples can be run in this assay.

Due to a low level of cross-reactivity of the biotinylated anti-TGF- β 1 antibody with TGF- β 2, multiplexing TGF- β 1 and TGF- β 2 can result in falsely elevated values for TGF- β 2 in some sample types. We found in human serum samples, falsely elevated values for TGF- β 2 when multiplexing TGF- β 1 and TGF- β 2. Similar cross-reactivity was observed in competitor's multiplex assays. For studies using human serum samples, TGF- β 1 and TGF- β 3, or TGF- β 2 and TGF- β 3 can be multiplexed; however, careful consideration should be given when deciding to evaluate TGF- β 1 and TGF- β 2 as a multiplex. Testing with samples from healthy donors found the impact of the cross-reactivity observed became insignificant with human milk, platelet-poor plasma, cell culture supernates and urine samples. It is suggested that each lab evaluate their samples to determine if multiplexing will be problematic (as TGF- β levels will vary with disease state and serum source for media).

Analyte	Catalog Number	Microparticle Region
TGF- β 1	LTGM100	34
TGF- β 2	LTGM200	15
TGF- β 3	LTGM300	22

PRINCIPLE OF THE ASSAY

Magnetic Luminex® Performance Assay multiplex kits are designed for use with the Luminex MAGPIX® CCD Imager. Alternatively, kits can be used with the Luminex 100™, Luminex 200™ or Bio-Rad® Bio-Plex®, dual laser, flow-based sorting and detection platforms.

Analyte-specific antibodies are pre-coated onto color-coded magnetic microparticles. Microparticles, standards and samples are pipetted into wells and the immobilized antibodies bind the analytes of interest. After washing away any unbound substances, a biotinylated antibody cocktail specific to the analytes of interest is added to each well. Following a wash to remove any unbound biotinylated antibody, streptavidin-phycoerythrin conjugate (Streptavidin-PE), which binds to the biotinylated antibody, is added to each well. A final wash removes unbound Streptavidin-PE, the microparticles are resuspended in buffer and read using the Luminex MAGPIX Analyzer. A magnet in the analyzer captures and holds the superparamagnetic microparticles in a monolayer. Two spectrally distinct Light Emitting Diodes (LEDs) illuminate the microparticle. One LED identifies the analyte that is being detected and the second LED determines the magnitude of the PE-derived signal, which is in direct proportion to the amount of analyte bound. Each well is imaged with a CCD camera.

LIMITATIONS OF THE PROCEDURE

- FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.
- The kit should not be used beyond the expiration date on the kit label.
- Do not mix or substitute reagents with those from other lots or sources.
- If samples fall outside the dynamic range of the assay, further dilute the samples with Calibrator Diluent and repeat the assay.
- Any variation in standard diluent, operator, pipetting technique, washing technique, incubation time or temperature, and kit age can cause variation in binding.
- Variations in sample collection, processing, and storage may cause sample value differences.
- This assay is designed to eliminate interference by other factors present in biological samples. Until these factors have been tested in the Luminex Performance Assay, the possibility of interference cannot be excluded.
- Magnetic Luminex Performance Assays afford the user the benefit of multianalyte analysis in a single complex sample. For each sample type, a single multipurpose diluent is used to optimize recovery, linearity, and reproducibility. Such a multipurpose diluent may not optimize any single analyte to the same degree that a unique diluent selected for analysis of that analyte can optimize conditions, therefore, some performance characteristics may be more variable than those for assays designed specifically for single analyte analysis.
- **Only the analytes listed on the Standard Value Card can be measured with this base kit.**

TECHNICAL HINTS

- When mixing or reconstituting protein solutions, always avoid foaming.
- To avoid cross-contamination, change pipette tips between additions of each standard level, between sample additions, and between reagent additions. Also, use separate reservoirs for each reagent.
- To ensure accurate results, proper adhesion of plate sealers during incubation steps is necessary.
- Protect microparticles and Streptavidin-PE from light at all times to prevent photobleaching.

