

# DuoSet<sup>®</sup> IC

## Human/Mouse/Rat Total GAPDH

Catalog Number DYC5718-2

DYC5718-5

DYC5718E

For the development of sandwich ELISAs to measure GAPDH in cell lysates.

This package insert must be read in its entirety before using this product.  
For research use only. Not for use in diagnostic procedures.

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## PRINCIPLE OF THE ASSAY

This DuoSet IC<sup>®</sup> ELISA contains the basic components required for the development of sandwich ELISAs to measure GAPDH in cell lysates. An immobilized capture antibody specific for human/mouse/rat GAPDH binds both phosphorylated and unphosphorylated protein. After washing away unbound material, a biotinylated detection antibody specific for human/mouse/rat GAPDH is used to detect captured protein, utilizing a standard Streptavidin-HRP format.

## MATERIALS PROVIDED & STORAGE CONDITIONS

Store the unopened kit at 2-8 °C. Do not use past kit expiration date.

DESCRIPTION	PART #	CATALOG # DYC5718-2	CATALOG # DYC5718-5	STORAGE OF OPENED/ RECONSTITUTED MATERIAL
Human/Mouse/Rat Total GAPDH Capture Antibody	843639	1 vial	2 vials	Store for up to 1 month at 2-8 °C or aliquot and store at ≤ -20 °C for up to 3 months in a manual defrost freezer.*
Human/Mouse Rat Total GAPDH Detection Antibody	843640	1 vial	2 vials	
Human/Mouse/Rat Total GAPDH Standard	843641	3 vials	5 vials	<b>Use within one hour of reconstitution. Use a fresh standard for each assay.</b>
Streptavidin-HRP	890803	1 vial	1 vial	Store at 2-8 °C. <b>DO NOT FREEZE.</b>

\* Provided this is within the expiration date of the kit.

DYC5718-2 contains sufficient materials to run ELISAs on at least two 96 well plates.<sup>†</sup>

DYC5718-5 contains sufficient materials to run ELISAs on at least five 96 well plates.<sup>†</sup>

This kit is also available in an Economy Pack (R&D Systems, Catalog # DYC5718E).

Economy Packs contain sufficient materials to run ELISAs on 15 microplates.<sup>†</sup> Specific vial counts of each component may vary. Please refer to the literature accompanying your order for specific vial counts.

<sup>†</sup> Provided the following conditions are met:

- The reagents are prepared as described in this package insert.
- The assay is run as described in the General ELISA Protocol on page 5.
- The recommended microplates, buffers, diluents, substrates, and solutions are used.

## OTHER MATERIALS REQUIRED

- Aprotinin (Sigma # A6279)
- Leupeptin (Sigma # L8511)
- Pepstatin (Sigma # P4265)
- Phenylmethylsulfonylfluoride (PMSF) (Sigma # P7626)
- Sodium Azide ( $\text{NaN}_3$ ) (Sigma # S2002)
- Sodium Fluoride (NaF) (Sigma # 201154)
- Sodium Orthovanadate ( $\text{Na}_3\text{VO}_4$ ) (Sigma # S6508), activated
- Sodium Pyrophosphate ( $\text{Na}_4\text{P}_2\text{O}_7$ ) (Sigma # P8010)
- Triton<sup>®</sup> X-100 (Sigma # T9284)
- Urea
- Pipettes and pipette tips
- Deionized or distilled water
- 96 well microplates (R&D Systems Catalog # DY990)
- Plate sealers (R&D Systems, Catalog # DY992)
- Squirt bottle, manifold dispenser, or automated microplate washer

## SOLUTIONS REQUIRED

**PBS** - 137 mM NaCl, 2.7 mM KCl, 8.1 mM  $\text{Na}_2\text{HPO}_4$ , 1.5 mM  $\text{KH}_2\text{PO}_4$ , pH 7.2-7.4, 0.2  $\mu\text{m}$  filtered (R&D Systems, Catalog # DY006).

**Wash Buffer** - 0.05% Tween<sup>®</sup> 20 in PBS, pH 7.2-7.4 (R&D Systems, Catalog # WA126).

**Block Buffer** - 1% BSA\*, 0.05%  $\text{NaN}_3$ , in PBS, pH 7.2-7.4.

**IC Diluent #1** - 1% BSA\* in PBS, pH 7.2-7.4, 0.2  $\mu\text{m}$  filtered. (R&D Systems, Catalog # DY995)

**IC Diluent #8\*\*** - 1 mM EDTA, 0.5% Triton X-100, 5 mM NaF in PBS, pH 7.2-7.4.

**Note:** IC Diluent #8 is also the base diluent for IC Diluent #3, IC Diluent #7, and Lysis Buffer #6. Approximately 50 mL of this diluent is required to run the assay on one plate.

