

# **Venor®GeM Advance**

Pre-aliquoted Mycoplasma Detection Kit for conventional PCR

**INSTRUCTIONS FOR USE** 

FOR USE IN RESEARCH AND QUALITY CONTROL

## **Symbols**



Lot No.



Order No.



**Expiry date** 



Storage temperature



**Number of extractions** 



Manufacturer

#### INDICATION

The Venor®GeM Advance is designed for direct detection of mycoplasma contamination in cell cultures.

#### **TEST PRINCIPLE**

Venor®GeM Advance utilizes the polymerase chain reaction (PCR), which is established as the method of choice for highly sensitive detection of Mycoplasma and Acholeplasma contamination in cell cultures and other cell culture derived biologicals. Detection requires as little as 5 fg of mycoplasma DNA corresponding to 20 mycoplasma per sample volume. The detection procedure can be performed within 3 hours. Venor®GeM Advance is intended for research use only, not for clinical diagnostics or testing of human samples.

The primer set is specific to the highly conserved 16S rRNA coding region in the mycoplasma genome. This allows the detection of *M. orale, M. hyorhinis, M. arginini, M. fermentans, M. salivarium, M. hominis,* usually encountered as contaminants in cell cultures as well as *M. pneumoniae, Acholeplasma laidlawii, M. synoviae* and *Ureaplasma* species. Cross-detection of the following bacteria is not monitored: *Clostridium acetobutylicum, Lactobacillus acidophilus* and *Streptococcus pneumoniae*. Furthermore, human DNA is not detectable with this kit. See further details on page 6.

The provided reaction tubes are pre-coated with lyophilized Taq polymerase, nucleotides, primer, and loading dye to significantly reduce the hands-on time. Hence, the PCR product can be loaded directly on the agarose gel. *The pre-coated tubes* also contain an internal control DNA to verify the PCRs' success.

The reaction mixture contains dUTP instead of dTTP, so the option is available to degrade amplicons from previous analysis by use of uracil-DNA glycosylase (UNG). Thus, the occurrence of false-positive result can be minimized. UNG is not part of the product.

## **COMPONENTS**

Each kit contains reagents for 24, 48, 96 or 240 reactions. The expiry date of the unopened package is marked on the package label. The kit components are stored until use at +2 to +8 °C and must be stored after opening and rehydration at < -18 °C.

| Component                       | Description  | Content   |
|---------------------------------|--|---|
| Test Reaction Tubes             | tubes pre-coated with freeze-dried<br>primers, polymerase, dNTPs, inter-<br>nal control DNA, and gel loading<br>buffer/dye | 3, 6, 12, or 30 strips of 8 transparent tubes each, depending on package size |
| Positive Control Reaction Tubes | Test Reaction Tubes containing additional DNA of <i>Mycoplasma</i> orale genome  | 1, 2, 3, or 5 strips of 8 red tubes each, depending on package size           |
| Rehydration Buffer              |  | 1, 2, or 5 tubes à 1.6 ml, depending on package size                          |
| Caps for PCR Tubes              |  | 4, 6, 15, or 35 strips of 8 tubes each, depending on package size             |

The lot specific quality control certificate (*Certificate of Analysis*) can be downloaded from our website (www.minerva-biolabs.com).

## **USER-SUPPLIED CONSUMABLES AND EQUIPMENT**

The Venor®GeM Advance kit contains all necessary components for setting up the PCR. Additional consumables and equipment is supplied by the user:

- PCR cycler
- Pipettes with corresponding filter tips (10 and 100  $\mu$ l)
- 1.5 ml reaction tubes
- Agarose gel electrophoresis system including DNA stain

## SPECIMEN FOR CELL CULTURE SCREENING

Samples should be obtained from cell cultures that are highly confluent (90 % or higher). Cell culture supernatant is very well suited for the mycoplasma test without the need of additional sample preparation. However, PCR inhibiting substances may accumulate in the medium of cell cultures, which may make is necessary to extract the DNA prior to the PCR test (see further information below). Note that penicillin or streptomycin in the culture media are not known to inhibit mycoplasma nor affect the tests' sensitivity.

The average mycoplasma number in cell culture is  $\sim 10^6$  particles per ml with a maximum of  $10^8$  particles per ml. Within this range, a sufficient amount of mycoplasma DNA is present in the supernatant for successfully applying the PCR test. Prepare the PCR template as follows:

- 1. Transfer 500  $\mu$ I of the cell culture supernatant from the test culture to a 1.5 ml reaction tube. The lid should be sealed tightly to prevent opening during heating.
- 2. Incubate the sample at 95 °C for 10 minutes.
- 3. Briefly centrifuge (5 seconds) the sample at approx. 13,000 rpm to pellet cellular debris.
- 4. Use 2  $\mu$ I of the supernatant directly for PCR.

