

Protein G Agarose

Item No
223-51-01

Size
10 mL



INTRODUCTION

Protein G Agarose consists of recombinant protein G, which is produced in *E. coli* and after purification, is covalently immobilized onto 4% cross-linked agarose beads. Protein G agarose is suitable for the isolation of IgG antibodies using column or immunoprecipitation methods. DNA sequencing of native protein G (from Streptococcal group G) has revealed two IgG-binding domains as well as sites for albumin and cell surface binding (1 - 6). Protein G has been designed to eliminate the albumin and cell surface binding domains to reduce nonspecific binding while maintaining efficient binding of the Fc region of IgG's. With the removal of these binding domains, Protein G can be used to separate albumin from crude human IgG samples⁽⁷⁾.

Covalently coupled Protein G Agarose has been widely used for the isolation of a wide variety of immunoglobulin molecules from several mammalian species. Protein G has greater affinity for many more mammalian IgGs than Protein A (Table 1).

FORM/STORAGE

Protein G Agarose is supplied in a total volume of 15 mL consisting of 10 mL Protein G agarose suspended in 20% ethanol/PBS. Store at 2 - 8°C. Stable for a minimum of 1 year from date of receipt when stored at 2 - 8°C. Non-sterile.

SPECIFICATIONS

Ligand density:	~2 mg Protein G/mL drained gel
Bead structure:	4% highly cross-linked agarose
Bead size range:	45 - 165 um
Recommended working pH:	3 - 9
Binding capacity:	≥ 20mg/mL Human IgG

Note: Different immunoglobulins derived from the same species and from the same subclass can demonstrate deviations in the binding capacity; Protein G may hydrolyze at low pH.

Table 1. Relative Affinity of Immobilized Protein G and Protein A for Various Antibody Species and Subclasses of polyclonal and monoclonal IgG's⁽⁸⁾.

<u>Species/ Subclass</u>	<u>Protein G</u>	<u>Protein A</u>
MONOCLONAL		
Human		
IgG ₁	++++	++++
IgG ₂	++++	++++
IgG ₃	++++	---
IgG ₄	++++	++++
Mouse		
IgG ₁	++++	+
IgG _{2a}	++++	++++
IgG _{2b}	+++	+++
IgG ₃	+++	++
Rat		
IgG ₁	+	---
IgG _{2a}	++++	---
IgG _{2b}	++	---
IgG _{2c}	++	+
POLYCLONAL		
Rabbit	+++	++++
Cow	++++	++
Horse	++++	++
Goat	++	---
Guinea pig	++	++++
Sheep	++	+/-
Pig	+++	+++
Rat	++	+/-
Mouse	++	++
Chicken	+	---
Human IgG	++++	++++
Human IgM	+	---
Human IgE	+	---
Human IgA	+	---
--- (weak or no binding) → ++++ (Strong binding)		

PROCEDURE

PURIFICATION OF IgG MOLECULES

1. User Supplied Materials

- Buffers: see number 2 below.
- Disposable column with frits and reusable caps. KPL recommends using Pharmacia Biotech PD-10 empty disposable columns (Catalog No. 17-0438-01) or equivalent.

2. Buffer Preparation

- Wash/Binding Buffer:**
KPL Catalog No. 50-70-01 or prepare 0.1 M Sodium phosphate, 0.15 M NaCl, pH 7.4.
- Elution Buffer:**
KPL Catalog No. 50-68-01 or prepare 0.2 M Glycine, pH 2.85.
- Storage Buffer:**
KPL Buffer Kit, Catalog No. 50-69-01 or prepare 0.01 M NaH₂PO₄, 0.15M NaCl, 2.7 mM KCl, pH 7.4, 20% ethanol.

- Sample Preparation:** To insure proper ionic strength and pH are maintained for optimal binding, it is necessary to dilute serum samples, ascites fluid or tissue culture supernatant at least 1/1 with binding buffer. Alternatively, the sample may be dialyzed overnight against wash/binding buffer. KPL recommends using a 12,000 MW cutoff dialysis tubing with at least two buffer exchanges. Remove any particulate matter from the sample by centrifugation or filtration through a 0.8 µm filter.