**IC Diluent #3\*\*** - 1 mM EDTA, 0.5% Triton X-100, 5 mM NaF, 1 M urea in PBS, pH 7.2-7.4.

**IC Diluent #7\*\*** - 1 mM EDTA, 0.5% Triton X-100, 5 mM NaF, 6 M urea in PBS, pH 7.2-7.4.

**Lysis Buffer #6\*\*** - 1 mM EDTA, 0.5% Triton X-100, 5 mM NaF, 6 M urea, 1 mM activated sodium orthovanadate, 2.5 mM sodium pyrophosphate, 10  $\mu\text{g}/\text{mL}$  Leupeptin, 10  $\mu\text{g}/\text{mL}$  Pepstatin, 100  $\mu\text{M}$  PMSF, 3  $\mu\text{g}/\text{mL}$  Aprotinin in PBS, pH 7.2-7.4.

**Substrate Solution** - 1:1 mixture of Color Reagent A ( $\text{H}_2\text{O}_2$ ) and Color Reagent B (Tetramethylbenzidine) (R&D Systems, Catalog # DY999).

**Stop Solution** - 2 N  $\text{H}_2\text{SO}_4$  (R&D Systems, Catalog # DY994).

\*The use of R&D Systems Reagent Diluent Concentrate 2 (Catalog # DY995) or Millipore Bovine Serum Albumin, Fraction V, Protease free (Catalog # 82-045) is recommended. All buffers containing BSA must be stored at 2-8 °C.

\*\*Sample Diluent Concentrate 1 (5X) (R&D Systems, Catalog # DYC001), prepared as described in the DYC001 insert.

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## REAGENT PREPARATION

**Bring all reagents to room temperature before use.**

**Human/Mouse/Rat Total GAPDH Capture Antibody** (Part 843639) - Each vial contains 360 µg/mL of mouse anti-human GAPDH antibody when reconstituted with 200 µL of PBS.

**Human/Mouse/Rat Total GAPDH Detection Antibody** (Part 843640) - Each vial contains 36 µg/mL of biotinylated mouse anti-human GAPDH antibody when reconstituted with 1.0 mL of IC Diluent #1.

**Human/Mouse/Rat Total GAPDH Standard** (Part 843641) - Each vial contains 160 ng/mL of recombinant human GAPDH when reconstituted with 500 µL of IC Diluent #7. Immediately before use, an initial 6-fold dilution should be made in IC Diluent #8. Additional dilutions should be made in IC Diluent #3. A seven point standard curve using 2-fold serial dilutions and a high standard of 10,000 pg/mL is recommended.

**Streptavidin-HRP** (Part 890803) - Immediately before use, dilute the Streptavidin-HRP to the working concentration specified on the vial label using IC Diluent #1.

## PREPARATION OF SAMPLES

**Cell Lysates** - Rinse cells two times with PBS, making sure to remove any remaining PBS after the second rinse. Solubilize cells at  $1 \times 10^7$  cells/mL in Lysis Buffer #6 and allow samples to sit on ice for 15 minutes. Assay immediately or store at  $\leq -70$  °C. Before use, centrifuge samples at 2000 x g for 5 minutes and transfer the supernate to a clean test tube. Sample protein concentration may be quantified using a total protein assay. For assaying, dilute lysates 6-fold with IC Diluent #8 and make further serial dilutions in IC Diluent #3.

**Note:** *The final concentration of urea in all samples and standards should be 1 M prior to addition to the plate.*

## PRECAUTIONS

The Stop Solution provided with this kit is an acid solution.

Some components in this kit contain ProClin® which may cause an allergic skin reaction. Avoid breathing mist.

Wear protective gloves, clothing, eye, and face protection. Wash hands thoroughly after handling. Please refer to the MSDS on our website prior to use.

