



CM13 Interference-Resistant Helper Phage

INSTRUCTION MANUAL

CM13 Interference-Resistant Helper Phage for Phage Display

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Table of Contents

Description	4
Protocol	5
Overview	5
Procedures	5
Working with filamentous phage	5
Preparation of bacterial cells	5
Helper phage transduction	6
Troubleshooting	6
Appendix	7
MSDS Information	7
Quality Control	7
Technical Support	7
References	7

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Description

CM13 is a helper phage especially engineered for phage display. CM13 is a derivative of M13KO7 (1) containing the interference-resistant ir3B A->G mutation (2) at position 8418 of M13KO7. CM13-infected cells produce similar or higher levels of virions than M13KO7 while giving similar levels of display on the minor coat protein III. This helper phage makes preparation of virions particularly efficient at small scale, thus facilitating the overall screening process. This preparation contains enough helper phage to superinfect up to 1 L (1000 ml) of TG1 culture. Other bacterial strains may require different amounts of CM13 helper phage.

CM13 virions were isolated from the supernatant of infected *E. coli* TG1 cells, purified by PEG precipitation and resuspended in 50% glycerol TBS buffered. The suspension is in liquid state at -20°C.

Components

Product: Purified CM13 Helper Phage

Catalog number: PH020L

Quantity: 1 ml

Titer: 2.0×10^{12} pfu/ml

Storage conditions

Storage at -20°C is recommended.

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Protocol

Overview

The following guidelines are given for the production of virions displaying a polypeptide fused to the minor coat protein III of the M13 filamentous bacteriophage. For the production of single-stranded DNA using CM13, please consult reference (3).

This protocol is limited to the step of transduction (or superinfection) by CM13 helper phage of phagemid-containing bacteria. For additional methods and protocols such as purifying virions, quantifying virions, measuring titers, and/or preparation of basic media & solutions please consult **Antibody Design Labs** online technical resources at <http://www.abdesignlabs.com/technical-resources/>.

CM13 helper phage is suitable for the infection of F⁺ or F' *E. coli* bacteria containing a phagemid vector having an f1 or f1-like origin of replication. The protein III fusion is encoded by the phagemid vector while the helper phage brings all the necessary elements for efficient DNA packaging and assembly of virions.

The following protocol is given for the phagemid vector pADL-10b (**Antibody Design Labs** product number PD0105), an ampicillin-resistant phagemid with a full-length g3p fusion protein under the control of a *lac* promoter. Conditions are also optimized for single phage production in SS320 or TG1 *E. coli* strains in 2xYT medium and may require custom adjustments for different vectors, other strains, other growth media, and/or preparations of libraries.

Procedures

Working with filamentous phage

Keep the bench clean and regularly wiped with 2% bleach to limit phage cross-contamination and only use filtered tips to prevent aerosol contaminations. Phages are known to survive standard autoclaving conditions and are not removed by 0.22 µm filtration. Phages are either killed by heat-treating dry, autoclaved materials in an oven for 4 hours at 105°C (4) or by incubation in 2% bleach for at least 1 hour. We recommend to extensively wash with hot water all glass and plastic-ware, then submerge (tubes) or incubate (flasks) with a 2% solution of bleach for at least one hour. Heat-resistant glassware can then be autoclaved in an autoclave that is never used for biological waste while sensitive plastic-ware can be used directly or at best heat-treated as described above.

Preparation of bacterial cells

1. Pick a single colony from a freshly streaked plate with the phagemid-containing bacteria and inoculate a 3-ml culture of 2xYT medium supplemented with ampicillin 100 µg/ml and glucose 1% w/v. In absence of IPTG and presence of glucose 1% w/v, the expression of the g3p fusion protein is repressed; this helps preventing loss of phagemid or selection of insertless clones, and improves library integrity.
2. Incubate o/n at 37°C with agitation at 250 rpm.

