# **Assay Performance Characteristics:**

Standard range: 40-0.08ng/mL Limit of Detection: 0.32ng/mL Background: OD<0.1 at 450nm

Coefficient of Determination: R-squared>0.98

# **Plate Template:**

	1	2	3	4	5	6	7	8	9	10	11	12
A												
В												
С												
D												
Е												
F												
G												
Н												

#### References:

- Arruda, LK, Platts-Mills, TAE, Fox, JW and Chapman, MD (1990). Aspergillus fumigatus allergen I, a major IgE binding protein, is a member of the mitogillin family of cytotoxins. J. Exp. Med., 172:1529-1532.
- Arruda LK, Platts-Mills TAE, Longbottom JL, Chapman MD (1992). Aspergillus fumigatus: Identification of 16kd, 18kd and 45kd antigens recognized by human IgG and IgE antibodies and murine monoclonal antibodies. J. Allergy Clin. Immunol. 89:1166-1176.











Indoor Biotechnologies, Inc. 700 Harris Street, Charlottesville, VA 22903 United States

Tel: (434) 984-2304 Fax: (434) 984-2709

Email: mail@inbio.com

Indoor Biotechnologies Ltd. Vision Court Caxton Place Cardiff, Wales CF23 8HA United Kingdom

Tel: +44 (0) 29 2167 4640 Email: Info@indoorbiotech.co.uk Indoor Biotechnologies, India Private Limited Bangalore Bioinnovation Centre, BioTech Park, Electronic City Phase 1 Bangalore-560100, India

Tel: +91-9901722009

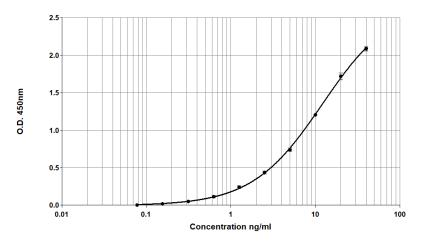
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# Asp f 1 ELISA 2.0 Pre-coated Plate Kit

Product Code: EPC-AF1-X Lot Number: XXXXX

## Sample curve:



#### Contents:

Microtiter plate coated with anti-Asp f 1 monoclonal antibody 4A6

Asp f 1 allergen standard (white cap) Concentration: 400ng/mL

Rabbit anti-Asp f 1 polyclonal antiserum (brown cap)

Peroxidase-conjugated goat anti-rabbit IgG (blue cap)

Wash buffer (10x concentrate)
Assay buffer (10x concentrate)
TMB developing substrate
Stop solution (0.5N sulfuric acid)

Store kit at 2-8°C Expiry: XXX. XX, XXXX

For research and commercial use in vitro: not for human in vivo or therapeutic use.



**Certificate of Analysis** 

**Pre-coated Plate:** 96-well polystyrene microtiter plate coated with

monoclonal antibody 4A6and treated with stabilizing

agent. Sealed in foil pouch with desiccant.

Monoclonal Antibody: 4A6 (clone 4A6 B5 E7)

Immunogen: Asp f 1

Isotype: Mouse IgG1

Specificity: Binds to a species specific epitope on Aspergillus

fumigatus allergen, Asp f 1.

Purification: Produced in tissue culture and purified by chromatog

raphy using Protein G. Single heavy and light chain

bands on SDS-PAGE.

Lot Number: XXXXX

**Detection Antibody:** Polyclonal antibody raised against partially purified Asp f

1 (1,2)

Specificity: The pAb contains IgG Ab to Asp f 1 as well as IgG Ab to

other Aspergillus allergens.

Activity: The pAb is in phosphate buffered saline, pH 7.4,

containing 1% BSA/50% glycerol/PBS. The pAb has been filtered and should be diluted 1/1000 for use in Asp f

1 ELISA.

Lot Number: XXXXX

Allergen Standard: nAsp r 1

Composition: Naturally purified Asp r 1 prepared in 1% BSA.

