

## Instructions for Use

### **Huma OsteoGelMA- Human Gelatin Methacrylate, Lyophilized, High Bloom**

For 2D-3D tissue engineering research, hydrogels and cell culture applications

**SKU:** BMHL

**Tissue Source:** Human Bone



**Humabiologics**

EMPOWERING TOMORROW'S INNOVATIONS

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## Product Description

**Huma OsteoGelMA** is a methacrylated version of Humabiologics', first of its kind, human bone gelatin. Its ability to be photocrosslinked allows for a tunable 3D gel with a wide range of applications including 3D bioprinting, 3D culturing, and many other tissue engineering and regenerative medicine research applications.

*Note: requires a photoinitiator such as Irgacure 2959 or LAP.*

Gelatin is a heterogenous mixture of water-soluble proteins with high average molecular masses. Gelatin is extracted by denaturing collagen-rich tissue in water [1]. The mechanical properties of gelatin, such as gel stiffness and compressibility, are related to the average molecular weight of the gelatin proteins and characterized by Bloom number [2]. **Humabiologics offers the first and only human gelatin methacrylate in the market derived from bone at Bloom numbers between 90-300 g to meet the needs of translational research and regenerative therapies.** Bloom number is proportional to the average gelatin molecular mass as described by the table below:

<u>Bloom Number</u>	<u>Average Molecular Mass</u>
50-125 (Low Bloom)	20,000-25,000
175-225 (Medium Bloom)	40,000-50,000
225-325 (High Bloom)	50,000-100,000

Biomedical applications of human gelatin include coating tissue culture surfaces with a thin layer of protein to improve cell attachment for a variety of cell types [3]. Due to the excellent biocompatibility of gelatin, it has been used to generate 3D scaffolds and hydrogels for tissue engineering applications [4] as well as a delivery material to control the release of bioactive molecules [5]. In the pharmaceutical industry, gelatin is used as an encapsulating agent and binder for tablet production [6]. Gelatin can be combined with other human-derived biomaterials- such as Collagen Type I from human skin (**HumaDerm offered by Humabiologics**) – to make completely human-derived tissue engineered composites [7].

## Source Tissue

**Huma OsteoGelMA** is extracted and purified from human bone sourced strictly from American Association of Tissue Banks (AATB), accredited and FDA registered tissue banks and organ procurement organizations (OPOs). Humabiologics strives to meet research needs by providing high quality biomaterials obtained from tissue partners who comply with requirements for transplantable human tissues under 21 CFR 1271 of the Food and Drug Administration (FDA).

## Precautions and Disclaimer

**Huma OsteoGelMA** is obtained from human tissue that has been tested and found negative for HIV-1 and -2, hepatitis B, and hepatitis C, as well as other infectious agents. Please review the Safety Data Sheet (SDS) for information regarding hazards and safe handling practices. **Huma OsteoGelMA is for research use only and is not intended for human use, diagnosis, screening, household, food or other uses.**

## Storage/Reconstitution

This product can be stored in airtight containers at ambient temperature (15-30 °C) upon receipt and prior to solubilization. The expiration date is 1 year from receipt of the bottle. Dry gelatin, when stored in airtight containers at ambient temperature, can remain unchanged for extended periods of time.

**Huma OsteoGelMA** can be reconstituted in 37°C water to the desired concentration with agitation/vortexing for several minutes. Dissolving time depends on the gelatin concentration but usually takes a few minutes for low concentration. Centrifugation might be used to remove undissolved particulate. After reconstitution, solution should be used immediately or stored at 2-10°C. Solubilized gelatin can be sterilized by filtration through a 0.22 µm membrane.

## Gelatin Crosslinking

*Note: The following are general recommendations. Researchers should optimize parameters based on their specific application.*

1. Dissolve Huma OsteoGelMA in 60°C 1x PBS or 1x cell culture media at the desired concentration. Recommended final concentration is  $\geq 5\%$ .
  - a. Mechanical properties of the hydrogel can be controlled by varying the Huma OsteoGelMA concentration, the photoinitiator concentration, and degree of crosslinking.
2. Dissolve photoinitiator and keep solution protected from light.
  - a. Irgacure 2959 can be dissolved in neat methanol or DMSO at a 10% concentration.
  - b. LAP can be dissolved in PBS at a 2.5% concentration.
3. Add the desired amount of photoinitiator to the Huma OsteoGelMA solution.
  - a. Depending on the application and bioprinting parameters, a photoinitiator concentration between 0.1 – 0.5% can be used.
  - b. Based on the photoinitiator used, crosslinking settings will vary.
    - i. For LAP, the crosslinking light should be at a wavelength of 405 nm.
    - ii. For Irgacure 2959, the crosslinking light should be at a wavelength of 365 nm.
4. In a sterile environment, use a syringe filter to sterile filter the Huma OsteoGelMA.
5. Place sterile Huma OsteoGelMA in 37°C incubator to cool.
6. Once cooled, add the cells to the sterile Huma OsteoGelMA/photoinitiator solution.
7. Load the cell/Huma OsteoGelMA/photoinitiator solution into the bioprinting syringe specific to the bioprinter according to the manufacturer's instructions.
8. Set up bioprinter and optimize parameters based on the printing condition, intended application, and bioprinter manufacturer instructions. The parameters needing to be optimized include nozzle diameter, pressure, nozzle temperature, print bed temperature, print speed, and crosslinking light source distance and intensity.
  - a. These are important parameters to optimize in order to achieve the desired extrudability of the Huma OsteoGelMA from the nozzle. Good extrudability means the bioink flows out of the nozzle as a filament as opposed to droplets.

## References

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4. Kang, H.W., Y. Tabata, and Y. Ikada, *Fabrication of porous gelatin scaffolds for tissue engineering*. Biomaterials, 1999. **20**(14): p. 1339-44.
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7. Gautam, S., et al., *Surface modification of nanofibrous polycaprolactone/gelatin composite scaffold by collagen type I grafting for skin tissue engineering*. Materials Science and Engineering: C, 2014. **34**: p. 402-409.