MATERIALS PROVIDED & STORAGE CONDITIONS

Store the unopened kit at 2-8 °C. Do not use past the kit expiration date.

PART	PART #	DESCRIPTION	STORAGE OF OPENED, DILUTED, OR RECONSTITUTED MATERIAL
TGF- β Standard Cocktail	893244	2 vials of recombinant TGF- β 1, 2, and 3 in a buffered protein base with preservatives; lyophilized.	Discard after use. Use a fresh standard for each assay.
Biotin Antibody Diluent 2	895832	5.5 mL of a buffered protein base with preservative.	May be stored for up to 1 month at 2-8 °C.*
Microparticle Diluent 2	895815	6 mL of a buffered protein base with preservatives.	
Calibrator Diluent RD5-49	895584	21 mL of a buffered protein base with preservatives. <i>For cell culture supernate/urine samples.</i>	
Calibrator Diluent RD6-50	895912	21 mL of a buffered delipidized animal serum with preservatives. <i>For serum/plasma/human milk samples.</i>	
Streptavidin-PE	892525	0.07 mL of a 100-fold concentrated streptavidin-phycoerythrin conjugate with preservatives.	
Wash Buffer Concentrate	895003	21 mL of a 25-fold concentrated solution of buffered surfactant with preservative. <i>May turn yellow over time.</i>	
Microplate	641385	1 flat-bottomed 96-well microplate used as a vessel for the assay.	
Mixing Bottles	895505	2 empty 8 mL bottles used for mixing microparticles with Microparticle Diluent.	
Plate Sealers	640445	4 adhesive foil strips.	
Standard Value Card	749814	1 card listing the Standard reconstitution volume and working standard concentrations for this lot of base kit.	

*Provided this is within the expiration date of the kit.

PRECAUTIONS

Some components in this kit contain a preservative which may cause an allergic skin reaction. Avoid breathing mist.

Wear protective gloves, clothing, eye, and face protection. Wash hands thoroughly after handling. Please refer to the MSDS on our website prior to use.

ADDITIONAL REAGENTS REQUIRED

For sample activation:

- Hydrochloric acid (A.C.S. Grade, 12 N)
- Sodium hydroxide (A.C.S. Grade, 10 N)
- HEPES, free acid (Reagent Grade, M.W. 238.3)

OTHER SUPPLIES REQUIRED

- **Luminex Performance Assay analyte-specific kit(s) (see Introduction on page 1).**
- Luminex MAGPIX, Luminex 100/200, or Bio-Rad Bio-Plex analyzer with X-Y platform.
- Hand-held microplate magnet or platewasher with a magnetic platform.
- Pipettes and pipette tips.
- Deionized or distilled water.
- Multi-channel pipette, manifold dispenser, or automated dispensing unit.
- 100 mL and 500 mL graduated cylinders.
- Horizontal orbital microplate shaker (0.12" orbit) capable of maintaining a speed of 800 ± 50 rpm.
- Microcentrifuge.
- **Polypropylene** test tubes for dilution of standards and activation of samples.

SAMPLE COLLECTION AND STORAGE

The sample collection and storage conditions listed below are intended as general guidelines. Sample stability has not been evaluated.

Samples must be assayed immediately after activation. Do not freeze activated samples.

Cell Culture Supernates - Remove particulates by centrifugation and assay (see activation procedure) immediately or aliquot and store unactivated samples at $\leq -70^{\circ}\text{C}$. Avoid repeated freeze-thaw cycles.

Notes: *Animal serum used in the preparation of cell culture media may contain high levels of latent TGF- β 1. For best results, do not use animal serum for growth of cell cultures when assaying for TGF- β 1 production. If animal serum is used as a supplement in the media, precautions should be taken to prepare the appropriate control and run the control in the assay to determine the baseline concentration of TGF- β 1.*

A 10% solution of bovine serum used in cell culture can be expected to contribute typically 100-200 pg/mL of bovine TGF- β 2 upon activation. Prepare, activate, and assay an appropriate media control to determine the baseline bovine TGF- β 2 in the sample.

