

Biotinylated Recombinant SARS-CoV-2 B.1.617.2 (G446V) Spike RBD His-tag Avi-

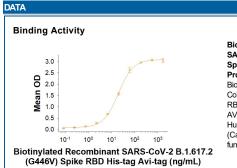
tag

Catalog Number: AVI10982

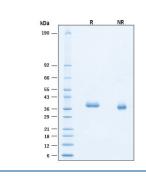
DESCRIPTION				
Source	Human embryonic kidney cell, HEK293-derived sars-cov-2 Spike RBD protein			
	Human embryonic kidney cell, HEK293- derived sars-cov-2 Spike RBD protein Arg319-Phe541 (Gly446Val, Leu452Arg, Thr478Lys) Accession # YP_009724390.1	6-His tag	Avi-tag	
	N-terminus C-tr			
N-terminal Sequence Analysis	Arg319			
Structure / Form	Biotinylated via Avi-tag.			
Predicted Molecular Mass	26 kDa			

SPECIFICATIONS		
SDS-PAGE	34-41 kDa, under reducing conditions.	
Activity	Measured by its binding ability in a functional ELISA with Recombinant Human ACE-2 Fc Chimera (Catalog # 10544-ZN).	
Endotoxin Level	<0.10 EU per 1 µg of the protein by the LAL method.	
Purity	>95%, by SDS-PAGE visualized with Silver Staining and quantitative densitometry by Coomassie® Blue Staining.	
Formulation	Lyophilized from a 0.2 µm filtered solution in PBS with Trehalose. See Certificate of Analysis for details.	

PREPARATION AND STORAGE		
Reconstitution	Reconstitute at 500 µg/mL in PBS.	
Shipping	The product is shipped at ambient temperature. Upon receipt, store it immediately at the temperature recommended below.	
Stability & Storage	 Use a manual defrost freezer and avoid repeated freeze-thaw cycles. 12 months from date of receipt, -20 to -70 °C as supplied. 1 month, 2 to 8 °C under sterile conditions after reconstitution. 3 months, -20 to -70 °C under sterile conditions after reconstitution. 	



Biotinylated Recombinant SARS-CoV-2 B.1.617.2 (G446V) Spike RBD His-tag Avi-tag Protein Binding Activity. Biotinylated Recombinant SARS-CoV-2 B.1.617.2 (G446V) Spike RBD His-tag Avi-tag (Catalog # AV/10982) binds Recombinant Human ACE-2 Fc Chimera (Catalog # 10544-ZN) in a functional ELISA. SDS-PAGE



Biotinylated Recombinant SARS-CoV-2 B.1.617.2 (G446V) Spike RBD His-tag Avitag Protein SDS-PAGE. 2 µg/lane of Biotinylated Recombinant SARS-CoV-2 B.1.617.2 (G446V) Spike RBD His-tag Avitag Protein (Catalog # AVI10982) was resolved with SDS-PAGE under reducing (R) and non-reducing (NR) conditions and visualized by Coomassie® Blue staining, showing bands at 34-41 kDa.

