Assay Performance Characteristics:

Standard range: 500.0-0.98ng/mL
Limit of Detection: 7.8ng/mL
Background: OD<0.2 at 450nm
Coefficient of Determination: R-squared>0.98

Plate Template:

	1	2	3	4	5	6	7	8	9	10	11	12
A												
B												
C												
D												
E												
F												
G												
H												

References:

www.inbio.com

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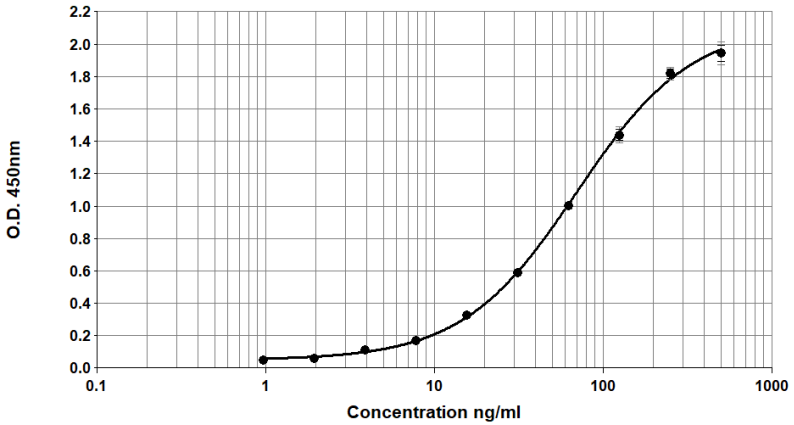
Tyr p 2 ELISA 2.0

Pre-coated Plate Kit

Product Code: EPC-TP2-X

Sample curve:



Contents:
Microtiter plate coated with purified polyclonal anti-*Tyrophagus putrescentiae*

Tyr p 2 allergen standard (white cap)
Concentration: 5,000ng/mL

Biotinylated purified polyclonal anti-*Tyrophagus putrescentiae* (brown cap)

Streptavidin-peroxidase (blue cap)

Wash buffer (10x concentrate)
Assay buffer (10x concentrate)
TMB developing substrate
Stop solution (0.5N sulfuric acid)

Store kit at 2-8^{O}C
Expiry: xxxxx xx, xxxx

For research and commercial use in vitro: not for human in vivo or therapeutic use.

Certificate of Analysis

Pre-coated Plate:	96-well polystyrene microtiter plate coated with polyclonal anti-*Tyrophagus putrescentiae* and treated with stabilizing agent. Sealed in foil pouch with desiccant.
Polyclonal Antibody:	Purified polyclonal anti-*Tyrophagus putrescentiae*
Immunogen:	Recombinant *Tyrophagus putrescentiae*, Tyr p 2
Isotype:	Multiple
Specificity:	The pAb contains IgG antibodies to storage mite *Tyrophagus putrescentiae*.
Purification:	The pAb is in phosphate buffered saline, pH 7.4 and has been 0.22µm filtered
Lot Number:	xxxxx

Detection Antibody:	Purified polyclonal anti-*Tyrophagus putrescentiae*
Immunogen:	Recombinant *Tyrophagus putrescentiae*, Tyr p 2
Isotype:	Multiple
Specificity:	Contains IgG antibodies to *Tyrophagus putrescentiae* allergen.
Purification:	Purified by affinity chromatography using Protein G. Single heavy and light chain bands on SDS-PAGE.
Biotinylation:	Biotinylated and titrated for use in ELISA at 1/1000 dilution. Prepared in 1% BSA/50% glycerol/PBS, pH 7.4, 0.22µm filtered, preservative free.
Lot Number:	xxxxx

Allergen Standard:	Purified recombinant Tyr p 2 prepared in 1% BSA/30% glycerol/PBS, pH 7.4.
Concentration:	5,000ng/mL (based on advanced protein assay)
Lot Number:	xxxxx

Materials required, but not provided:

- Type I ultrapure water or 18.2MΩ de-ionized water
- Volumetric measuring equipment (e.g. serological pipettes, graduated cylinders)
- Clean containers for buffer and reagent preparation
- Calibrated single and multi-channel micropipettes and tips
- Vortex mixer
- Plate reader capable of reading absorbance at 450nm
- Analysis software (recommended, but not required)

Protocol

Please read the entire protocol before starting the assay
Bring all reagents to room temperature before use

1. Prepare 1x working dilutions of the 10x wash and assay buffers in clean containers using 18.2MΩ de-ionized water or Type I ultrapure water. For one plate:
   Wash buffer: add 15mL concentrate to 135mL water
   Assay buffer: add 2.5mL concentrate to 22.5mL water
   Adjust volumes accordingly for multi-plate assays. Diluted buffers may be stored at 4^{O}C for up to 1 week
   *Use of an automated plate washer will require a larger volume of wash buffer than provided. Most PBS-based wash buffers should be compatible with the assay. Visit the support page at www.inbio.com for recommended wash buffer formulary.
2. Remove the plate from the foil pouch and wash by adding 150µL wash buffer to each well. Empty the wells by inverting the plate and then tap on absorbent paper to remove residual buffer. Repeat the wash cycle two more times.
3. Add standards, samples, and blanks to the plate (final volume in all wells is 100µL).

   Standards: add 180µL assay buffer into wells A1 and B1, and 100µL into remaining wells of rows A and B. Vortex the Tyr p 2 standard and add 20µL to wells A1 and B1. Mix well by pipetting up and down 7-10 times and then transfer 100µL into wells A2 and B2. Mix well and continue the serial doubling dilution scheme across the plate to column 10.
   The assay buffer in wells A11, B11 and A12, B12 will serve as **Blanks.**
   Samples: dust extracts are routinely tested starting at 1/10 dilution and can be prepared directly on the pre-coated plate: add 20µL sample to 180µL assay buffer. Mix, then transfer 100µL into 100µL assay buffer in the next well. Continue across the plate for the desired number of dilutions. A minimum of three dilutions per sample should be tested; 6-12 dilutions are recommended.
   Air filter extracts, allergen extracts, and other types of samples may require a different dilution scheme.
   *Sample dilutions may also be prepared in tubes or on a 96-well dilution plate and transferred to the pre-coated plate.
4. Incubate the plate at room temperature (away from direct sunlight) for 1 hour.
5. Wash the plate 3x with 150µL wash buffer per well. Vortex the biotinylated polyclonal antibody and prepare a 1:1,000 detection antibody mix by adding 10µL biotinylated pAb to 10mL assay buffer. **Mix thoroughly** and add 100µL to each well.
6. Incubate the plate at room temperature (away from direct sunlight) for 1 hour.
7. Wash the plate 3x with 150µL wash buffer per well. Vortex the conjugate and prepare a 1:1,000 conjugate mix by adding 10µL streptavidin-peroxidase to 10mL assay buffer. **Mix thoroughly** and add 100µL to each well.
8. Incubate the plate at room temperature (away from direct sunlight) for 30 minutes.
9. Pour the TMB substrate and stop solution into separate basins so they are ready to use in the next step. Wash the plate 3x with 150µL wash buffer per well.
10. Use a **multi-channel** pipette to add 100µL TMB to each well. Gently tap the plate and monitor the reaction as the blue color develops. Once OD450 reaches 0.08-0.09 for Standard 1, use a **multi-channel** pipette to add 50µL stop solution to each well (the color will change to yellow)
11. Read the plate at 450nm. The OD for Standard 1 should be between 1.2 and 3.5, with an ideal range of 2.0 - 2.5.