Cell pellets cannot be used directly for the test due to cell debris that will interfere with the PCR reaction. However, cell pellets as well as foetal calf serum, vaccines, cryo stocks, and paraffin-embedded samples require DNA extraction in advance. The *OneStep* assay was qualified with these DNA extraction kits:

- Venor®GeM Sample Preparation kit (Order No. 56-1050) for manual DNA extraction, or
- Venor®GeM Sample Preparation Kit—IP C16 (Order No. 56-2096) for automated DNA extraction.

Extracted DNA can be stored at 4  $^{\circ}$ C for up to 6 days at –20  $^{\circ}$ C for long time storage.

## **PRECAUTIONS**

The Venor®GeM Advance kit is for research use only. The kit should be used by trained laboratory staff only.

All samples should be considered as potentially infectious and handled with all due care and attention. Always wear suitable lab coat and disposable gloves.

This kit does not contain hazardous substances. Remnants can be discarded according to local regulations.

Cross contamination may lead to false-positive test results. Thus all tests should be performed according to good laboratory practice.

## **IMPORTANT NOTES**

- These instructions must be understood to successfully use the *Venor®GeM Advance* kit. The components supplied should not be mixed with reagents from different lot and used as an integral unit. The reagents of the kit must not be used beyond their shelf life.
- ⇒ Follow the exact protocol. Any deviation may affect the test method and can affect the results.
- Avoid cross contamination by preparing the Positive Controls after the Negative Controls and Test Reactions
- ⇒ Set up at least one negative control sample (non template control) in each PCR. Use fresh cell culture medium or elution buffer for the NTC in case of extracted DNA.
- ⇒ PCR inhibition is likely to be caused by the sample matrix, or, in case of extracted DNA, caused by the elution buffer. Thus we recommend our *Venor®GeM Sample Preparation* kits. Any other DNA extraction kit needs to be qualified.
- ⇒ The appearance of the pellets in the pre-coated PCR tubes may vary on technical reasons which has no effect on the outcome of the test.

## **PROCEDURE**

- ⇒ Set up negative and positive controls with each test.
- ⇒ The components must be equilibrated to room temperature prior use.

## 1. Component preparation

- 1. Remove required number of PCR tubes from the kit (cut tubes from strips if necessary) and close bags with remaining tubes properly.
- 2. Spin down PCR tubes and Rehydration Buffer.
- Peel off the sealing film from the PCR tubes and add 23  $\mu$ l of the Rehydration Buffer.
  - **Note:** Rehydrate Positive Control Tubes with 25  $\mu$ l of Rehydration Buffer.

## 2. Add samples

Follow the pipetting sequence and close tubes after the sample was added.

| 1. | Negative Controls: | add 2 $\mu\text{I}$ fresh cell culture medium or elution buffer supplied with the DNA extraction kit |  |
|----|--------------------|--|--|
| 2. | Samples:           | add 2 $\mu$ I of cell culture supernatant or DNA extract.  |  |
| 3. | Positive Controls: | no addition of reagents necessary  |  |

4. Close and spin all PCR tubes briefly, load the PCR cycler and start the program.

## 3. Start PCR amplification

1. Place the PCR tubes in the cycler and close the lid tightly.

Program the PCR cycler or check stored temperature profiles.

1 cycle 94 °C for 2 min 39 cycles 94 °C for 30 sec 55 °C for 30 sec 72 °C for 30 sec

Hold from 4 °C to 8 °C

3. Start the program.

#### 4. Agarose gel electrophoresis

- 1. Cast a 1.5 or 2 % agarose gel including DNA stain (max. 5 mm thick, 5 mm comb).
- 2. Load 5  $\mu$ l of each PCR product. No loading dve is required.
- 3. Perform the gel electrophoresis (e.g. 20 min at 100 V).
- 4. Visualize the PCR results on a suitable transilluminator.

2.

## RESULT INTERPRETATION

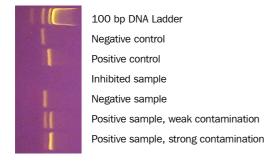
The Internal Control DNA gives rise to a distinct 191 bp band in every lane indicating a successfully performed PCR. This band will fade out with increased amounts of primary target amplification (e.g. mycoplasma DNA input of  $> 5 \times 10^6$  copies per ml. The initial concentration of positive control DNA is higher than  $5 \times 10^6$  copies per ml. Consequently, the internal control is usually not visible in the positive control reaction.