Serum - Use a serum separator tube (SST) and allow samples to clot for 30 minutes at room temperature. Centrifuge for 15 minutes at 1000 x g. Remove serum and assay (see activation procedure) immediately or aliquot and store unactivated samples at $\leq -70^{\circ}\text{C}$. Avoid repeated freeze-thaw cycles.

Platelet-poor Plasma* - Collect plasma on ice using EDTA or heparin as an anticoagulant. Centrifuge for 15 minutes at 1000 x g within 30 minutes of collection. An additional centrifugation step of the plasma at 10,000 x g for 10 minutes at 2-8 $^{\circ}\text{C}$ is recommended for complete platelet removal. Assay (see activation procedure) immediately or aliquot and store unactivated samples at $\leq -70^{\circ}\text{C}$. Avoid repeated freeze-thaw cycles.

Note: *Grossly hemolyzed or lipemic samples are not suitable for the measurement of TGF- β in this assay.*

**TGF- β 1 is present in platelet granules and is released upon platelet activation. Therefore, to measure circulating levels of TGF- β 1, platelet-poor plasma should be collected for measurement. It should be noted that many protocols for plasma preparation, including procedures recommended by the Clinical and Laboratory Standards Institute (CLSI), result in incomplete removal of platelets from blood. This will cause variable and irreproducible results for assays of factors contained in platelets and released by platelet activation. The recommended plasma collection protocol is designed to minimize platelet degranulation. However, since even the best methods for plasma collection may result in some platelet degranulation on occasion, it is recommended that markers for platelet degranulation be determined in samples containing elevated TGF- β 1 levels.*

Urine - Aseptically collect the first urine of the day (mid-stream), voided directly into a sterile container. Centrifuge to remove particulate matter. Assay immediately (see activation procedure) or aliquot and store unactivated samples at $\leq -70^{\circ}\text{C}$. Avoid repeated freeze/thaw cycles.

Note: *Neat unactivated urine samples exhibit a decrease in TGF- β 1 concentration in the first 24 hours of storage (frozen or refrigerated). Care should be taken that all samples are assayed under identical storage conditions and durations.*

Human Milk - Centrifuge for 15 minutes at 1000 x g at 2-8 $^{\circ}\text{C}$. Collect the aqueous fraction and repeat this process a total of 3 times. Assay immediately or aliquot and store samples at $\leq -20^{\circ}\text{C}$. Avoid repeated freeze-thaw cycles.

ACTIVATION/NEUTRALIZATION REAGENT PREPARATION

The following solutions may be stored at room temperature for up to one month in polypropylene bottles.

Caution: *Wear protective clothing and safety glasses during preparation or use of these reagents. Refer to the appropriate MSDS prior to use.*

1N HCl (100 mL) - Slowly add 8.33 mL of 12N HCl to 91.67 mL of deionized water. Mix well.

1.2N NaOH/0.5M HEPES (100 mL) - Slowly add 12 mL of 10N NaOH to 75 mL of deionized water. Mix well, and add 11.9 g of HEPES. Mix well, and bring the final volume to 100 mL with deionized water.

SAMPLE ACTIVATION PROCEDURE

To activate latent TGF- β to immunoreactive TGF- β detectable in this assay, follow the activation procedure outlined below for all sample types. Assay samples immediately after neutralization (pH 7.2-7.6). **Use polypropylene test tubes. Do not activate the TGF- β Standard Cocktail.**

1. If samples contain particulates, centrifuge for 2 minutes at 5000 x g before proceeding.
2. Add 20 μ L of 1N HCl to 100 μ L of sample. Mix well, and incubate for 10 minutes at room temperature.
3. Add 20 μ L of 1.2N NaOH/0.5M HEPES. Mix thoroughly.

