Venor®GeM qOneStep

Mycoplasma Detection Kit for qPCR

INSTRUCTIONS FOR USE

FOR USE IN RESEARCH AND QUALITY CONTROL

Symbols

Lot No.

Order No.

Expiry date

Storage temperature

Number of reactions

Manufacturer

INDICATION

Venor®GeM qOneStep mycoplasma detection kit is designed for the direct detection of mycoplasma contaminations in cell cultures, cell culture media, and other biological matrices.

TEST PRINCIPLE

The Venor®GeM qOneStep assay utilizes qPCR as the established method of choice for the rapid, robust and sensitive detection of mycoplasma contaminations. The assay targets a highly conserved region within the mycoplasma genome to detect prevalent cell culture contaminants such as *M. orale*, *M. hyorhinis*, *M. arginini*, *M. fermentans*, *M. salivarium*, and *M. hominis*, but also the less frequent strains *M. pneumoniae*, *Acholeplasma laidlawii*, *M. synoviae* and *Ureaplasma* species (see list of detected mycoplasma species in section "Assay Characteristics"). Venor®GeM qOneStep kit is able to detect all mycoplasma species in a single experiment, whereas eukaryotic and bacterial DNA is not amplified. The procedure takes less than 3 hours, and, in contrast to other methods like luminescence-linked enzymology, fluorescent staining or culture methods, there is no need for vital cells. Notably, the detection by qPCR is considered to be superior in terms of sensitivity and precision.

Mycoplasma are specifically detected by amplifying the 16S rRNA coding region in the mycoplasma genome. The kit contains all necessary PCR components including hot-start Taq polymerase, primers, and dNTPs. False-negative results caused by PCR inhibition and/or DNA extraction issues will be reliably identified by means of the Internal Control DNA, already included in the qOneStep Mix. The internal control amplification is detected at 560 nm (HEX™ channel), whereas the mycoplasma-specific amplification is detected at 520 nm (FAM™ channel).

The qOneStep Mix contains dUTP instead of dTTP to facilitate the degradation of amplicon carryover by use of uracil-DNA glycosylase (UNG). Thus, the probability of false-positive results is minimized. Please note that UNG is not included in the Venor®GeM qOneStep kit.

REAGENTS

Each kit contains reagents for 25, 100, or 250 reactions. The expiry date of the unopened package is marked on the package label. The kit components must be stored at +2 to +8 °C until use. The rehydrated mix must be stored at ≤ – 18 °C.

Component	Quantity			Cap color
	25 reactions Order No. 11-91025	100 reactions Order No. 11-91100	250 reactions Order No. 11-91250	
qOneStep Mix	1 × (lyophilized)	4 × (lyophilized)	10 × (lyophilized)	red
Rehydration Buffer	1 × 1.8 ml	2 × 1.8 ml	5 × 1.8 ml	blue
Positive Control DNA	1 × (lyophilized)	1 × (lyophilized)	1 × (lyophilized)	green
PCR grade water	1 × 2.0 ml	1 × 2.0 ml	1 × 2.0 ml	white

The LOT-specific quality control certificate (Certificate of Analysis) can be downloaded from our website (www.minerva-biolabs.com).

USER-SUPPLIED CONSUMABLES AND EQUIPMENT

The Venor®GeM qOneStep kit contains necessary reagents for setting up the PCR. Additional consumables and equipment are supplied by the user:

- qPCR device with filter sets for detecting the fluorescence dyes FAM™ and HEX™
- PCR reaction tubes and caps for the specific qPCR device
- 1.5 ml reaction tubes, DNase- and RNase-free
- Microcentrifuge for 1.5 ml reaction tubes
- Pipettes with corresponding filter tips (10, 100, and 1000 µl)
- Optional for carry-over prevention: Uracil DNA glycosylase (UNG)

SPECIMEN

Samples should be obtained from cell cultures with 80 to 90 % confluence. Cell culture supernatant is very well suited for the mycoplasma test without the need of additional sample preparation. However, PCR inhibiting substances may accumulate in the medium of cell cultures, making it necessary to extract the DNA prior to the PCR test (see below for further information). Note that penicillin or streptomycin in culture media are not known to inhibit mycoplasma nor affect the test's sensitivity.

