

GFP-Expressing Human Brain Astrocytes
ORDER INFORMATION

Name of Cells: Human Brain Astrocyte (**GFP-HBAs**)
Catalogue Number: **cAP-0031GFP**
Product Format: Frozen Vial
Cell Number: >5 x 10⁵/vial

General Information

GFP expressing human brain astrocytes (GFP-HBAs) are selected from HBAs (**cAP-0031**) after transfected with GFP expressing lentiviral particles, which are resistant to Puromycin. The cells are shipped in frozen vials (the cells are provided @ passage 3). Astrocyte Growth Medium (cAP-29, containing FBS and growth factor supplements) is recommended for cell culture and these cells have an average population doubling capacity > 8 when cultured following the detailed protocol described below.

Characterization of the cells

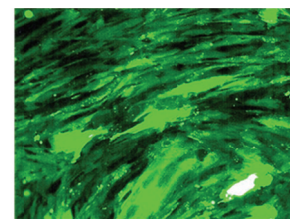
Cytoplasmic GFAP > 98% positive by immunofluorescence
 GFP-HBAs are negative for HIV-1, HBV, HCV, and mycoplasma.

Product Use: GFP-HBAs are for Research Use Only.

Shipping: Frozen Vial.

Handling of Arriving Cells

When you receive the cells in a frozen vial, you can transfer the vial of cells into a -80°C freezer for short period storage or a liquid nitrogen tank for long term storage. Thaw the cells in a 37°C water bath, and then transfer the cells in a T25 flask pre-coated with Quick coating solution (cAP-01) as described in details in Subculture Protocol.



cAP-0031 GFP GFP-Expressing Human Brain Human Astrocyte (HBAs)

Subculture Protocol

- Pre-coating of T25 flasks: Add 2ml of Quick Coating Solution (**cAP-01**) into one T25 flask and make sure whole surface of the flask is covered with the coating solution. Five minutes later, dispose excessive Quick Coating Solution by aspiration and the flask is ready to be used (no need for overnight incubation when using Quick Coating Solution). Other extracellular matrix can be used including gelatin, collagen, and fibronectin and you are advised to test the conditions for using those materials in advance.
- Rinse the cells in T25 flask with 5ml HBSS (**Room Temperature, RT**) twice.
- Add 2ml of Trypsin/EDTA (**RT**) (cAP-23) into one T25 flask (make sure the whole surface of the T25 flask is covered with Trypsin/EDTA), and gently dispose the excessive Trypsin/EDTA solution **within 20 seconds** with aspiration.
- Leave the T25 flask with the cells at **RT** for 1 minute (the cells usually will detach from the surface within 1-2 minutes). You can monitor the cells under microscope and when most of cells become rounded up, hit the flask against the bench surface, and the cells will move on the surface of the flask when monitoring under microscope.
- Add 5ml Trypsin Neutralization Buffer and spin the cells down with 800g for 5 minutes.
- Re-suspend the cell pellet with 10ml of Astrocyte Growth Medium (cAP-29) and the cell suspension is transferred directly into 2 pre-coated T25 flasks (5ml each, and the cells are sub-cultured at 1:2 ratios)
- Change medium every 2-3 days and cells usually become confluent within 7 days.

Related products

Quick Coating Solution	cAP-01	240ml	Angio-Proteomie
Astrocyte Growth Medium	cAP-29	500ml	Angio-Proteomie
Endothelial Basal Medium	cAP-03	500ml	Angio-Proteomie
HBSS w/o Ca ²⁺ , Mg ²⁺	cAP-11	100ml	Angio-Proteomie
Cell Freezing Solution (FBS)	cAP-22	50ml	Angio-Proteomie
Cell Freezing Solution (Non-FBS)	cAP-22B	50ml	Angio-Proteomie
Trypsin/EDTA Solution	cAP-23	100ml	Angio-Proteomie
Trypsin Neutralization Solution	cAP-28	100ml	Angio-Proteomie
ITS (100x)	cAP-26	10ml	Angio-Proteomie
L-Glutamine-MAXIMUM (100x)	cAP-27	100ml	Angio-Proteomie
Human Plasma Fibronectin Solution	cAP-42	1mg/ml	Angio-Proteomie

Caution: Handling human tissue derived products is potentially bio-hazardous. Although each cell strain is tested negative for HIV, HBV and HCV DNA, diagnostic tests are not necessarily 100% accurate; therefore, proper precautions must be taken to avoid inadvertent exposure. Always wear gloves and safety glasses when working these materials. Never mouth pipette. We recommend following the universal procedures for handling products of human origin as the minimum precaution against contamination.