4. Column and Resin Preparation:

- Pour 20% Ethanol in the bottom of a petri dish or in a flat bottom container. Float the frit on top of the ethanol. Using the large round end of a 1 mL pipette tip, press the frit firmly into the ethanol to force air out. Repeat this step until the frit is completely wet.
- Push the frit into the barrel of the column until it rests firmly on the bottom.
- With the cap removed, clip the end of the column to create a hole to allow liquid to flow through.
- Wash the frit with 5 column volumes of 1X Wash/Binding Buffer.
- Prepare a 1/1 suspension of resin in 1X Wash/Binding buffer. The required amount of agarose per mg immunoglobulin to be purified can be estimated by the binding capacity.

- Pour slurry into column. Allow column to flow by gravity to pack the column bed.
- Equilibrate the packed affinity resin with 10 column volumes (CV) of the wash/binding buffer. For example, if the packed bed is 1 mL, equilibrate with 10 mL Wash/Binding Buffer.

5. Sample Purification

- Gently apply sample to the column by layering onto the top of the resin. Be careful not to disturb the bed surface.
- Wash column with 10 CV of the 1X Wash/Binding buffer or until the absorbance of eluate at 280 nm approaches the background level.
- Before beginning the elution step, set up enough tubes to collect the entire elution volume as 1 mL fractions (4 CV will be used to elute the antibody). To each collection tube add 240 µL 5X Wash/Binding Buffer. To elute the antibody, gently add 1 mL 1X Elution Buffer to the top of the resin collecting the eluate in a prepared collection tube. Repeat until the entire volume has been collected up to 4 column volumes. **Note:** If the eluate is to be collected in a single bulk volume, add 240 µL 5X Wash/Binding Buffer per mL Elution Buffer to the collection vessel before starting the elution. Elution of bound immunoglobulin can be monitored by absorbance at 280 nm, if desired.

- Column Regeneration:** Once the sample has been eluted, wash the affinity matrix with 2 CV of elution buffer. Re-equilibrate the column with at least 10 CV of 1X Wash/Binding Buffer. When column is equilibrated, pH of eluate will be the same as that of the wash/binding buffer.

- Clean-in-Place:** With certain applications, substances which contain denatured proteins or lipids do not elute in the regeneration procedure. The following steps can be taken to clean the column:

- To remove strongly bound hydrophobic proteins, lipoproteins and lipids, wash the column with a non-ionic detergent (e.g. 0.1% Triton X-100) at 37°C, with a contact time of ~1 minute.
- Immediately re-equilibrate the column with 5 - 10 CV of 1X Wash/Binding Buffer.
- As an alternative, wash the column with 70% ethanol. Allow the column to stand for 12 hours. Re-equilibrate the column with 5 - 10 CV of 1X Wash/Binding Buffer.
- To remove precipitated or denatured substances, wash the column with 2 CV of 6M guanidine hydrochloride. Immediately re-equilibrate the column with 5 - 10 CV of 1X Wash/Binding Buffer (see step 5).

Recommended Column Volumes

Antibody Source	Recommended bed volume (mL) per mL sample
Immune Serum	2 mL
Tissue Culture Supernatant (with 10% fetal bovine serum)	0.2 mL
Tissue Culture Supernatant (serum-free)	0.01 mL
Ascites Fluid	2 mL

6. **Resin Storage:** Store affinity matrix in Storage Buffer at 2 - 8°C. **Do not** store the matrix frozen or at room temperature. The matrix can be stored in the column by sealing the outlets or removed from the column and stored as a slurry.

The product listed herein is for research use only and is not intended for use in human or clinical diagnosis. Nothing disclosed herein is to be construed as a recommendation to use this product in violation of any patents. However, said information and product are offered without warranty or guarantee since the ultimate conditions of use and the variability of the materials treated are beyond our control. We cannot be responsible for patent infringements or other violations that may occur with the use of this product. No claims beyond replacement of unacceptable material or refund of purchase price shall be allowed.

IMMUNOPRECIPITATION

For immunoprecipitation protocols, see references 9 - 12.

PRODUCT SAFETY AND HANDLING

See MSDS (Material Safety Data Sheet) for this product.

REFERENCES

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RELATED PRODUCTS

Protein G Agarose Kit	Catalog. No. 553-51-00
Protein A Agarose Kit	Catalog. No. 553-50-00
Protein A Agarose	Catalog. No. 223-50-01

See KPL's catalog for a complete list of antibodies, substrates, Western blotting and ELISA kits.

L-286-04