

Enzyme Immunoassay for Recombinant (Therapeutic) Human IgG1

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INTRODUCTION

Recombinant human monoclonal antibodies have become a major focus of new drug discovery in the Pharmaceutical and Biotech industries, with hundreds of new human monoclonal antibodies currently in preclinical development. Fully human recombinant antibodies have gradually replaced humanized mouse or rat antibodies, with IgG1 and IgG4 as the preferred engineered isotypes². As part of the developmental process, human therapeutic antibody drug candidate molecules must be tested in non-human primate species to assess their pharmacokinetic profile and potential toxicity³. Primate and human antibodies are highly homologous, making it difficult to distinguish the human antibody drug candidate from the endogenous primate antibodies in a serum or plasma sample by traditional human IgG-specific immunoassays. Therefore, we developed two highly sensitive ELISAs that distinguish human IgG1 or human IgG4 from rhesus or cynomolgus monkey immunoglobulins. This Recombinant (Therapeutic) Human IgG1 ELISA can be used to accurately quantify human IgG1 in the serum or plasma of rhesus or cynomolgus monkeys without cross reacting with antibodies of those species. This same ELISA can also be used to quantify human IgG1 in the plasma of other preclinical test species (mouse, rat, rabbit, guinea pig, etc.) if desired. This assay is supplied with a recombinant human IgG1 kappa allotype G1m17.1 standard. The customer may wish to substitute their own specific drug candidate IgG1 for this standard in the assay.

PRINCIPLES OF PROCEDURE

This is a sandwich enzyme-linked immunosorbent assay (ELISA). The plate is pre-coated with anti-Human IgG Antibody and blocked, ready for the addition of samples and standards. The assay should take approximately 3 hours to run, plus any required sample preparation time.

MATERIALS PROVIDED

Component	Description	Volume	Storage	Cat no.
Anti-Human IgG Plate	96-well plate coated and blocked	1 plate	4°C	NF01a
Assay Buffer	Buffer to dilute kit components and samples	100mL	4°C	NF01b
10x Wash Buffer	Buffer used to wash the plate	30mL	4°C	NF01c
Human IgG1 Standard	Lyophilized Recombinant Human IgG1	0.2µg	4°C	NF01d
10x Detection Antibody	Anti-Human IgG1-HRP Conjugate	1.5mL	4°C	NF01e
TMB Substrate	Stabilized TMB color reagent	15mL	4°C	NF01f

MATERIALS NEEDED BUT NOT PROVIDED

1. Microplate reader with 450 nm filter
2. Adjustable micropipettes (10 – 1000 µL) and tips
3. 3N Sulfuric Acid (H₂SO₄)
4. Plate shaker

5. Plate cover or plastic film

STORAGE

1. Store the components of this kit at the temperatures specified on the labels.
2. Unopened reagents are stable until the indicated kit expiration date.
3. Reconstituted Standard may be used up to 1 month after reconstitution if stored at 4°C
4. Diluted Detection Antibody must be used within 2 hours of dilution

WARNINGS AND PRECAUTIONS

1. Use aseptic technique when opening and dispensing reagents.
2. This kit is designed to work properly as provided and instructed. Additions, deletions or substitutions to the procedure or reagents are not recommended, as they may be detrimental to the assay.

PROCEDURAL NOTES

1. To minimize errors in absorbance measurements due to handling, wipe the exterior bottom of the microplate wells with a lint-free paper towel prior to inserting into the plate reader.

SAMPLE PREPARATION

If assessing human IgG1 in monkey plasma or serum, dilute the serum at least 1:5 into Assay Buffer before testing. Undiluted monkey plasma or serum contains contaminants that interfere with the accurate functioning of this ELISA.

REAGENT PREPARATION

1. **Human IgG1 Standard:** Resuspend lyophilized standard vial by adding 0.2mL of Assay Buffer directly to vial, gently invert to suspend completely, to create 1µg/mL stock solution.
2. **10x Wash Buffer:** Dilute the wash buffer 1:10 by adding 30mL of 10x Wash Buffer to 270mL of dH₂O.
3. **10x Detection Antibody:** Immediately prior to use, dilute detection antibody 1:10 by adding 1.2mL 10x Detection Antibody to 10.8mL Assay Buffer. Use within 2 hours of diluting.