The average mycoplasma concentration in cell culture is ~ 10^6 particles per ml with a maximum of 10^8 particles per ml. Within this range, a sufficient amount of mycoplasma DNA is present in the supernatant for successfully applying the qPCR test. Prepare the qPCR template as follows:

1.	Transfer 100 µl of cell culture supernatant to a sterile 1.5 ml reaction tube. Close the lid tightly.
2.	Incubate the sample at 95 °C for 5 minutes.
3.	Centrifuge the sample briefly (15 sec) at max speed to pellet any cellular debris.
4.	Use 2 µl of the supernatant directly for qPCR, or store the sample for up to 6 days at +2 to +8 °C or at ≤ – 18 °C for long term storage.

Please note, that cell pellets, just like foetal calf serum, vaccines, cryo stocks, and paraffin-embedded samples, cannot be used directly for the test due to PCR inhibiting substances that will interfere with the reaction. These samples would therefore require DNA extraction prior to PCR. The Venor®GeM qOneStep assay was qualified with the following DNA extraction kits:

- Venor®GeM Sample Preparation kit (Order No. 56-1100) for manual DNA extraction, or
- Venor®GeM Sample Preparation kit—IP C16 (Order No. 56-2096) for automated DNA extraction.

Extracted DNA can be stored at +2 to +8 °C for up to 6 days or at ≤ – 18 °C for long term storage.

PRECAUTIONS

Venor®GeM qOneStep kit is for research use only. The kit should be used by trained laboratory staff only.

All samples should be considered as potentially infectious and handled with all due care and attention. Always wear a suitable lab coat and disposable gloves.

This kit does not contain hazardous substances. Remnants can be discarded according to local regulations.

Cross contaminations may lead to false-positive test results. Thus, all tests should be performed according to good laboratory practice.

IMPORTANT NOTES

⇨ These instructions must be understood to successfully use the Venor®GeM qOneStep kit. The reagents supplied should not be mixed with reagents from different batches but used as an integral unit. The reagents of the kit must not be used beyond their shelf life.

⇨ Follow the exact protocol. Any deviation may affect the test method and results.

⇨ PCR inhibition is likely to be caused by the sample matrix. Thus, we recommend our Venor®GeM Sample Preparation kits. Any other DNA extraction kit needs to be qualified.

⇨ It is important to include control samples on a regular basis to monitor the reliability of your results. Positive and negative controls are essential in case of troubleshooting.

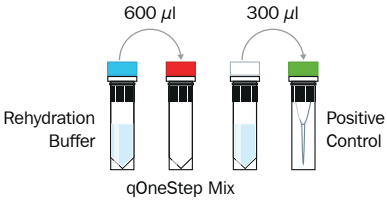
⇨ Set up at least one negative control sample (non template control) in each PCR. Use the elution buffer for the NTC in case of extracted DNA.

⇨ The control samples must be processed in the same manner as the test samples. You may want to include other laboratory specific control samples such as high, median and low DNA levels (e.g. $3 \times LOD_{95}$).

PROCEDURE – OVERVIEW

1. Reagent Preparation

qOneStep Mix and Positive Control

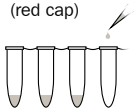


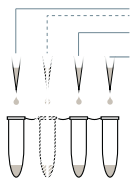
for 5 min RT
briefly
for 5 sec

2. Preparation of PCR Reactions

loading the test tubes

aliquot 23 µl qOneStep Mix (red cap)

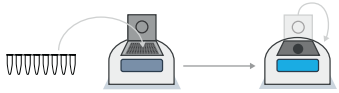




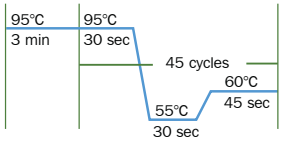
+ 2 µl sample
+ 2 µl Positive Control (green cap)
+ 2 µl fresh cell culture medium (Negative Control)

briefly

3. Start PCR amplification



Start PCR program



Rehydration Buffer
qOneStep Mix
PCR grade water
Positive Control

incubate
vortex
centrifuge
add

This procedure overview is not a substitute for the detailed manual.

MB_SI_Venor-GeM-qOneStep_01_EN

PROCEDURE - STEP BY STEP

The test should be carried out with negative and positive controls and samples in duplicates. All reagents and samples must be equilibrated to +2 to +8 °C prior use. After reconstitution, the reagents must be stored at ≤ -18 °C. Repeated freezing and thawing should be avoided. For small sample numbers, we recommend the preparation of aliquots of reconstituted qOneStep Mix and Positive Control DNA.