## TECHNICAL HINTS AND LIMITATIONS

- This DuoSet IC ELISA should not be used beyond the expiration date on the kit label.
- Individual results may vary due to differences in technique, plasticware, and water sources.
- It is important that the diluents selected for reconstitution and for dilution of the standard reflect the environment of the samples being measured. The diluent suggested in this protocol should be suitable for most cell lysates.
- The type of enzyme and substrate and the concentrations of capture/detection antibodies used can be varied to create an immunoassay with a different sensitivity and dynamic range. A basic understanding of immunoassay development is required for the successful use of these reagents in immunoassays.
- A thorough and consistent wash technique is essential for proper assay performance. Wash Buffer should be dispensed forcefully and removed completely from the wells by aspiration or decanting. Remove any remaining Wash Buffer by inverting the plate and blotting it against clean paper towels.
- Use a fresh reagent reservoir and pipette tips for each step.
- It is recommended that all standards and samples be assayed in duplicate.
- Avoid microbial contamination of reagents and buffers. This may interfere with the sensitivity of the assay. Buffers containing protein should be made under aseptic conditions and stored at 2-8 °C or be prepared fresh daily.

## GENERAL ELISA PROTOCOL

### Plate Preparation

1. Dilute the Capture Antibody to a working concentration of 2.0 µg/mL in PBS, without carrier protein. Immediately coat a 96 well microplate with 100 µL per well of the diluted Capture Antibody. Seal the plate and incubate overnight at room temperature.
2. Aspirate each well and wash with Wash Buffer, repeating the process two times for a total of 3 washes. Wash by filling each well with Wash Buffer (400 µL) using a squirt bottle, manifold dispenser, or autowasher. Complete removal of liquid at each step is essential for good performance. After the last wash, remove any remaining Wash Buffer by aspirating or by inverting the plate and blotting it against clean paper towels.
3. Block plates by adding 300 µL of Block Buffer to each well. Incubate at room temperature for 1-2 hours.
4. Repeat the aspiration/wash as in step 2. The plates are now ready for sample addition.

### Assay Procedure

1. Add 100 µL of sample or standards in IC Diluent #3 per well. Use IC Diluent #3 as the zero standard. Cover with a plate sealer and incubate 2 hours at room temperature.  
**Note:** *A seven point standard curve using 2-fold serial dilutions and a high standard of 10,000 pg/mL is recommended.*
2. Repeat the aspiration/wash as in step 2 of Plate Preparation.
3. Dilute the Detection Antibody to a working concentration of 1.0 µg/mL in IC Diluent #1. Add 100 µL of the diluted Detection Antibody to each well. Cover with a new plate sealer and incubate 2 hours at room temperature.
4. Repeat the aspiration/wash as in step 2 of Plate Preparation.
5. Immediately before use, dilute the Streptavidin-HRP to the working concentration specified on the vial label using IC Diluent #1. Add 100 µL of the diluted Streptavidin-HRP to each well. Incubate for 20 minutes at room temperature. Avoid placing the plate in direct light.
6. Repeat the aspiration/wash as in step 2 of the Plate Preparation.
7. Add 100 µL of Substrate Solution to each well. Incubate for 20 minutes at room temperature. Avoid placing the plate in direct light.
8. Add 50 µL of Stop Solution to each well. Gently tap the plate to ensure thorough mixing.
9. Determine the optical density of each well immediately, using a microplate reader set to 450 nm. If wavelength correction is available, set to 540 nm or 570 nm. If wavelength correction is not available, subtract readings at 540 nm or 570 nm from the readings at 450 nm. This subtraction will correct for optical imperfections in the plate. Readings made directly at 450 nm without correction may be higher and less accurate.