Other PCR products may be visible in the gel as faint, diffuse bands of different sizes (neither 191 bp nor  $\sim$ 270 bp). This does not indicate positive results. These products are unspecific and caused by non-specific annealing (e.g. high DNA input of > 100  $\mu$ g/ml). Also, primer self-annealing may give rise to a band of 80-90 bp in size. This again does not affect the sensitivity and precision or results of the test.

If the PCR test shows inhibition due to the sample (lower band intensity compared to negative control) a DNA extraction needs to be performed prior to re-testing the sample (see chapter "Specimen").

| Detection of <i>Mycoplasma</i><br>band at 265-278 bp | Internal control<br>band at 191 bp | Interpretation                         |
|--|------------------------------------|--|
| Positive   | Irrelevant                         | Mycoplasma present in the sample       |
| Negative   | Negative                           | PCR inhibition                         |
| Negative   | Positive                           | No mycoplasma detectable in the sample |

Fig. 1: A typical agarose gel image



## ANALYTICAL CHARACTERISTICS OF THE TEST

## **Analytical Sensitivity**

The detection limit was determined with approx. 20 genome copies/PCR reaction for different mycoplasma species. Due to the small sample volume of 2  $\mu$ l this kit does not comply with the detection limit of 10 CFU/ml according to *European Pharmacopoeia* (EP) 2.6.7. For EP 2.6.7 compliant lot release testing of biopharmaceuticals please consider the product versions Venor®GeM Classic for conventional PCR or Venor®GeM qEP for real-time qPCR.

## **Cross Reactivity**

A cross-reactivity with eukaryotic DNA origin could not be found. Rarely unspecific PCR products can be formed and become visible in the gel as faint, diffuse bands of different sizes due to overloading the PCR (ref. chapter "Interpretation of Results"). The kit is also not detecting any of the phylogenetically related microorganisms, *Clostridium*, *Lactobacillus* and *Streptococcus*. Likewise, the water-born germ *Burgholderia* is not detected. But, at concentrations above 10<sup>4</sup> genomes/µl the test is positive for *Staphylococcus aureus*, *Staphylococcus epidermidis* and *Bacillus subtilis*.

## **Sequence Alignment**

A large number of *Mollicut*es sequences have been published. The primers of the kit were aligned with the NCBI data base and inspected for homologies within the target region of the 16S rRNA. At least 1 *Ureaplasma*, 7 *Acholeplasma* and 85 *Mycoplasma* show relevant sequence homologies and highest presumption of a positive PCR result.

| Positive tested Mollicutes   | Negative tested            |                        |                     |
|------------------------------|----------------------------|------------------------|---------------------|
| Positive tested infollicutes | EP listed bacteria         | Other bacteria         | Mammals             |
| Acholeplasma laidlawii       | Clostridium acetobutylicum | Chlamydia trachomatis  | Vero-B4             |
| Mycoplasma arginini          | Lactobacillus acidophilus  | Legionella pneumophila | Per.C6              |
| Mycoplasma arthritidis       | Streptococcus pneumoniae   | Micrococcus Iuteus     | RK13                |
| Mycoplasma fermentans        |                            | Candida albicans       | CHO-K1              |
| Mycoplasma genitalium        |                            | Enterococcus faecalis  | Murine genomic DNA  |
| Mycoplasma hominis           |                            | Enterobacter aerogenes | Calf thymus DNA     |
| Mycoplasma hyorhinis         |                            | Escherichia coli       | Foetal bovine serum |
| Mycoplasma orale             |                            | Proteus mirabilis      | Horse serum         |
| Mycoplasma penetrans         |                            | Bacillus cereus        | Goat serum          |
| Mycoplasma pneumoniae        |                            |                        |                     |
| Mycoplasma salivarium        |                            |                        |                     |
| Mycoplasma synoviae          |                            |                        |                     |
| Ureaplasma urealyticum       |                            |                        |                     |

## **APPENDIX**

## Limited Product Warranty

This warranty limits our liability for replacement of this product. No warranties of any kind, express or implied, including, without limitation, implied warranties of merchantability or fitness for a particular purpose, are provided. Minerva Biolabs shall have no liability for any direct, indirect, consequential, or incidental damages arising from of the use, the results of use, or the inability to use this product.

#### Trademarks

Venor® is a registered trademark of Minerva Biolabs GmbH, Germany.