SAMPLE PREPARATION

Activated cell culture supernates and activated urine samples require a 1:3.57 dilution in Calibrator Diluent RD5-49 after activation. This is a 1:5 final dilution factor. A suggested 1:3.57 dilution is 112 μ L of activated/neutralized sample + 288 μ L of Calibrator Diluent RD5-49. Mix thoroughly.

Activated serum, plasma, and human milk samples require a 1:10.7 dilution in Calibrator Diluent RD6-50 after activation. This is a 1:15 final dilution factor. A suggested 1:10.7 dilution is 50 μ L of activated/neutralized sample + 485 μ L of Calibrator Diluent RD6-50. Mix thoroughly.

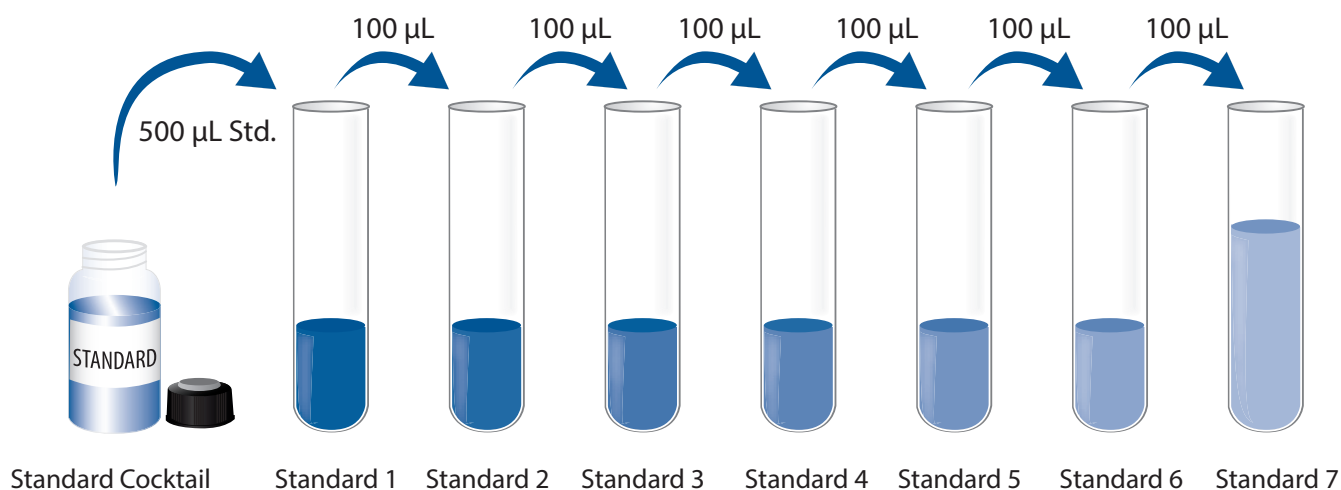
REAGENT PREPARATION

Bring all reagents to room temperature before use.

Wash Buffer - If crystals have formed in the concentrate, warm to room temperature and mix gently until the crystals have completely dissolved. Add 20 mL of Wash Buffer Concentrate to deionized or distilled water to prepare 500 mL of Wash Buffer.

Standard - Reconstitute the Standard Cocktail with Calibrator Diluent RD5-49 (*for cell culture supernate and urine samples*) or Calibrator Diluent RD6-50 (*for serum, plasma, and human milk samples*). Refer to the Standard Card for the reconstitution volume and assigned values. Allow the standard to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions.

Use polypropylene tubes. Pipette 500 μL of the reconstituted Standard into the Standard 1 tube. Pipette 200 μL of the appropriate Calibrator Diluent into the remaining tubes. Use Standard 1 to produce a 3-fold dilution series (below). Mix each tube thoroughly before the next transfer. Standard 1 serves as the high standard. The appropriate Calibrator Diluent serves as the blank.