1. Reagent preparation

1.	qOneStep Mix Positive Control DNA	Red cap Green cap	Spin down all lyophilized components at max speed for 5 sec
2.	qOneStep Mix	Red cap	Add 600 µl Rehydration Buffer (blue cap) For sample kit only: Add 240 µl Rehydration Buffer
3.	Positive Control DNA	Green cap	Add 300 µl of PCR grade water (white cap)
4.	qOneStep Mix Positive Control DNA	Red cap Green cap	Incubate at room temperature for 5 min
5.	qOneStep Mix Positive Control DNA	Red cap Green cap	Vortex briefly and spin down for 5 sec

2. Preparation of PCR reactions

Follow this scheme to set up the test:

1.	Aliquot 23 µl of qOneStep Mix to each PCR tube.
2.	Negative Controls: add 2 µl fresh cell culture medium or elution buffer from DNA extraction kit (see chapter “Specimen“)
3.	Samples: add 2 µl of cell culture supernatant or DNA extract.
4.	Positive Control: add 2 µl Positive Control DNA (green cap).
5.	Close the PCR tubes tightly and spin down briefly.

3. Start qPCR amplification

1.	Place PCR tubes in the qPCR device and close the lid.
2.	Program the qPCR cycler (a technical note with detailed cycler programs of selected qPCR cyclers is available on our website www.minerva-biolabs.com).
3.	Start the program.

This assay was tested on the following qPCR devices:

qPCR device	Manufacturer
CFX96™	Bio-Rad Laboratories
LightCycler® 2.0	Roche Diagnostics
ABI Prism® 7500	Applied Biosystems
RotorGene® 6000	Corbett Research
Mx3005P®	Agilent Technologies
AriaMx	Agilent Technologies

For the detailed qPCR cycler programs please visit our website www.minerva-biolabs.com

INTERPRETATION OF RESULTS

The presence of mycoplasma is indicated by an increasing fluorescence signal in the FAM™ channel. The quantification is based on threshold cycle (C_t) values and a DNA standard curve. The exact procedure for obtaining C_t-values including baseline calculation/normalization depends on the particular qPCR device and cycler control software. Please see the documentation of your device for further details. We recommend the assessment of the amplification curve progression of all samples including control samples.

A positive PCR is indicated by $C_t < 40$. PCR reactions with $C_t \geq 40$ are considered negative. In addition, a successful PCR is displayed by an increasing fluorescence signal in either the FAM™ or the HEX™ channel, or both. The mycoplasma DNA and Internal Control function as competitors in the PCR. Thus, the more mycoplasma DNA is in the sample, the higher the signal in the FAM™ channel and the lower the internal control signal in the HEX™ channel. The following table will help with the interpretation of PCR results:

Detection of Mycoplasma FAM™ channel	Internal control HEX™ channel	Interpretation
positive	irrelevant	Mycoplasma are detected in the sample
negative	negative	PCR inhibition
negative	positive	No Mycoplasma are detected in the sample

ASSAY CHARACTERISTICS

Venor®GeM qOneStep assay detects the following Mollicute species, but not any of the phylogenetically related microorganisms, such as *Clostridium*, *Lactobacillus* and *Streptococcus*. Also, the waterborne germ *Burkholderia* is not detected. Cross-reactivity with other bacterial and mammalian DNA was tested negative:

Positively tested Mollicutes	**Negatively tested**	
	Bacteria	**Mammals**
Acholeplasma laidlawii	*Clostridium acetobutylicum*	Vero-B4
Mycoplasma hyorhinis	*Lactobacillus acidophilus*	Per.C6
Mycoplasma fermentans	*Streptococcus pneumoniae*	RK13
Mycoplasma orale	*Chlamydia trachomatis*	CHO-K1
Mycoplasma synoviae	*Legionella pneumophila*	Murine genomic DNA
Mycoplasma pneumoniae	*Micrococcus luteus*	Calf thymus DNA
Mycoplasma arginini	*Candida albicans*	Foetal bovine serum
Mycoplasma gallisepticum	*Enterococcus faecalis*	Horse serum
Spiroplasma citri	*Enterobacter aerogenes*	Goat serum
Mycoplasma arthritidis	*Escherichia coli*	
Mycoplasma genitalium	*Proteus mirabilis*	
Mycoplasma hominis	*Burgholderia cepacia*	
Mycoplasma penetrans		
Mycoplasma salivarium		
Ureaplasma urealyticum		

APPENDIX

Limited Product Warranty

This warranty limits our liability for replacement of this product. No warranties of any kind, express or im-plied, including, without limitation, implied warranties of merchantability or fitness for a particular purpose, are provided. Minerva Biolabs shall have no liability for any direct, indirect, consequential, or incidental damages arising from of the use, the results of use, or the inability to use this product.

