DuoSet® IC

Human/Mouse/Rat Total p38α

Catalog Number DYC8691-2 DYC8691-5

For the development of sandwich ELISAs to measure human, mouse, and rat total p38 α in cell lysates.

This package insert must be read in its entirety before using this product.

FOR RESEARCH USE ONLY.
NOT FOR USE IN DIAGNOSTIC PROCEDURES.

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PRINCIPLE OF THE ASSAY

This DuoSet [®] IC ELISA contains the basic components required for the development of sandwich ELISAs to measure p38 mitogen-activated protein kinase alpha (p38 α) in cell lysates. An immobilized capture antibody specific for p38 α , also known as stress-activated protein kinase 2A (SAPK2A), CSAID-binding protein 1 (CSBP1), and MAPK14, binds both phosphorylated and unphosphorylated p38 α . After washing away unbound material, a biotinylated detection antibody specific for total p38 α is used to detect both phosphorylated and non-phosphorylated protein, utilizing a standard Streptavidin-HRP format. Cross-reactivity with other p38 family members (p38 β , p38 γ , and p38 δ), and other MAPKs is minimal (see page 8). This DuoSet IC ELISA has been demonstrated to work with lysates prepared from human, mouse, and rat cells (see pages 8 - 9).

MATERIALS PROVIDED

Store the unopened kit at 2 - 8° C. Do not use past kit expiration date.

			Vials Provided	
Description	Part #	Storage Conditions	Cat. # DYC8691-2	Cat. # DYC8691-5
Total p38α Capture Antibody	841064	2 - 8° C	1	2
Total p38α Detection Antibody	841065	2 - 8° C	1	2
Total p38 α Standard	841066	2 - 8° C	3	5
Streptavidin-HRP	890803	2 - 8° C	1	1

DYC8691-2 contains sufficient materials to run ELISAs on at least two 96 well plates.*

DYC8691-5 contains sufficient materials to run ELISAs on at least five 96 well plates.*

*Provided the following conditions are met:

- The reagents are prepared as described in this package insert.
- The assay is run as described in the General ELISA Protocol on page 6.
- The recommended microplates, buffers, diluents, substrates, and solutions are used.

OTHER MATERIALS REQUIRED

- Aprotinin (Sigma # A6279)
- Leupeptin (Sigma # L8511)
- Pepstatin (Sigma # P4265)
- Phenylmethylsulfonylfluoride (PMSF) (Sigma # P7626)
- Sodium Fluoride (NaF) (Sigma # S6521)
- Triton X-100 (Sigma # T9284)
- Urea
- Pipettes and pipette tips
- Deionized or distilled water
- 96 well microplates [Costar EIA Plates (Catalog # 2592 or R&D Systems' Catalog # DY990) are suggested]
- Plate sealers (R&D Systems, Catalog # DY992)
- Squirt bottle, manifold dispenser, or automated microplate washer

SOLUTIONS REQUIRED

PBS - 137 mM NaCl, 2.7 mM KCl, 8.1 mM Na₂HPO₄, 1.5 mM KH₂PO₄, pH 7.2 - 7.4, 0.2 μ m filtered.

Wash Buffer - 0.05% Tween[®] 20 in PBS, pH 7.2 - 7.4 (R&D Systems, Catalog # WA126).

Block Buffer - 1% BSA*, 0.05% NaN3, in PBS, pH 7.2 - 7.4.

IC Diluent #1 - 1% BSA* in PBS, pH 7.2 - 7.4, 0.2 μm filtered.

IC Diluent #8 - 1 mM EDTA, 0.5% Triton X-100, 5 mM NaF in PBS, pH 7.2 - 7.4.

Note: *IC Diluent #8 is also the base diluent for IC Diluent #3, IC Diluent #7, and Lysis Buffer #3. Approximately 50 mL of this diluent is required to run the assay on one plate.*

IC Diluent #3 - 1 mM EDTA, 0.5% Triton X-100, 5 mM NaF, 1 M urea in PBS, pH 7.2 - 7.4.

IC Diluent #7 - 1 mM EDTA, 0.5% Triton X-100, 5 mM NaF, 6 M urea in PBS, pH 7.2 - 7.4.

Lysis Buffer #3 - 1 mM EDTA, 0.5% Triton X-100, 5 mM NaF, 6 M urea, 25 μ g/mL Leupeptin, 25 μ g/mL Pepstatin, 100 μ M PMSF, 3 μ g/mL Aprotinin in PBS, pH 7.2 - 7.4.