#### **Related Products**

| MAD  | T   | DAIA | Polymerase |
|------|-----|------|------------|
| IVID | ıau | DINA | Polymerase |

| 53-0050/-0100/-0200/-0250 | MB Taq DNA Polymerase (5 U/μl) | 50/100/200/250 units |
|---------------------------|--------------------------------|----------------------|
| 53-1050/-1100/-1200/-1250 | MB Tag DNA Polymerase (1 U/ul) | 50/100/200/250 units |

#### **Contamination Control Kits for conventional PCR**

| 11-1025/-1050/-1100/-1250 | Venor®GeM Classic Mycoplasma Detection Kit | 25/50/100/250 tests |
|---------------------------|--|---------------------|
| 11-7024/-7048/-7096/-7240 | Venor®GeM Advance Mycoplasma Detection Kit | 24/48/96/240 tests  |
| 11-8025/-8050/-8100/-8250 | Venor®GeM OneStep Mycoplasma Detection Kit | 25/50/100/250 tests |
| 12-1025/-1050/-1100/-1250 | Onar® Bacteria Detection Kit               | 25/50/100/250 tests |

#### **Sample Preparation**

| 56-1010/1050/1200 | Venor®GeM Sample Preparation Kit          | 10/50/200 extractions |
|-------------------|---|-----------------------|
| 56-2096           | Venor®GeM Sample Preparation Kit - IP C16 | 96 extractions        |

#### **Mycoplasma Elimination**

| 10-0200/0500/1000 | Mynox® Mycoplasma Elimination Reagent      | 2/5/10 treatments |
|-------------------|--|-------------------|
| 10-0201/0501/1001 | Mynox® Gold Mycoplasma Elimination Reagent | 2/5/10 treatments |

## PCR Quantification Standards, 1 x 108 genomes / vial

| 52-0112  | Mycoplasma orale         |  |
|--|--------------------------|--|
| 52-0115  | Mycoplasma gallisepticum |  |
| 52-0116  | Acholeplasma laidlawii   |  |
| 52-0117  | Mycoplasma fermentans    |  |
| 52-0119  | Mycoplasma pneumonia     |  |
| 52-0124  | Mycoplasma synoviae      |  |
| 52-0129  | Mycoplasma arginini      |  |
| 52-0130  | Mycoplasma hyorhinis     |  |
| 52-0164  | Spiroplasma citri        |  |
| See Minerva homepage for further available species |                          |  |
|  |                          |  |

## 10CFU™ Sensitivity Standards, 3 vials with 10 CFU each, 2 vials negative control

| 102-1003 | Mycoplasma arginini      |
|----------|--------------------------|
| 102-2003 | Mycoplasma orale         |
| 102-3003 | Mycoplasma gallisepticum |
| 102-4003 | Mycoplasma pneumoniae    |
| 102-5003 | Mycoplasma synoviae      |
| 102-6003 | Mycoplasma fermentans    |
| 102-7003 | Mycoplasma hyorhinis     |
| 102-8003 | Acholeplasma laidlawii   |
| 102-9003 | Spiroplasma citri        |

102-0002 Mycoplasma Set, all EP 2.6.7 listed species 2 vials per species, 10 CFU each

## PCR Clean™ (formerly DNA Remover™)

| 15-20 | 25/15-2200 | DNA Decontamination Reagent, spray bottle/refill bottles | 250 ml/4x 500 ml             |
|-------|------------|--|------------------------------|
| 15-22 | 201        | Wipes  | 120 wipes in a dispenser box |
| 15-22 | 202        | Wipes, refill packs                                      | 5 x 120 wipes in a bag       |
| 15-22 | 203        | Wipes, single wrapped                                    | 30 Sachets                   |

#### Mvcoplasma Off™

| myoopiaoma on |   |               |
|---------------|---|---------------|
| 15-1000       | Surface Disinfectant Spray, spray bottle    | 1000 ml       |
| 15-5000       | Surface Disinfectant Spray, refill bottles  | 5 x 1000 ml   |
| 15-1001       | Surface disinfectant Wipes in dispenser box | 120 wipes     |
| 15-5001       | Surface Disinfectant Wipes, refill pack     | 5 x 120 wipes |
| 15-1030       | Wipes, single wrapped                       | 30 sachets    |
|               |   |               |

## ZellShield™

|               |   | 1000 ml/ 5 x 1000 ml |
|---------------|---|----------------------|
| 13-0050/-0150 | Contamination Prevention Reagent 100x concentrate |                      |

#### WaterShield™

| 15-3025/-3075 | Water Disinfection Additive for incubators and water baths | 30 x 5 ml/500 ml   |
|---------------|--|--------------------|
| 13-3023/-3013 | Water Distriction Additive for incubators and water baths  | 30 X 3 HII/300 HII |

200x concentrate



## Manufacturer

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## **Ordering**

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## **Technical Service**

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## **Made in Germany**

Minerva Biolabs GmbH develops and manufactures products in accordance with DIN EN ISO 9001:2008 and DIN EN ISO 13485:2012 quality system requirement. Reg.No. SY 60096693 0001 & SX 60096692 0001