DILUTED MICROPARTICLE COCKTAIL PREPARATION

1. Centrifuge each Microparticle Concentrate vial for 30 seconds at 1000 x g prior to removing the cap.
2. Gently vortex the vials to resuspend the microparticles, taking precautions not to invert the vials.
3. Dilute the Microparticle Concentrates in the mixing bottle provided. The volume of the Microparticle Concentrate listed in the table below is for each analyte (e.g. if measuring a full plate, add 50 μ L of each Microparticle Concentrate to 5 mL of Microparticle Diluent).

Number of Wells Used	Microparticle Concentrate	+	Microparticle Diluent
96	50.0 μ L	+	5.00 mL
72	37.5 μ L	+	3.75 mL
48	25.0 μ L	+	2.50 mL
24	12.5 μ L	+	1.25 mL

Note: Protect microparticles from light during handling. Diluted microparticles cannot be stored. Prepare microparticles within 30 minutes of use.

DILUTED BIOTIN ANTIBODY COCKTAIL PREPARATION

1. Centrifuge each Biotin Antibody Concentrate vial for 30 seconds at 1000 x g prior to removing the cap.
2. Gently vortex the vials, taking precautions not to invert the vials.
3. Add 50 μ L of each Biotin Antibody Concentrate to the Biotin Antibody Diluent. Mix gently.

STREPTAVIDIN-PE PREPARATION

Use a polypropylene amber bottle or a polypropylene tube wrapped with aluminum foil. Protect Streptavidin-PE from light during handling and storage.

1. Centrifuge the Streptavidin-PE vial for 30 seconds at 1000 x g prior to removing the cap.
2. Gently vortex the vial, taking precautions not to invert the vial.
3. Dilute the 100X Streptavidin-PE to a 1X concentration by adding 55 μ L of Streptavidin-PE to 5.5 mL of Wash Buffer.

INSTRUMENT SETTINGS

Luminex MAGPIX analyzer:

- a) Assign the microparticle region for each analyte being measured (see page 1)
- b) 50 events/bead
- c) Sample size: 50 μ L
- d) Collect Median Fluorescence Intensity (MFI)

Luminex 100/200 and Bio-Rad Bio-Plex analyzers:

Note: *Calibrate the analyzer using the proper reagents for superparamagnetic microparticles (refer to instrument manual).*

- a) Assign the microparticle region for each analyte being measured (see page 1)
- b) 50 events/bead
- c) Minimum events: 0
- d) Flow rate: 60 μ L/minute (fast)
- e) Sample size: 50 μ L
- f) Doublet Discriminator gates at approximately 8000 and 16,500
- g) Collect MFI

Note: *The CAL2 setting for the Bio-Rad Bio-Plex analyzer should be set at the low RP1 target value.*

ASSAY PROCEDURE

Bring all reagents and samples to room temperature before use. It is recommended that all samples and standards be assayed in duplicate.

Note: *Protect microparticles and Streptavidin-PE from light at all times.*

1. Prepare all reagents, working standards, and samples as directed in the previous sections.
2. Add 50 μ L of Standard or sample* per well.
3. Resuspend the diluted microparticle cocktail by inversion or vortexing. Add 50 μ L of the microparticle cocktail to each well of the microplate. Securely cover with a foil plate sealer. Incubate for 2 hours at room temperature on a horizontal orbital microplate shaker (0.12" orbit) set at 800 ± 50 rpm. A plate layout is provided to record standards and samples assayed.
4. Using a magnetic device designed to accommodate a microplate, wash by applying the magnet to the bottom of the microplate, removing the liquid, filling each well with Wash Buffer (100 μ L) and removing the liquid again. Removal of liquid is essential for good performance. A small amount of residual liquid (10-15 μ L) will not interfere in this assay. Perform the wash procedure three times.

Note: *Refer to the magnetic device user manual for proper wash technique using a round bottom microplate.*
5. Add 50 μ L of diluted Biotin Antibody Cocktail to each well. Securely cover with a foil plate sealer and incubate for 1 hour at room temperature on the shaker set at 800 ± 50 rpm.
6. Repeat the wash as in step 4.
7. Add 50 μ L of Streptavidin-PE to each well. Securely cover with a foil plate sealer and incubate for 30 minutes at room temperature on the shaker set at 800 ± 50 rpm.
8. Repeat the wash as in step 4.
9. Resuspend the microparticles by adding 100 μ L of Wash Buffer to each well. Incubate for 2 minutes at room temperature on the shaker set at 800 ± 50 rpm.
10. Read within 90 minutes using a Luminex or Bio-Rad Analyzer.