50% glycerol/PBS, pH 7.4

Concentration: 400ng/mL (based on amino acid analysis)

Calibration: The Asp r 1 concentration of the purified natural Asp r 1

was determined by amino acid analysis. Asp f 1 and Asp r 1 (mitogillin) are 95% homologous (differing by 4

amino acid residues) and are antigenically and

Functionally indistinguishable (1).

Lot Number: XXXXX

# Materials required, but not provided:

- Type I ultrapure water or 18.2MΩ de-ionized water
- Volumetric measuring equipment (e.g. serological pipette, graduated cylinder)
- Clean containers for buffer and reagent preparation
- Calibrated single and multi-channel micropipettes and tips
- Vortex mixer
- Plate reader capable of reading absorbance at 450nm
- Analysis software (recommended, but not required)

#### Protocol

## Please read the entire protocol before starting the assay

Bring all reagents to room temperature before use

1. Prepare 1x working dilutions of the 10x wash and assay buffers in clean containers using  $18.2M\Omega$  de-ionized water or Type I ultrapure water. For one plate:

**Wash buffer:** add 15mL concentrate to 135mL water **Assay buffer:** add 2.5mL concentrate to 22.5mL water Adjust volumes accordingly for multi-plate assays. \*Diluted buffers may be stored at 4°C for up to 1 week

- Remove the plate from the foil pouch and wash by adding 150µL wash buffer to
  each well. Empty the wells by inverting the plate and then tap on absorbent paper
  to remove residual buffer. Repeat the wash cycle two more times.
- 3. Add standards, samples, and blanks to the plate (final volume in all wells is 100 µL).

**Standards:** add 180 $\mu$ L assay buffer into wells A1 and B1, and 100 $\mu$ L into remaining wells of rows A and B. Vortex the Bla g 5 standard and add 20 $\mu$ L to wells A1 and B1. Mix well by pipetting up and down 7-10 times and then transfer 100 $\mu$ L into wells A2 and B2. Mix well and continue the serial doubling dilution scheme across the plate to column 10.

The assay buffer in wells A11, B11 and A12, B12 will serve as **Blanks**. **Samples:** Dust extracts are routinely tested starting at 1/10 dilution and may be prepared on the pre-coated plate: add  $20\mu L$  sample to  $180\mu L$  assay buffer in column A, mix, then transfer  $100\mu L$  into  $100\mu L$  assay buffer in the next well. Continue across the plate for the desired number of dilutions. A minimum of three dilutions per sample should be tested; 6-12 are recommended. Air filter extracts, allergen extracts, and other types of samples may require a different dilution scheme. \*Sample dilutions may also be prepared in tubes or on a 96-well dilution plate and transferred to the pre-coated plate.

- 4. Incubate the plate at room temperature (away from direct sunlight) for 1 hour.
- 5. Wash the plate 3x with  $150\mu$ L wash buffer per well. Vortex the polyclonal antibody and prepare a 1:1,000 detection antibody/conjugate mix by adding  $10\mu$ L polyclonal antibody and  $10\mu$ L peroxidase-conjugated goat ant-rabbit IgG to 10mL assay buffer.

Mix thoroughly and add 100µL to each well.

- 6. Incubate the plate at room temperature (away from direct sunlight) for 1 hour.
- 7. Pour the TMB substrate and stop solutions into separate basins so they are ready to use in the next step. Wash the plate 3x with 150µL wash buffer per well.
- Use a <u>multi-channel</u> pipette to add 100µL TMB to each well and monitor the reaction as the blue color develops. After 10-15 minutes, use a <u>multi-channel</u> pipette to add 50µL stop solution to each well (the color will change to yellow).
- 9. Read the plate at 450nm. The OD for Standard 1 should be between 1.2 and 3.5, with an ideal range of 2.0 2.5.

A list of frequently asked questions and troubleshooting guide can be found under the 'Support' tab on our web site: www.inbio.com.