

**GFP Expressing Human Dermal Fibroblasts-Adult**
**ORDER INFORMATION**

**Name of Cells:** GFP Expressing Human Dermal Fibroblasts (HDFCs-adGFP)  
**Catalogue Number:** **cAP-0008-adGFP**  
**Product Format:** Frozen Vial  
**Cell Number:** >5x 10<sup>5</sup>/vial

**General Information**

HDFCs-ad are isolated from normal adult skin tissue samples and transfected with GFP-Lentiviral particles at passage one. Puromycin resistant GFP-HDFCs-ad are selected and shipped in frozen vials (the cells are provided @ passage 3). Fibroblast Growth Medium (FGM, cAP-44, full medium) is recommended for cell culture and these cells have an average minimum population doubling levels > **14** when cultured following the detailed protocol described below).

HDFCs-ad are tested negative for HIV-1, HBV, HCV, and mycoplasma.

**Product Use: HDFCs-adGFP** cells are for research use only.

**Shipping:** Frozen Vials in dry ice package.

**Handling of Arriving Cells**

When you receive the cells in a frozen vial, you can transfer the vial of cells into a -80°C freezer for short term storage or a liquid nitrogen tank for long term storage. Thaw the cells in a 37°C water bath, and then transfer the cells into a T25 flask pre-coated with Quick coating solution (cAP-01) as described in details in Subculture Protocol.

**Subculture Protocol**

- A) Pre-coating of T25 flasks: Add 2ml of Quick Coating Solution (**cAP-01**) into one T25 flask and make sure whole surface of the flask is covered with the coating solution. Five minutes later, dispose excessive Quick Coating Solution by aspiration and the flask is ready to be used (no need for overnight incubation when using Quick Coating Solution). Other extracellular matrix can be used including gelatin, collagen, and fibronectin and you are advised to test the conditions for using those materials in advance.
- B) Rinse the cells in T25 flask with 5ml HBSS (**Room Temperature, RT**) twice.
- C) Add 2ml of Trypsin/EDTA (**RT**) (cAP-23) into one T25 flask (make sure the whole surface of the T25 flask is covered with Trypsin/EDTA), and gently dispose the excessive Trypsin/EDTA solution **within 20 seconds** with aspiration.
- D) Leave the T25 flask with the cells at **RT** for 1 minute (the cells usually will detach from the surface within 1-2 minutes). You can monitor the cells under microscope and when most of cells become rounded up, hit the flask against the bench surface, and the cells will move on the surface of the flask when monitoring under microscope.
- E) Add 5ml Trypsin Neutralization Buffer (cAP-28) and spin the cells down with 800g for 5 minutes.
- F) Re-suspend the cell pellet with 10 - 20ml of full medium and the cell suspension is transferred directly into 2 or 4 pre-coated T25 flasks (5ml each, and the cells are sub-cultured at 1:2 to 1: 4 ratios)
- G) Change medium every 2-3 days and cells usually become confluent within 7 days (when split at a 1:4 ratio).

**Related products**

Quick Coating Solution	cAP-01	240ml	Angio-Proteomie
Fibroblast Growth Medium	cAP-44	500ml	Angio-Proteomie
Fibroblast Basal Medium	cAP-44B	500ml	Angio-Proteomie
HBSS w/o Ca <sup>2+</sup> , Mg <sup>2+</sup>	cAP-11	100ml	Angio-Proteomie
Cell Freezing Solution (FBS)	cAP-22	50ml	Angio-Proteomie
Cell Freezing Solution (Non-FBS)	cAP-22B	50ml	Angio-Proteomie
Trypsin/EDTA Solution	cAP-23	100ml	Angio-Proteomie
Trypsin Neutralization Solution	cAP-28	100ml	Angio-Proteomie
ITS (100x)	cAP-26	10ml	Angio-Proteomie
L-Glutamine-MAXIMUM (100x)	cAP-27	100ml	Angio-Proteomie
Human Plasma Fibronectin Solution	cAP-42	1mg/ml	Angio-Proteomie

**Caution: Handling human tissue derived products is potentially bio-hazardous. Although each cell strain is tested negative for HIV, HBV and HCV DNA, diagnostic tests are not necessarily 100% accurate; therefore, proper precautions must be taken to avoid inadvertent exposure. Always wear gloves and safety glasses when working these materials. Never mouth pipette. We recommend following the universal procedures for handling products of human origin as the minimum precaution against contamination.**