3. In the morning, dilute an aliquot of the culture 1:20 v/v with fresh 2xYT medium in a new culture tube and incubate for one hour at 37°C with agitation at 250 rpm. It is important to dilute the glucose to 0.1% or less to prevent the catabolite repression of the *lac* operator.
4. Take a small aliquot in sterile conditions and measure the absorbance at 600 nm of a 1:10 dilution in 2xYT medium (A_{600}). With SS320 or TG1 strains, absorbance should be close to 0.5 OD and cells are ready to be transduced. If necessary adjust the length of incubation to your conditions.

Helper phage transduction

We recommend incubating directly at 37°C with agitation at 250 rpm for 30 min to one hour after adding the helper phage. The total number of transductants is not increased by either pre-incubating bacteria without agitation for 10 min prior to adding the helper phage (sometimes said to regenerate pili destroyed by strong agitation) or after adding the phage (sometimes said to promote transduction). In fact we have noted that in our conditions a lack of agitation during the transduction period results in smaller number of transductants and variable virion production. We also recommend adding the helper phage when the bacterial culture reaches an optical density at 600 nm between 0.4 OD and 0.5 OD; large amounts of non-superinfected cells may decrease virion production above 0.5 OD while disparities caused by differences in phage growth rates will be amplified at lower ODs.

1. Optionally vortex vigorously CM13 and collect with a brief microcentrifugation; this treatment will disrupt potential phage aggregates and is indicated when reproducible and precise quantities of helper phage are desired.
2. Add 1 μ l of CM13 phage per 1 ml of bacterial culture with A_{600} at 0.5 OD. This corresponds to a multiplicity of infection (MOI) around 14 and a number of pfu roughly 3 time to the number of bacteria in the culture medium.
3. Incubate for 1 h at 37°C and 250 rpm; some protocols only recommend 30 min incubation for this step.
4. Add ampicillin 100 μ g/ml, kanamycin 50 μ g/ml, and IPTG 200 μ M; lower the temperature to 30°C and incubate 8 h to overnight before proceeding to phage purification. The amount of IPTG to add depends on the phagemid, e.g. phagemids such as pADL-23c, pHEN2 or pComb3 do not require IPTG while pADL-10b does require IPTG.

Troubleshooting

The production of virions using helper phage is usually straightforward.

- ❖ Insure the presence of pili by applying selective conditions during the growth of bacterial stocks (streak TG1 bacteria on M9-minimal medium plates) or during the culture itself if maintenance of the *F'* episome is mediated by antibiotic resistance, e.g. tet^R phenotype for SS320 (add tetracycline 50 μ M to the growth medium).
- ❖ Streaking a kanamycin-containing plate before adding CM13 may reveal contamination by another helper phage.
- ❖ Lack or low virion production when bacterial cultures are too dense at the time of transduction. This problem is amplified by immunity to superinfection induced by endogenous expression of full-length g3p fusion protein by the phagemid. Make sure to only use bacterial cultures with A_{600} equal or inferior to 0.5 OD when adding the helper phage.
- ❖ Phagemid whose phage coat protein fusion expression is not properly regulated may lead to toxicity and arrest of cell division as soon as the glucose is removed from the culture medium. Therefore a culture not dense enough when the helper phage is added may produce very few virions. Always use freshly prepared bacteria and a culture to the recommend density value to maximize phage yields.

Appendix

MSDS Information

MSDSs (Material Safety Data Sheets) are available on **Antibody Design Labs** website at the corresponding product page.

Quality Control

Specifications are detailed on the online product page. **Antibody Design Labs** certifies that the product will perform according to these specifications.

Technical Support

Visit **Antibody Design Labs** website at www.abdesignlabs.com for technical resources, including manuals, vector maps and sequences, application notes, FAQs, etc.

FOR MORE INFORMATION OR TECHNICAL ASSISTANCE, CALL, WRITE, FAX, OR EMAIL US AT:

Antibody Design Labs

11175 Flintkote Ave, Suite B
San Diego, CA 92121

Email: support@abdesignlabs.com

Phone: 1-877-223-3104 (Toll Free)

Fax: 1-858-272-6007 (24 hour)

(Monday – Friday 9:00 AM – 5:00 PM PST)

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