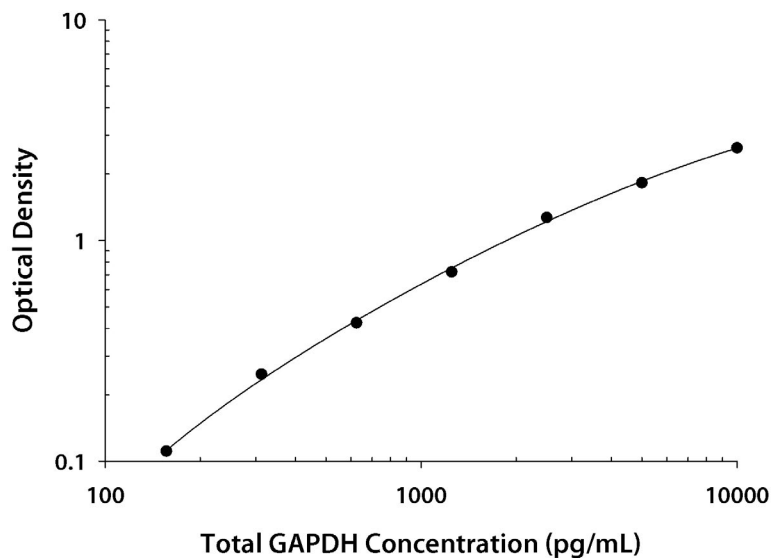
## CALCULATION OF RESULTS

Average the duplicate readings for each standard and sample, then subtract the average zero standard optical density (O.D.). Results may be normalized to total protein or cell number.

Create a standard curve by reducing the data using computer software capable of generating a four parameter logistic (4-PL) curve-fit. As an alternative, construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph. The data may be linearized by plotting the log of the human/mouse/rat GAPDH concentrations versus the log of the O.D. and the best fit line can be determined by regression analysis. This procedure will produce an adequate but less precise fit of the data.

## TYPICAL DATA

A standard curve should be generated for each set of samples assayed. The graph below represents typical data generated when using this Human/Mouse/Rat Total GAPDH DuoSet IC ELISA. The standard curve was calculated using a computer generated 4-PL curve-fit. This standard curve is for demonstration purposes only.



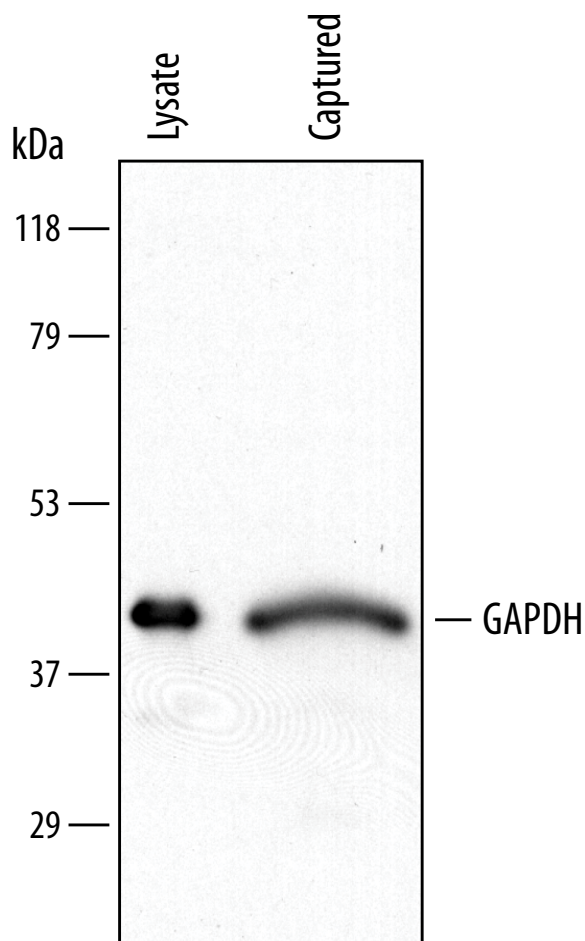
## CALIBRATION

The Human/Mouse/Rat Total GAPDH DuoSet ELISA is calibrated against a highly purified *E. coli*-expressed recombinant human GAPDH produced at R&D Systems. Samples containing natural GAPDH showed linear dilution parallel to the standard curve obtained using the Human/Mouse/Rat Total GAPDH Standard. These results indicate that O.D. values from this DuoSet IC ELISA can be used to determine the relative concentrations of GAPDH in natural samples.



## SPECIFICITY

The Human/Mouse/Rat Total GAPDH DuoSet IC ELISA specifically recognizes GAPDH. Specificity was demonstrated by Western blot analysis of the protein bound by the capture antibody supplied in the kit.

