

Anti-Common carp (Cyprinus carpio carpio) / Koi carp (Cyprinus carpio koi) IgM monoclonal antibody labelled with horseradish peroxidase

Product no: C16-HRP

# **Product Information**



#### **Product Description**

This monoclonal antibody (Mab) reacts with Common carp (*Cyprinus carpio carpio*) / Koi carp (*Cyprinus carpio koi*) immunoglobulin M (IgM) and has been conjugated to horseradish peroxidase. The Mab is of an IgG2b isotype and recognises the heavy chain of the IgM molecule.

### Use of product

The Mab is recommended for use in an Enzyme-Linked Immunosorbent Assay (ELISA) to measure antibody levels of antigen-induced IgM. The optimal conditions for use of this product will vary depending on the procedure used. The user must determine the suitability of the product for a particular procedure. This product is for *in vitro* use only.

#### Vial Contents

The Mab is prepared from bovine-free culture medium and contains no animal-derived stabilisers. Each vial contains sufficient conjugate for five 96-well ELISA plates.

The product should be reconstituted as follows:

Add 1 ml of phosphate buffered saline (PBS) (see buffers) to the vial to make a stock solution. For use transfer the contents of the vial into 54 ml of antibody buffer.



#### Storage

Store at -20°C prior to reconstitution. Once reconstituted, the stock solution can be stored at -20°C in 200  $\mu$ l aliquots and then added to 10.8 ml antibody buffer when required. Repeated freeze/thawing of the product should be avoided.

Protocol

Suggested protocol for the detection of Common carp and Koi carp IgM by indirect - Enzyme-Linked Immunosorbent Assay (ELISA) The method outlined here is only a guideline and assay conditions

may vary depending on the antigen used to coat the ELISA plate and environmental conditions.

#### Procedure

The coating procedure depends on the type of antigen used in the screening.

Plates coated with particulate antigens (e.g. bacteria)

- Coat 96-well ELISA plate with 0.05% (w/v) poly-L-lysine in coating buffer, 50 µl well<sup>-1</sup> for 60 min
- " Wash plate with 2 washes of low salt wash buffer
- Resuspend bacteria in PBS (1 x10<sup>6</sup> bacteria ml<sup>-1</sup>) and add to the wells of the ELISA plate at 100 µl well<sup>-1</sup>. Incubate overnight at 4°C or centrifuge plate at x 200 g for 5 min and incubate for 60 min at 22°C
- Add 50 µl well<sup>-1</sup> 0.05% (v/v) gluteraldehyde, diluted in PBS, to the antigen and incubate for a further 20 min at 22°C

Plates coated with soluble antigen (e.g. fish IgM)

Coat 96 well ELISA plate with 100 μl well-1 antigen [(1-20 μg ml-1) this will need to be optimised by the user] dissolved in coating buffer. Cover and incubate overnight a 4°C.

The remainder of procedure is as follows:

- " Wash plate 3 times with low salt wash buffer
- Post-coat plate (to block non-specific binding sites) with either 1% (w/v) bovine serum albumin (BSA) or 3% (w/v) casein (dried milk). Add 250 μl well<sup>-1</sup> and incubate for 2 h at 22°C
- " Wash plate with 3 washes of low salt wash buffer
- <sup>\*\*</sup> Prepare doubling-dilutions of the fish serum in PBS starting with a 1/2 dilution. Also use doubling-dilutions of pre-immune serum or serum from non-vaccinated/non-diseased fish, and PBS as negative controls. Add serum and control dilutions to the wells (100 µl well<sup>-1</sup>) and incubate for 3 h at 22°C or overnight at 4°C
- " Wash plate with 5 washes of high salt wash buffer, incubating for 5 min on last wash
- Add 100 μl well<sup>-1</sup> of the reconstituted anti-fish Mab-HRP and incubate for 60 min at 22°C
- " Wash plate with 5 washes of high salt wash buffer, incubating for 5 min on last wash
- " Add 100  $\mu l$  well-1 chromogen in substrate buffer and incubate for 10 min at 22°C
- Stop reaction with 50  $\mu$ l well<sup>-1</sup> of stop solution
- Read plate at 450 nm in an ELISA reader. Blank ELISA reader against wells filled with chromogen and stop solution. If an ELISA reader is unavailable compare colour with that of background (i.e. negative control).

NB. WEAR GLOVES WHEN USING CHROMOGEN AND STOP SOLUTION



 Buffers

 Coating buffer (Carbonate-bicarbonate solution)

 Na<sub>2</sub>CO<sub>3</sub>
 1.59 g

 NaHCO<sub>3</sub>
 2.93 g

Dissolve in one litre of distilled water. Adjust to pH 9.6. N.B. prepare fresh coating buffer on each occasion

 Phosphate Buffered Saline
 (PBS)

 0.02M Phosphate, 0.15M NaCl
 NaL

 NaH\_PO\_4.2H\_2O
 0.876g

 Na\_PHPO\_4.2H\_2O
 2.56g

 NaCl
 8.77g

Dissolve in one litre of distilled water. Adjust to pH 7.3 with conc. HCl

Wash buffer (x10)	(low salt)
Trisma base	24.2 g
NaCl	222.2 g
Merthiolate	1 g
Tween 20	5 ml

Dissolve in one litre of distilled water. Adjust to pH 7.3 with conc. HCl

Wash buffer(x10)(high salt)Trisma base24.2 gNaCl292.2 gMerthiolate1 gTween 2010 ml

Dissolve in one litre of distilled water. Adjust to pH 7.7 with conc. HCl

Antibody buffer Add 1 g of BSA to 100 ml of PBS (i.e. 1 % BSA solution)

<u>Conjugate buffer</u> Add 1g of BSA to 100 ml of low salt wash buffer

Substrate buffer(Sodium acetate/ citric acid buffer)Citric acid21.0 gSodium acetate8.2 g

Dissolve in one litre of distilled water. Adjust to pH 5.4 with 1 M NaOH Add 5  $\mu$ l of H<sub>2</sub>O<sub>2</sub> to 15 ml substrate buffer

<u>Substrate</u> Prepare 3'3'5'5'-Tetramethylbenzidine dihydrochloride (TMB) (42 mM) in 1:2 acetic acid: distilled water. Add 150  $\mu$ l of this solution to 15 ml substrate buffer

<u>Stop reagent</u> 2M  $H_2SO_4$  in distilled water



#### **Certificate of Analysis**

Anti-Common carp (*Cyprinus carpio carpio*) / Koi carp (*Cyprinus carpio koi*) monoclonal antibody labelled with horseradish peroxidase

Product no. C16-HRP

Batch no.

Date of expiry

## Absorbance of reconstituted conjugate measured by indirect ELISA:

The reconstituted conjugate gives an absorbance of \$at\$ 450 nm by ELISA when plates are coated with 10  $\mu$ g ml<sup>-1</sup> purified IgM.



Aquatic Diagnostic Ltd., Institute of Aquaculture, University of Stirling, Stirling, Scotland, FK9 4LA

Telephone: +44 (0)1786 466568 Fax: +44 (0)1786 472133 E-mail: aquaticdiagnostics@stir.ac.uk http://www.aquaticdiagnostics.com