Substrate Solution - 1:1 mixture of Color Reagent A (H₂O₂) and Color Reagent B (Tetramethylbenzidine) (R&D Systems, Catalog # DY999).

Stop Solution - 2 N H₂SO₄ (R&D Systems, Catalog # DY994).

*The use of Serological Proteins Inc. Bovine Serum Albumin, Fraction V, Protease-free (Cat. # 82-045) or Sigma Bovine Serum Albumin (Cat. # A7030) is recommended. All buffers containing BSA must be stored at 2 - 8° C.

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REAGENT PREPARATION

Bring all reagents to room temperature before use.

Total p38 α **Capture Antibody** (Part 841064) - 720 μ g/mL of mouse anti-human p38 α antibody when reconstituted with 200 μ L of PBS. After reconstitution, store at 2 - 8° C for up to 30 days or aliquot and store at \leq -20° C in a manual defrost freezer or at \leq -70° C for up to 3 months.*

Total p38 α **Detection Antibody** (Part 841065) - 7.2 µg/mL of biotinylated rabbit anti-human p38 α antibody when reconstituted with 1.0 mL of IC Diluent #1. After reconstitution, store at 2 - 8° C for up to 30 days or aliquot and store at \leq -20° C in a manual defrost freezer or at \leq -70° C for up to 3 months.*

Total p38 α Standard (Part 841066) - 280 ng/mL of recombinant human p38 α when reconstituted with 500 μ L of IC Diluent #7. **A fresh standard should be used for each assay.** Immediately before use, an initial 6-fold dilution should be made in IC Diluent #8. Additional dilutions should be made in IC Diluent #3. A seven point standard curve using 2-fold serial dilutions and a high standard of 20 ng/mL is recommended.

Streptavidin-HRP (Part 890803) - 1 mL of Streptavidin conjugated to horseradish-peroxidase. Store at 2 - 8° C. **DO NOT FREEZE.**

*Provided this is within the expiration date of the kit.

PREPARATION OF SAMPLES

Cell lysates - Rinse cells two times with PBS, making sure to remove any remaining PBS after the second rinse. Solubilize cells at 1 x 10^7 cells/mL in Lysis Buffer #3. Vortex lysates briefly and allow to sit at room temperature for 1 hour before use or store at \leq -20° C in a manual defrost freezer. Sample protein concentration may be quantified using a total protein assay. Before use, centrifuge at 2000 x g for 5 minutes and transfer the supernate into a clean test tube. For assaying, dilute lysates 6-fold with IC Diluent #8 and make further serial dilutions in IC Diluent #3.

Note: The final concentration of urea in all samples and standards should be 1 M prior to addition to the plate.

PRECAUTION

The Stop Solution suggested for use with this kit is an acidic solution. Wear eye, hand, face, and clothing protection when using this material.

TECHNICAL HINTS AND LIMITATIONS

- This DuoSet IC ELISA should not be used beyond the expiration date on the kit label.
- Individual results may vary due to differences in technique, plasticware and water sources.
- It is important that the diluents selected for reconstitution and for dilution of the standard reflect the environment of the samples being measured. The diluents suggested in this protocol should be suitable for most cell lysates.
- The type of enzyme and substrate and the concentrations of capture/detection antibodies used can be varied to create an immunoassay with a different sensitivity and dynamic range. A basic understanding of immunoassay development is required for the successful use of these reagents in immunoassays.
- A thorough and consistent wash technique is essential for proper assay performance. Wash Buffer should be dispensed forcefully and removed completely from the wells by aspiration or decanting. Remove any remaining Wash Buffer by inverting the plate and blotting it against clean paper towels.
- Use a fresh reagent reservoir and pipette tips for each step.
- It is recommended that all standards and samples be assayed in duplicate.
- Avoid microbial contamination of reagents and buffers. This may interfere with the sensitivity of the assay. Buffers containing protein should be made under sterile conditions and stored at 2 - 8° C or be prepared fresh daily.

GENERAL ELISA PROTOCOL

A plate layout is provided to record standards and samples assayed.