*Samples require activation and dilution. See Sample Activation and Sample Preparation sections.

CALCULATION OF RESULTS

Use the Standard concentrations on the Standard Value Card and calculate 3-fold dilutions for the remaining levels. Average the duplicate readings for each standard and sample and subtract the average blank Median Fluorescence Intensity (MFI).

Create a standard curve for each analyte by reducing the data using computer software capable of generating a five parameter logistic (5-PL) curve-fit.

Since samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

CALIBRATION

This assay is calibrated against highly purified recombinant human TGF- β 1, TGF- β 2, and TGF- β 3 produced at R&D Systems.

SPECIFICITY

This assay recognizes natural and recombinant TGF- β s.

The factors listed below were prepared at 50 ng/mL in Calibrator Diluent, assayed, and measured less than 0.5% cross-reactivity unless otherwise noted. The same factors prepared at 50 ng/mL in a mid-range recombinant TGF- β control were assayed for interference. No interference was observed except as noted.

Recombinant human:

6Ckine	HCC-1
Activin A	HCC-4
Activin RIA	I-309
Activin RIIA	IL-8
Activin RIIIB	Inhibin A
BLC/BCA-1	Inhibin B
BMP-2	IP-10
BMP-3	I-TAC
BMP-3b	LAP
BMP-4	MCP-1
BMP-5	MCP-2
BMP-6	MCP-3
BMP-8b	MCP-4
BMP-10	MDC
BMP-15	MIG
BMP RIA	MIP-1 α
BMP RIB	MIP-1 β
BMP RII	MIP-1 δ
CTACK	MIP-3 α
ENA-78	MIP-3 β
Eotaxin	MPIF-1
Eotaxin-2	NAP-2
Eotaxin-3	PARC
Follistatin288	RANTES
Follistatin300	SDF-1 α
Follistatin315	SDF-1 β
GCP-2	TECK
GRO α	TGF- α
GRO β	TGF- β RII
GRO γ	

Recombinant mouse:

6Ckine
BLC/BCA-1
BMP-3b
BMP RIA
BMP RIB
CRG-2/IP-10
CTACK
Eotaxin
GCP-2
JE/MCP-1
KC
MARC
MCP-5
MDC
MIG
MIP-1 α
MIP-1 β
MIP-1 γ
RANTES
SDF-1 α
TARC
TECK

Recombinant rat:

Agrin
MIP-3 α

Other recombinants:

porcine IL-8
amphibian TGF- β 5

Recombinant human TGF- β 1.2 exhibits 4.5% cross-reactivity in this assay.

Recombinant human TGF- β RIII interferes at concentrations \geq 50 ng/mL.

REFERENCES

1. Derynck, R. and K. Miyazono (2008) in *The TGF- β Family*, Derynck, R. and K. Miyazono eds., Cold Spring Harbor Laboratory Press, p. 29.
2. Dunker, N. and K. Krieglstein (2000) *Eur. J. Biochem.* **267**:6982.
3. Wahl, S.M. (2006) *Immunol. Rev.* **213**:213.
4. Chang, H. *et al.* (2002) *Endocr. Rev.* **23**:787.
5. de Caestecker, M. *et al.* (2004) *Cytokine Growth Factor Rev.* **15**:1.
6. Zuniga, J.E. *et al.* (2005) *J. Mol. Biol.* **354**:1052.

All trademarks and registered trademarks are the property of their respective owners.

PLATE LAYOUT

Use this plate layout to record standards and samples assayed.

12								
11								
10								
9								
8								
7								
6								
5								
4								
3								
2								
1								
	A	B	C	D	E	F	G	H