Trademarks

LightCycler is a registered trademark of a member of the Roche Group. ABI Prism is a registered trademark of Applera Corporation or its subsidiaries in the US and certain other countries. CFX96 Touch is a trademark of Bio-Rad Laboratories, Inc. FAM, and HEX are trademarks of Applied Biosystems LLC. Venor, Onar, Mynox, and ZellShield are registered trademarks and Mycoplasma Off, PCR Clean and WaterShield are trademarks of Minerva Biolabs GmbH.

Related Products

Contamination Control Kits for conventional PCR

11-1025/-1050/-1100/-1250	Venor®GeM Classic Mycoplasma Detection Kit	25/50/100/250 tests
11-7024/-7048/-7096/-7240	Venor®GeM Advance Mycoplasma Detection Kit	24/48/96/240 tests
11-8025/-8050/-8100/-8250	Venor®GeM OneStep Mycoplasma Detection Kit	25/50/100/250 tests
12-1025/-1050/-1100/-1250	Onar® Bacteria Detection Kit	25/50/100/250 tests

Contamination Control Kits for qPCR

11-9025/-9100/-9250	Venor®GeM qEP Mycoplasma Detection Kit	25/100/250 tests

Sample Preparation

56-1010/1050/1200	Venor®GeM Sample Preparation Kit	10/50/200 extractions
56-2096	Venor®GeM Sample Preparation Kit - IP C16	96 extractions

Mycoplasma Elimination

10-0200/0500/1000	Mynox® Mycoplasma Elimination Reagent	2/5/10 treatments
10-0201/0501/1001	Mynox® Gold Mycoplasma Elimination Reagent	2/5/10 treatments

PCR Quantification Standards, 1 x 10^8 genomes / vial

52-0112	*Mycoplasma orale*
52-0115	*Mycoplasma gallisepticum*
52-0116	*Acholeplasma laidlawii*
52-0117	*Mycoplasma fermentans*
52-0119	*Mycoplasma pneumonia*
52-0124	*Mycoplasma synoviae*
52-0129	*Mycoplasma arginini*
52-0130	*Mycoplasma hyorhinis*
52-0164	*Spiroplasma citri*

See Minerva homepage for further available species

Food and Water Assays

11-02-XX-025	Food Control™ qPCR	25 reactions
12-01-005/-020/-040	Meat ID™ Screen	5/20/40 tests
12-02-025/-100	Meat ID™ Halal	25/100 reactions
12-05-025/-100	Vegan Control™	25/100 reactions
33-2025/-2100/-2250	AquaScreen® Legionella species	25/100/250 reactions
34-2025/-2100/-2250	AquaScreen® Legionella pneumophila	25/100/250 reactions
34-6025/-6100/-6250	AquaScreen® Pseudomonas aeruginosa	25/100/250 reactions
34-7025/-7100/-7250	AquaScreen® Escherichia coli	25/100/250 reactions

PCR Cycler Validation

57-2102	PCR Cycler Check™ Advance	6 strips, 8 vials each
57-2103	PCR Cycler Check™ OneStep	100 reactions
57-2202	qPCR Cycler Check™	100 reactions

PCR Clean™ (formerly DNA Remover™)

15-2025/15-2200	DNA Decontamination Reagent, spray bottle/refill bottles	250 ml/4× 500 ml
15-2201	Wipes	120 wipes in a dispenser box
15-2202	Wipes, refill packs	5×120 wipes in a bag

Mycoplasma Off™

15-1000	Surface Disinfectant Spray, spray bottle	1000 ml
15-5000	Surface Disinfectant Spray, refill bottles	5×1000 ml
15-1001	Surface disinfectant Wipes in dispenser box	120 wipes
15-5001	Surface Disinfectant Wipes, refill pack	5×120 wipes

ZellShield®

13-0050/-0150	Contamination Prevention Reagent 100× concentrate	1000 ml/ 5×1000 ml

WaterShield™

15-3015/3020/3050	Water Disinfection Additive for incubators and water baths, 200× concentrate	30×5 ml/3×50 ml/500 ml

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