STANDARD CURVE PREPARATION

Set up for the standard curve by labeling dilution tubes and dispensing the indicated volumes of Assay Buffer and 1µg/mL Standard Stock Solution according to Table 1 below.

Table 1: Standard Curve Preparation

Standard	IgG1 Concentration (ng/mL)	Assay Buffer (µL)	Transfer Volume (µL)	Transfer Source	Final Volume (µL)
S ₇	20	980	20	1µg/mL Stock	500
S ₆	10	500	500	S ₇	500
S ₅	5	500	500	S ₆	500
S ₄	2.5	500	500	S ₅	500
S ₃	1.25	500	500	S ₄	500
S ₂	0.63	500	500	S ₃	500
S ₁	0.31	500	500	S ₂	1000
S ₀	0	1000	-	-	1000

ASSAY PROCEDURE

1. Add 100 μL of Standards and Samples to the corresponding wells on the microplate in duplicate. Incubate at room temperature for 1.5 hours on an orbital plate shaker. See Scheme 1 below for a suggested plate layout.
2. Dump the contents of the plate and wash each well three times with 300 μL of Wash Buffer. After the final wash, tap the plate upside-down on a lint-free paper towel to make sure there is no solution left in the wells.
3. Add 100 μL of the Detection Antibody to each well. Incubate at room temperature for 1 hour on orbital plate shaker.
4. Wash the plate as in step 2.
5. Add 100 μL of TMB Substrate to each well. Allow the color to develop for 10 minutes at room temperature.
6. Stop the reaction by adding 100 μL per well of 3N Sulfuric Acid (H_2SO_4).
7. Read the plate at 450 nm in a microplate reader within 15 minutes.

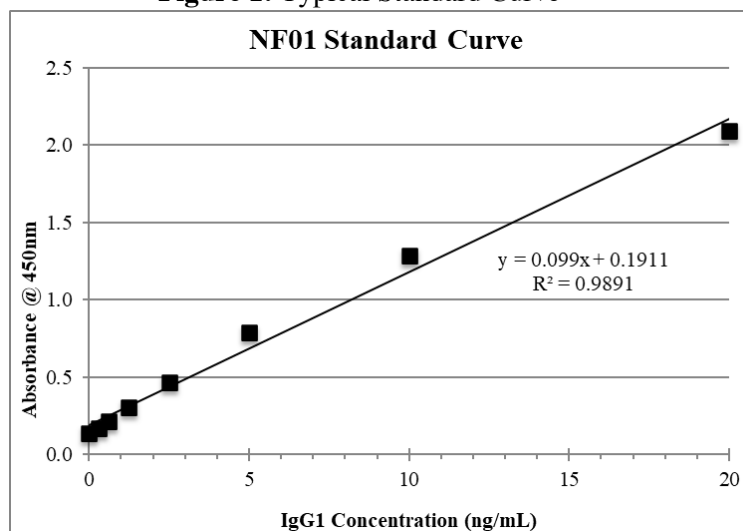
Scheme 1: Suggested Plate Layout

	1	2	3	4	5	6	7	8	9	10	11	12
A	S0	S0	U1	U1	U9	U9	U17	U17	U25	U25	U33	U33
B	S1	S1	U2	U2	U10	U10	U18	U18	U26	U26	U34	U34
C	S2	S2	U3	U3	U11	U11	U19	U19	U27	U27	U35	U35
D	S3	S3	U4	U4	U12	U12	U20	U20	U28	U28	U36	U36
E	S4	S4	U5	U5	U13	U13	U21	U21	U29	U29	U37	U37
F	S5	S5	U6	U6	U14	U14	U22	U22	U30	U30	U38	U38
G	S6	S6	U7	U7	U15	U15	U23	U23	U31	U31	U39	U39
H	S7	S7	U8	U8	U16	U16	U24	U24	U32	U32	U40	U40

CALCULATIONS

- a. Average all duplicate well absorbance values.
- b. Plot a standard curve using the absorbance values of each Standard (y-axis) versus the Standard concentration (x-axis).
- c. Determine the concentration of each unknown using the equation of the line.

Figure 1: Typical Standard Curve



REFERENCES

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3. Stubenrauch K, Wessels U, Lenz H.; (2009) Evaluation of an immunoassay for human-specific quantitation of therapeutic antibodies in serum samples from non-human primates. J Pharm Biomed Anal.; 49(4):1003-8

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