

# JB-4 Embedding Kit<sup>®</sup>

Catalog #00226

## Introduction

JB-4 Embedding Kit is a unique polymer embedding material that gives a higher level of morphological detail than paraffin processed tissues. A water-soluble media, JB-4 does not require dehydration to absolute alcohol except for dense, bloody, or fatty tissue specimens. JB-4 is excellent for small non-decalcified bone specimens, routine stains, special stains, and histochemical staining. *Clearing agents such as xylene and chloroform are not required.* The polymerization of JB-4 is exothermic, which is easily controlled by polymerizing on ice or by using refrigeration at 4°C. JB-4 Embedding Kits must be used under a chemical fume hood.

Sections of JB-4 embedded material can be cut at 0.5 to 3.0 microns or thicker. Microtomes designed for plastic sectioning are required as are glass, Ralph, or tungsten carbide knives. Immunohistochemical procedures are not recommended for JB-4 as the glycol methacrylate cannot be removed from the section and may block antigen sites for most antibody reactions. As an alternative we recommend the Polysciences, Inc. Osteo-Bed Bone Embedding Kit (Catalog #17734). The Osteo-Bed formulation is a methyl methacrylate that is well suited for bone or soft tissue for immunohistochemistry.

## NOTE

It is recommended that the Embedding Kit be used under a fume hood with appropriate gloves. For additional details, see **Warnings** and **Precautions**.

## Fixation

Specimens can be fixed in 10% Neutral Buffered Formalin or other routine histological fixative. We suggest using Poly/LEM, a methanol free formalin based fixative for light and electron microscopy. Routine specimen sizes for soft tissue should be no more than 2.0cm X 2.0cm X 2.0cm with fixation at a minimum of four hours to overnight. Fatty or dense tissues should be fixed overnight. Larger bone specimens will require fixation overnight or longer depending on the specimen size. Fixation can be at room temperature or 4°C. Cold fixation will extend the time required for the specimen to be penetrated and fixed. Decalcification is not required for JB-4 embedded specimens.

## Dehydration

Dehydration can be completed at room temperature or 4°C. This process can also be done with a routine tissue processor, stopped at the end of the last alcohol step and removed for infiltration. Please note that polymers cannot be used in routine histology tissue processors at any time. It may void the warranty and possibly begin to polymerize in the system thereby blocking the lines. Check with the manufacturer prior to attempting infiltration on any unit.

## Infiltration

*Infiltration Solution Mixing Procedure:*

The following amounts of material are used for one 100ml batch of Infiltration Solution:

JB-4 Solution A (Monomer)	100.00ml
Benzoyl Peroxide, Plasticized (Catalyst)	1.25gm

Carefully weigh 1.25gm of catalyst (benzoyl peroxide, plasticized) and add to 100.00ml Solution A while stirring on a magnetic stirrer. Mix until dissolved, approximately 10 to 20 minutes. Measurement of the catalyst is critical because it controls the rate of polymerization of the plastic and the exothermic reaction. This infiltration solution can be stored for up to two weeks in a dark cool area or in the refrigerator at 4°C.

## Infiltration Procedure

Infiltration is performed at room temperature or 4°C. Do not expose the samples to heat or direct light during infiltration. The specimens should be placed in two to three changes of Infiltration Solution to allow for the removal or replacement of all alcohols or tissue fluids.

Dehydration and infiltration can be completed simultaneously by using absolute alcohol in ascending grades of infiltration solution starting at 50% ethanol and 50% infiltration solution followed by 25% ethanol; 75% infiltration solution then 10% ethanol to 90% infiltration solution and finally in infiltration solution for a minimum of 3 changes prior to embedding. Proceed to the embedding steps.

The amount of infiltration solution used is approximately 8 to 10 times that of the volume of the specimen. The changes of fluid should occur every 10 to 90 minutes for smaller specimens. The time in each change is dependent on the size of the specimen. When infiltration is complete the tissue generally appears translucent and in most cases will sink to the bottom of the container. Infiltration should be done on a slow rotator, hematology shaker table or inverted several times during the process to allow complete saturation.

## Embedding

**The polymerization process should be under anaerobic conditions with the use of block holders, under light vacuum or in an air-tight container.**

Prior to mixing the Embedding Solution, collect and prepare the following materials; embedding molds, block holders, labels, gloves, instruments, an ice bath, and the specimens. Do not pre-cool the molds as this may cause condensation and prevent even polymerization of the block face. To prevent polymerization from occurring too fast and possible overheating of the tissue it is recommended that the polymerization process for embedding be slowed by completing it in the refrigerator or in a cold room at 4°C. Note that this may extend the polymerization from several hours to overnight.

Larger specimens with increased embedding solution may have an even greater exothermic reaction. This should be controlled by using a 4°C refrigerator or cold room. These larger specimens will require longer times for complete polymerization and may have more unpolymerized liquid on top of the block. This will appear as a syrup and should be allowed to drain off the blocks by inverting the molds upside down and then wiping off excess. It is recommended that gloves be worn to reduce exposure to un-polymerized resin.