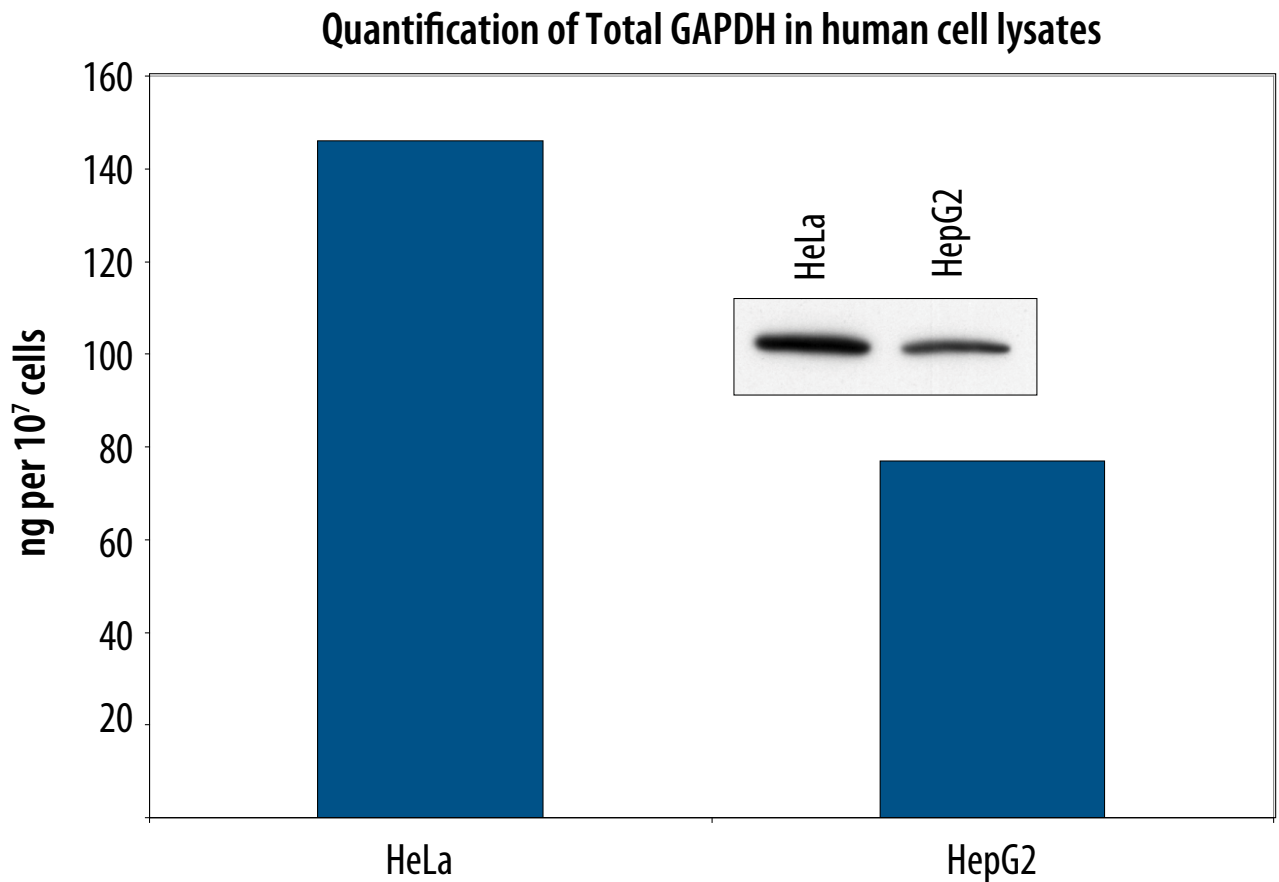


**Figure 1:** Lysate prepared from HeLa human cervical epithelial carcinoma cells was incubated in wells coated with Human/Mouse/Rat Total GAPDH Capture Antibody. Unbound material was removed by washing and bound material was solubilized in SDS gel sample buffer. The same lysate and captured proteins were resolved by SDS-PAGE, transferred to a PVDF membrane, and immunoblotted with Human/Mouse/Rat Total GAPDH Detection Antibody. Only a single band corresponding to GAPDH was detected in captured material, indicating that the ELISA is specific for GAPDH.

