

# **EDC-HCI Protocol and Product Information Sheet**

Zero-Length Crosslinkers
<u>c1100-3x10mg, c1100-100mg, c1100-5gm, c1100-25gm, c1100-100gm</u>
EDC-HCI
EDC; EDAC; EDAC-HCI; 1-(3-Dimethylaminopropyl)-3-ethylcarbodiimide HCI
25952-53-8
C <sub>8</sub> H <sub>17</sub> N <sub>3</sub> HCI
191.70
N/A
-20°C or below (shipped at ambient temperature).

N<sup>-C-N</sup>

## **Background Information**

Zero-length crosslinking allows scientists to immobilize protein-protein interactions without introducing a spacer arm. The EDC-HCI / Sulfo-NHS system has been perhaps the most successful way of creating zero-length crosslinks for decades. Facilitated through a reactive carbodiimide (EDC-HCI) and Sulfo-NHS as a catalyst, this coupling procedure is a highly efficient choice for crosslinking proteins or immobilizing proteins to a support. This procedure is designed to assist the scientist in crosslinking proteins or creating protein:protein conjugates. Immobilization techniques and other uses for this set of reagents can be obtained by contacting ProteoChem's technical support.

### **General EDC-HCI Protein Crosslinking Protocol**

Important Notes:

- EDC-HCl is moisture sensitive. Allow EDC-HCl to equilibrate to room temperature before opening vial.
- Except as indicated in this protocol, avoid using buffers that contain DTT, EDTA, or β-Mercaptoethanol, as these can interfere with conjugation reaction.
- It is recommended that the incubation steps of this procedure be done using a rotary stirring device, although intermittent light vortexing is also acceptable.
- When choosing Protein #1 (P<sub>1</sub>) and Protein #2 (P<sub>2</sub>), ensure that P<sub>1</sub> has free carboxyl groups available (COOH) and P<sub>2</sub> has free amine groups available (NH<sub>2</sub>).
  - P1: Terminal COOH, Asp, and Glu
  - P2: Terminal NH2, Lys

Reagents and Buffers Needed

Conjugation Buffer: 100 mM MES (<u>cr8107-25gm</u>), 500 mM NaCl, pH 6.0 *(Abr. MES* = 2(*N*-morpholino)ethanesulfonic acid) β-Mercaptoethanol Sulfo-NHS (<u>c1102-500mg</u>) Hydroxylamine-HCl (<u>cr8108-25gm</u>) Desalting Column(s): Sephadex® G-25 (<u>g4109</u>) or equivalent

Step 1. Activation of Protein #1



- a. Dissolve Protein #1 (P<sub>1</sub>) at 1-2 mg/mL in 1.0 mL Conjugation Buffer Record P<sub>1</sub> Concentration: (mg P<sub>1</sub>) / [(MW protein in mg/mmol) \* (mL Buffer)] = mM P<sub>1</sub>
- b. Weigh out 0.8 mg EDC-HCl and 2.2 mg of Sulfo-NHS and add directly to the protein solution in Step 1.a. This will give 4 mM EDC-HCl and 10 mM Sulfo-NHS.
- c. Gently vortex reaction mixture until all reagent is soluble.
- d. Allow Activation Reaction to proceed at room temperature for 15 minutes.
- e. Add 1.4  $\mu$ L of  $\beta$ -Mercaptoethanol to deactivate excess EDC-HCI. Note: If either protein has critical disulfide bonds, this deactivation step can be omitted. In such a case, proceed directly to Step 2, or desalt activated **P**<sub>1</sub> through gel filtration, then proceed to Step 2.

### Step 2. Conjugation of Two Proteins

- a. Dissolve Protein #2 ( $P_2$ ) in conjugation buffer at a concentration of 1-2 mg/mL. Ideally, this should be at the same concentration as  $P_1$ . Record  $P_2$  Concentration:
  - $(mg P_2) / [(MW protein in mg/mmol) * (mL Buffer)] = \_____ mM P_2$
- b. Gently vortex and allow the crosslinking reaction to proceed for 1.5 to 3 hours at room temperature. Use of rotary mixer is preferred.
- c. Quench the conjugation reaction by adding hydroxylamine (to give a final concentration of 5 to 10 mM).
- d. Purify the obtained conjugate from excess reagents and reaction by-products by gel filtration or dialysis. Exchange sample into buffer of choice. Conjugate bonds are stable under most biological conditions.

### References:

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