## Embedding Solution Mixing Procedure

***Make fresh Solution A following the directions in Infiltration Solution and Procedures above. Do not use old or used catalyzed Infiltration Solution for the embedding solutions.***

The following amounts of material are used for 25ml of embedding solution:

Infiltration Solution            25.0ml

JB-4 Solution B (Accelerator) 1.0ml (Must be an exact measurement.)

Mix 25ml of freshly made Infiltration Solution and 1.0ml of JB-4 Solution B thoroughly and begin embedding immediately. The small JB-4 Embedding Molds from Polysciences, Inc. require approximately 1.5 to 2ml of solution per mold. The Block Holder is essential to exclude oxygen during the polymerization process. If Block Holders are not used, cover the molds with an air tight film or place under vacuum at no more than 15psi, preferably in a cold room at 4°C or a refrigerator. If anaerobic conditions are not maintained, the JB-4 may polymerize incompletely or not all. BEEM® capsules may be capped for embedding.

We recommend polymerization in the refrigerator at 4°C or on an ice bath to reduce the exothermic reaction to 55°C or less. Room temperature polymerization will be complete in 1 to 2 hours for smaller blocks and can go up to three hours or more for very large blocks. Note that the exothermic reaction can exceed 100°C for larger specimens using 10 to 50ml of embedding solution at room temperature, therefore large blocks should be polymerized in the refrigerator or on ice. The blocks may range in color from light yellow to dark yellow or amber. This color shift is not a problem and will not effect the block hardness. The top of the block may have a liquid film on it that can be removed by draining or drying the block in a desiccator for several hours to overnight.

## Deplasticizing and Staining

JB-4 is a glycol methacrylate based polymer and cannot be removed from sections. Therefore, no organic solvents are required. Routine histology stains and most histochemistry can be run on the sections. High molecular weight special stains or immunohistochemical reactions may not penetrate the polymerized plastic in the sections.

## Warning

May be harmful if swallowed. Use under a hood with appropriate gloves. Components may cause irritation and or allergic skin reaction. Avoid contact with eyes, skin and clothing. Avoid inhalation of the vapors. Wash hands or exposed areas thoroughly after handling the solutions.

## Precautions

Do not heat over an open flame. Avoid electrical or static sparks. Store un-catalyzed resin in the original containers at room temperature in a dark cool area.

## First Aid

In case of contact with any component or mixed solution immediately flush area with water for at least 15 minutes. Should either unpolymerized or polymerized material contact the eyes flush with water for at least 15 minutes. If swallowed drink water to excess and call a physician immediately. Never give anything by mouth to someone who is unconscious.

## Storage

Refrigeration of the kit components is not required but they do require storage in a cool dark place. Do not store in the light or in a heated area as it may cause the monomer to polymerize. The catalyst, plasticized benzoyl peroxide, is an organic peroxide that is shipped dry and does not require special storage. Please note that the catalyst is formulated to remain stable and weigh correctly for this procedure without any adjustments to the amounts recommended. The catalyst should be kept tightly sealed. The catalyst may decompose with age, therefore we recommend carefully monitoring the date received and using the catalyst only with the kit it came in for best results.

Should any of our materials fail to perform to our specifications, we will be pleased to provide replacements or return the purchase price. We solicit your inquiries concerning all needs for life sciences work. The information given in this bulletin is to the best of our knowledge accurate, but no warranty is expressed or implied. It is the user's responsibility to determine the suitability for his own use of the products described herein, and since conditions of use are beyond our control, we disclaim all liability with respect to the use of any material supplied by us. Nothing contained herein shall be construed as a recommendation to use any product or to practice any process in violation of any law or any government regulation

## Catalyst Disposal Procedure

The catalyst can be destroyed by slowly adding and mixing it in small portions of the catalyst at 4 times or more the volume to weight of 10% sodium hydroxide solution in water. Do not allow material to settle in lumps or stand in layers and mix until dissolved completely. Dispose of this solution, Monomer A and the accelerator with other hazardous wastes in accordance with local, state, and federal regulations.

## Ordering Information

Catalog #	Description	Size
00226-1	JB-4 Embedding Kit	1 Kit
0226A-800	JB-4 Solution A	800 ml
0226B-3.8	JB-4 Solution B	3.8 L
02618-12	Catalyst (Benzoyl Peroxide, Plasticized)	12.0gm
16864-3.75	Poly/LEM Fixative	3.75 L/4x3.75 L
16643A-1	Polyethylene Molding Cup Trays 6 X 12 X 5mm (20 Cavities)	1 Each
16643B-1	Polyethylene Molding Cup Trays 12 X 16 X 5mm (20 Cavities)	1 Each
17177A-3	Polyethylene Molding Cup Trays 6 X 8 X 5mm (9 Cavities) Hex	3 Each
17177B-3	Polyethylene Molding Cup Trays 2 X 15 X 5mm (9 Cavities)	3 Each
17177C-3	Polyethylene Molding Cup Trays 13 X 19 X 5mm (9 Cavities)	3 Each
15899-50	JB-4 Plastic Block Holders	50/Pkg
24216-1	Tissue Tack Slides Silane Coated	Approx. 72/Box
24234-1	Tungsten Carbide Disposable Blades	2 per pkg

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### To Order In The US:

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Fax: 1-800-343-3291 • (215) 343-0214

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