

# Hot Start PCR Master Mix, lyophilized, 2×

LOT: See product label

EXPIRY DATE: See product label

### **ORDERING INFORMATION**

CAT. NO.	SIZE		PACKAGE CONTENT
BR0201103 200 rxn o		of 50 µl	4 × Lyo Hot Start PCR Master Mix
			$4 \times 1.25$ ml PCR Mix Reconstitution Buffer
COMPONENT		COMPOS	SITION
Lyo Hot Start PCR Mix	Master	Lyophilize	ed 2× Hot Start PCR Master Mix
PCR Mix Reconstitu Buffer	ution	Optimize	d PCR buffer for reconstituting Lyo Hot Start Master Mix
LYO HOT START F MASTER MIX RECONSTITUTION	PCR	1) Transfe Buffer to ( 2) Mix we 3) Store tl	r the whole content of one vial PCR Mix Reconstitution one vial Lyo Hot Start PCR Master Mix II – the lyophilisate will dissolve within seconds ne reconstituted Hot Start PCR Master Mix, 2× at -20°C
STORAGE		Store at ro label) Reconstite	com temperature or below (until expiry date – see product uted lyophilisate: store at $-20^{\circ}$ C for up to 12 months

## FEATURES

- Room-temperature stable enzymes and mixes
- Exceptionally pure Taq DNA Polymerase
- · Hot-start for high PCR specificity and sensitivity
- Optimized Master Mix for fast setup

## **APPLICATIONS**

- Ambient shipment and room-temperature storage
- Routine and demanding PCR applications
- PCR amplification up to 3 kb
- TA cloning

## DESCRIPTION

biotechrabbit<sup>™</sup> Lyo Hot Start PCR Master Mix is a freeze dried version of the well-established liquid equivalent. The stabilized format allows shipment and storage without cooling. The Master Mix is a perfect choice for a fast reaction setup that reduces the time required for calculation and pipetting and eliminates the need for buffer optimization. It is designed for low-background, high-throughput PCR of 0.2–3 kb DNA targets.

The 2x Hot Start PCR Master Mix contains pure biotechrabbit Hot Start *Taq* DNA Polymerase, extremely high-quality dNTPs and optimized PCR buffer; thus, only template, PCR primers and PCR-grade water are added.

The Hot Start *Taq* DNA Polymerase is inactive during reaction setup due to the bound antibody, which is quickly released at elevated temperatures, ensuring the enzyme is active only during PCR. There is no need for prolonged heating or denaturation steps. The hot start minimizes primer–dimers and mispriming.

Info: Recommended annealing temperature is 2°C above primer Tm (use gradient PCR to optimize the annealing temperature).

## PROTOCOL

#### Prevention of PCR contamination

When assembling the amplification reactions, care should be taken to eliminate the possibility of contamination with undesired DNA.

- Use separate clean areas for preparation of samples and reaction mixtures and for cycling.
- Wear fresh gloves. Use sterile tubes and pipette tips with aerosol filters for PCR setup.
- Use only water and reagents that are free of DNA and nucleases.
- With every PCR setup, perform a contamination control reaction that does not include template DNA.

#### Standard PCR setup

The standard PCR protocol using biotechrabbit reaction buffer provides excellent results for most applications. Optimization might be necessary for certain conditions, such as the amplification of long targets, high GC or AT content, strong template secondary structures or insufficient template purity. In such cases, optimization of template purification (see biotechrabbit nucleic acid purification kits), primer design and annealing temperature is recommended.

The best conditions for each primer-template can be optimized with the following:

- · Choosing the optimal quantities of template and primers
- Using a PCR Enhancer (i.e. BR1900201) for low amounts of template, impure or GC-rich templates
- Optimizing cycling conditions

## **BASIC PROTOCOL**

- The Master Mix is designed to be used without any optimization as it has all necessary reaction components in optimal amounts for successful PCR.
- Optionally, 5× PCR Enhancer (BR1900201) can be used to increase the yield and to lower the background in more complicated PCR reactions (low amounts of template, impure or GC-rich template).
- Thaw on ice and mix all reagents well. Keep all reagents and reactions on ice.
- Pipet the Master Mix into thin-walled 0.2 ml PCR tubes.
- Add template and primers separately if they are not used in all reactions.

COMPONENT	VOLUME	FINAL CONCENTRATION	
Hot Start PCR Master Mix, 2×	25	1×	
(reconstituted lyophilisate)	25 µi		
5× PCR Enhancer (optional)	10 µl	1×	
Forward primer	Variable	0.2–1 µM	
Reverse primer	Variable	0.2–1 µM	
Template DNA	Variable	10 pg–1 µg	
	Use 0.01–1 ng for plasmid or phage DNA and 0.1–1 µg for genomic		

DNA and 0.1-1 µg tol

Nuclease free water	Variable
Total volume	50 µl

- Mix and centrifuge briefly to collect the liquid in the bottom of the tube.
- Place in the PCR cycler.

## **CYCLING PROGRAM**

STEP	TEMPERATURE	TIME	CYCLES
Initial activation	95°C	2 min	1
Denaturation	95°C	30 s	25–35
Annealing*	(55-68°C)	15–30 s	25–35

\*Recommended annealing temperature is 2°C above Tm of primers, or use gradient PCR to optimize the annealing temperature

Extension	72°C	30–60 s/kb	25–35	
Final extension	72°C	5 min	1	
	To extend all incomplete PCR products			
Storage in the cycler	4°C	Indefinitely	1	

 Add loading dye solution (see DNA Loading Dye, 6×, cat. no. BR0800301) to the reactions to analyze PCR products on a gel or store them at −20°C.

## CERTIFICATE OF ANALYSIS

#### **Quality Control**

#### Functional assay

Human genomic DNA was amplified using the reconstituted Hot Start PCR Master Mix and specific primers to produce a distinct band.

Quality confirmed by: Head of Quality Control

## SAFETY INSTRUCTIONS

For safety instructions please see Safety Data Sheets (SDS)/Sicherheitshinweise finden Sie in den SDS unter:

http://www.biotechrabbit.com/support/documentation.html.

### **USEFUL HINTS**

- Visit Applications at www.biotechrabbit.com for more products and product selection guides.
- Most biotechrabbit products are available in custom formulations and bulk amounts.

## CONTACT BIOTECHRABBIT

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