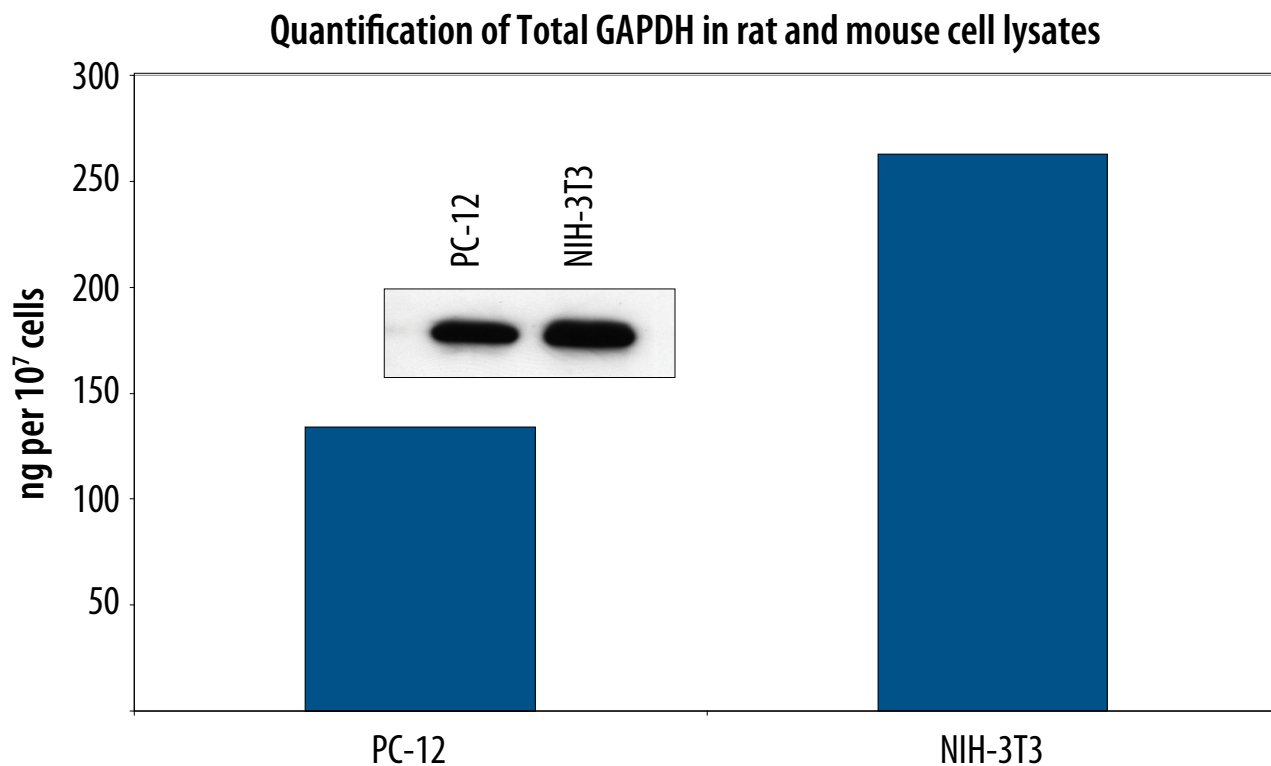
To further determine specificity, recombinant human GAPDH-2 was assayed at 100 ng/mL and did not cross-react or interfere in the assay.

## QUANTIFICATION

Amounts of GAPDH, as quantified by the Human/Mouse/Rat Total GAPDH DuoSet IC ELISA, are consistent with the relative amounts of GAPDH determined by qualitative Western blot analysis.



**Figure 2:** Lysates prepared from HeLa human cervical epithelial carcinoma cells and HepG2 human hepatocellular carcinoma cells were quantified with this DuoSet IC ELISA. The same lysates were also immunoblotted (inset) with anti-GAPDH monoclonal antibody (R&D Systems, Catalog #MAB5718). The DuoSet IC ELISA results correlate well with the relative amounts of GAPDH detected by Western blot.



**Figure 3:** Lysates prepared from PC-12 rat adrenal pheochromocytoma cells and NIH-3T3 mouse embryonic fibroblast cells were quantified with this DuoSet IC ELISA. The same lysates were also immunoblotted (inset) with anti-GAPDH polyclonal antibody (R&D Systems, Catalog #AF5718). The DuoSet IC ELISA results correlated well with the relative amounts of GAPDH detected by Western blot.

## PLATE LAYOUT

Use this plate layout to record standards and samples assayed.

A diagram of a 12x8 plate layout. The rows are numbered 1 through 12 on the left side, and the columns are labeled A through H at the bottom. The plate is represented as a grid of 96 circular wells, arranged in 12 rows and 8 columns. The top row is labeled 12, the second row 11, and so on, down to row 1. The columns are labeled A, B, C, D, E, F, G, and H from left to right. The plate has a notch at the top and a tab at the bottom.

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