Plate Preparation

- 1. Dilute the Capture Antibody to the working concentration of 4.0 μ g/mL in PBS without carrier protein. Immediately coat a 96 well microplate with 100 μ L per well of the diluted Capture Antibody. Seal the plate and incubate overnight at room temperature.
- 2. Aspirate each well and wash with Wash Buffer, repeating the process two times for a total of 3 washes. Wash by filling each well with Wash Buffer (400 μL) using a squirt bottle, manifold dispenser or autowasher. Complete removal of liquid at each step is essential for good performance. After the last wash, remove any remaining Wash Buffer by aspirating or by inverting the plate and blotting it against clean paper towels.
- 3. Block plates by adding 300 μ L of Block Buffer to each well. Incubate at room temperature for 1 2 hours.
- 4. Repeat the aspiration/wash as in step 2. The plates are now ready for sample addition.

Assay Procedure

- 1. Add 100 μ L of sample or standards in IC Diluent #3 per well. Use IC Diluent #3 as the zero standard. Cover with a plate sealer and incubate 2 hours at room temperature.
 - **Note:** A seven point standard curve using 2-fold serial dilutions and a high standard of 20 ng/mL is recommended.
- 2. Repeat the aspiration/wash as in step 2 of Plate Preparation.
- 3. Dilute the Detection Antibody to a working concentration of 200 ng/mL in IC Diluent #1 immediately before use. Add 100 μ L of the diluted Detection Antibody to each well. Cover with a new plate sealer and incubate 2 hours at room temperature.
- 4. Repeat the aspiration/wash as in step 2 of Plate Preparation.
- 5. Immediately before use, dilute the Streptavidin-HRP to the working concentration specified on the vial label using IC Diluent #1. Add 100 μ L of the diluted Streptavidin-HRP to each well. Incubate for 20 minutes at room temperature. Avoid placing the plate in direct light.
- 6. Repeat the aspiration/wash as in step 2 of Plate Preparation.
- 7. Add 100 μ L of Substrate Solution to each well. Incubate for 20 minutes at room temperature. Avoid placing the plate in direct light.
- 8. Add 50 μ L of Stop Solution to each well. Gently tap the plate to ensure thorough mixing.
- 9. Determine the optical density of each well immediately, using a microplate reader set to 450 nm. If wavelength correction is available, set to 540 nm or 570 nm. If wavelength correction is not available, subtract readings at 540 nm or 570 nm from the readings at 450 nm. This subtraction will correct for optical imperfections in the plate. Readings made directly at 450 nm without correction may be higher and less accurate.

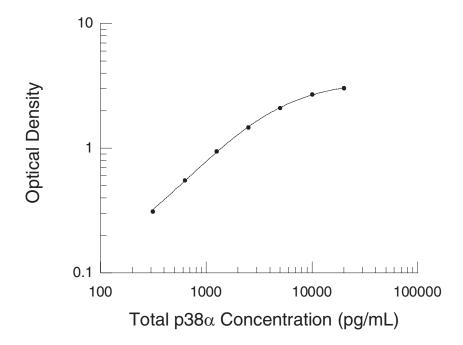
CALCULATION OF RESULTS

Average the duplicate readings for each standard and sample and subtract the average zero standard optical density. Results may be normalized to total protein or cell number.

Create a standard curve by reducing the data using computer software capable of generating a four parameter logistic (4-PL) curve-fit. As an alternative, construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph. The data may be linearized by plotting the log of the p38 α concentrations versus the log of the O.D. and the best fit line can be determined by regression analysis. This procedure will produce an adequate but less precise fit of the data.

TYPICAL DATA

A standard curve should be generated for each set of samples assayed. The graph below represents typical data generated when using the Human/Mouse/Rat Total p38 α DuoSet IC ELISA. The standard curve was calculated using a computer generated 4-PL curve-fit. This standard curve is for demonstration purposes only.



CALIBRATION

This DuoSet IC ELISA is calibrated against a highly purified *E. coli*-expressed recombinant human p38 α produced at R&D Systems.

SPECIFICITY

This DuoSet IC ELISA specifically recognizes p38 α . Specificity was demonstrated using cross-reactivity experiments. Recombinant human p38 β , p38 γ , and p38 δ were assayed at 200 ng/mL and did not cross-react or interfere. Recombinant human ERK2 and JNK1 were also assayed at 200 ng/mL and did not cross-react or interfere.

QUANTIFICATION

Amounts of p38 α , as quantified by the this DuoSet IC ELISA, are consistent with the amounts of p38 α determined by qualitative Western blot analysis.