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tag

Catalog Number: AVI10982

BACKGROUND

SARS-CoV-2, which causes the global pandemic coronavirus disease 2019 (Covid-19), belongs to a family of viruses known as coronaviruses that also include MERS-CoV and SARS-CoV-1. Coronaviruses are commonly comprised of four structural proteins: Spike protein (S), Envelope protein (E), Membrane protein (M) and Nucleocapsid protein (N) (1). The SARS-CoV-2 S protein is a glycoprotein that mediates membrane fusion and viral entry. The S protein is homotrimeric, with each ~180-kDa monomer consisting of two subunits, S1 and S2 (2). In SARS-CoV-2, as with most coronaviruses, proteolytic cleavage of the S protein into S1 and S2 subunits is required for activation. The S1 subunit is focused on attachment of the protein to the host receptor while the S2 subunit is involved with cell fusion (3-5). A receptor binding domain (RBD) in the C-terminus of the S1 subunit has been identified and the RBD of SARS-CoV-2 shares 73% amino acid (aa) identity with the RBD of the SARS-CoV-1, but only 22% aa identity with the RBD of MERS-CoV (6, 7). The low aa sequence homology is consistent with the finding that SARS and MERS-CoV bind different cellular receptors (8). The RBD of SARS-CoV-2 binds a metallopeptidase, angiotensin-converting enzyme 2 (ACE-2), similar to SARS-CoV-1, but with much higher affinity and faster binding kinetics (9). Before binding to the ACE-2 receptor, structural analysis of the S1 trimer shows that only one of the three RBD domains is in the "up" conformation. This is an unstable and transient state that passes between trimeric subunits but is nevertheless an exposed state to be targeted for neutralizing antibody therapy (10). Polyclonal antibodies to the RBD of the SARS-CoV-2 protein have been shown to inhibit interaction with the ACE-2 receptor, confirming RBD as an attractive target for vaccinations or antiviral therapy (11). There is also promising work showing that the RBD may be used to detect presence of neutralizing antibodies present in a patient's bloodstream, consistent with developed immunity after exposure to the SARS-CoV-2 (12). Several emerging SARS-CoV-2 genomes have been identified with mutations in the RBD compared to the Wuhan-Hu-1 SARS-CoV-2 reference sequence. First detected in India in December 2020, the B.1.617.2, or Delta variant, is considered a Variant of Concern (VOC) as it contains several mutations in the RBD domain that potentially affect viral fitness and transmissibility: L452R and T478K. The L452R mutation is known to increase affinity for ACE-2 receptors and is associated with resistance to neutralization by multiple monoclonal antibodies (13, 14). The T478K shows significant increase in ACE-2 binding affinity and may make the variant more transmissible and infectious (15). This construct also contains the G446V mutation which is predicted to be highly antigenic and described as affecting neutralization by antibodies present in polyclonal serum (13). Additionally, vaccines developed against SARS-CoV-2 show a decrease in efficacy towards the Delta variant (16). Our Avi-tag Biotinylated SARS-CoV-2 B.1.617.2/G446V RBD features biotinylation at a single site contained within the Avi-tag, a unique 15 amino acid peptide. Protein orientation will be uniform when bound to streptavidin-coated surface due to the precise control of biotinylation and the rest of the protein is unchanged so there is no interference in the protein's bioactivity

References:

- 1. Wu, F. et al. (2020) Nature 579:265.
- 2. Tortorici, M.A. and D. Veesler (2019) Adv. Virus Res. 105:93.
- 3. Bosch, B.J. et al. (2003). J. Virol. 77:8801.
- 4. Belouzard, S. et al. (2009) Proc. Natl. Acad. Sci. 106:5871.
- 5. Millet, J.K. and G.R. Whittaker (2015) Virus Res. 202:120.
- 6. Li, W. *et al*. (2003) Nature **426**:450.
- 7. Wong, S.K. et al. (2004) J. Biol. Chem. 279:3197.
- 8. Jiang, S. et al. (2020) Trends. Immunol. https://doi.org/10.1016/j.it.2020.03.007.
- 9. Ortega, J.T. et al. (2020) EXCLI J. 19:410.
- 10. Wrapp, D. et al. (2020) Science 367:1260.
- 11. Tai, W. et al. (2020) Cell. Mol. Immunol. https://doi.org/10.1016/j.it.2020.03.007.1.
- 12. Okba, N.M.A. et al. (2020). Emerg. Infect. Dis. https://doi.org/10.3201/eid2607.200841.
- 13. Liu, Z. et al. (2021) Cell Host Microbe. 29:477.
- 14. Motozono C, et al. (2021) bioRxiv, https://www.biorxiv.org/content/10.1101/2021.04.02.438288v1.
- 15. Wang, R. et al. (2021) Genomics 113:2158.
- 16. Pouwels, K.B. et al. (2021) Preprint at Univ. Oxford https://www.ndm.ox.ac.uk/files/coronavirus/covid-19-infection-survey/finalfinalcombinedve20210816.pdf.

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