Quantification of p38 α in human cell lines

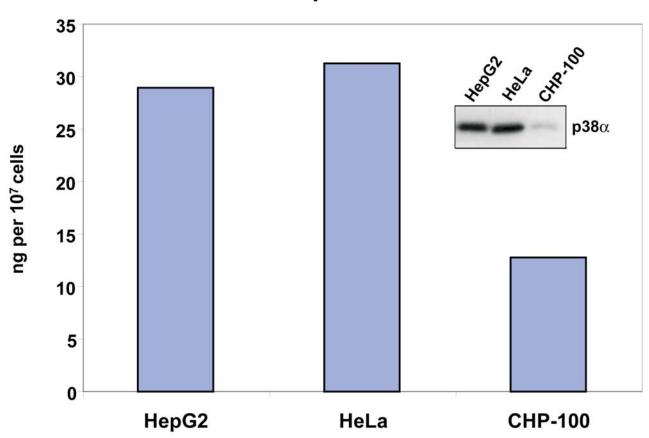


Figure 1: Lysates prepared from human HepG2, HeLa, and CHP-100 cells were quantified with this DuoSet IC ELISA. The same lysates were also immunoblotted (inset) with anti-p38 α polyclonal antibody (R&D Systems, Catalog # AF8691). The DuoSet IC ELISA results correlate well with the relative amounts of p38 α detected by Western blot.

Quantification of p38lpha in mouse and rat cell lines

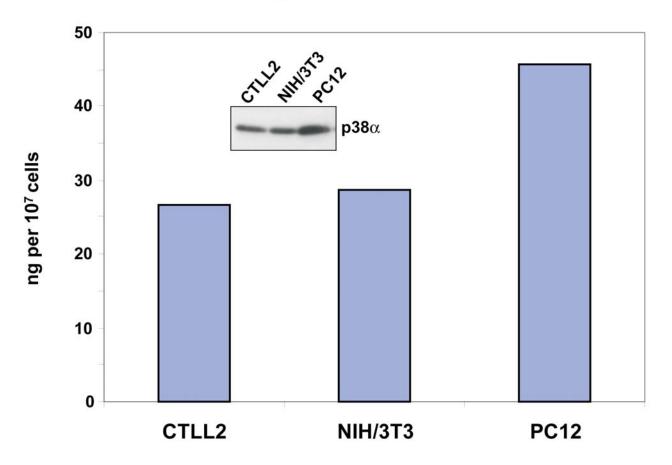
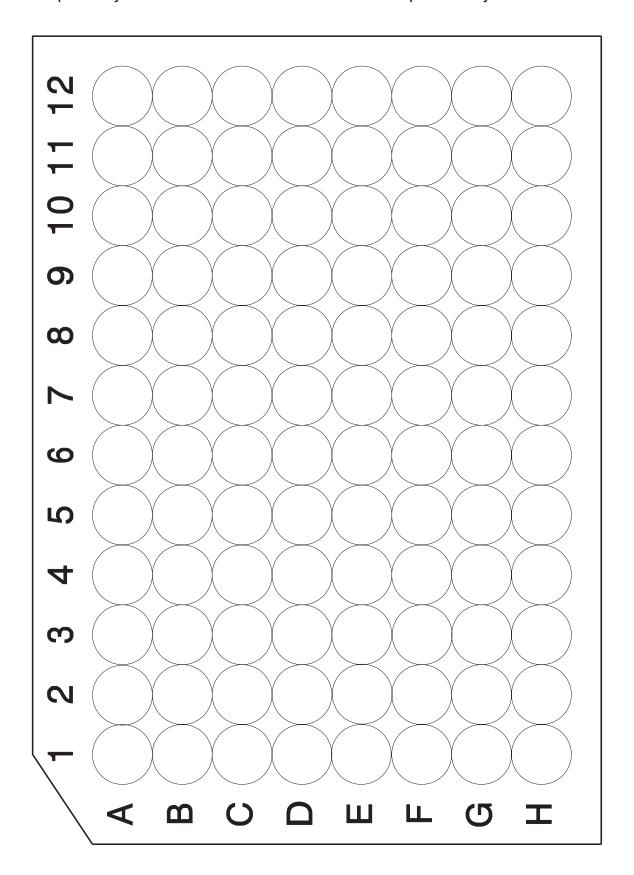


Figure 2: Lysates prepared from mouse CTLL2, mouse NIH/3T3, and rat PC12 cells were quantified with this DuoSet IC ELISA. The same lysates were also immunoblotted (inset) with anti-p38 α polyclonal antibody. The DuoSet IC ELISA results correlate well with the relative amounts of p38 α detected by Western blot.

PLATE LAYOUT

Use this plate layout as a record of standards and samples assayed.



NOTES