



~The Kit for a new type smear preparation~

Smear Gell®

~ Please read this manual before use ~

Product Number: SG-01

Lot Number: See labels of each reagent tube

Product: Kit for solidifying various types of cell suspensions including cultured cells, blood cells, 3D culture cells and extremely small samples.

Shipment: Shipped frozen in dry ice (-20°C)

Storage: Store reagents (I-solution and II-solution) at -20°C immediately upon arrival.

Do not store in frost-free freezer with automatic thaw-freeze.

Components of Kit:

Reagents: I-solution (One tube - 25 μ × 4 for 40 tests including extra 4 μ L for trial)

II-solution (One tube – 60 μ L × 4 for 40 tests including extra 10 μ L for trial)

Accessories: 40 Slides

Product Manual

Warranty: One year after receiving the product.

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« Any inquiries about this product »

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§ 1. Preparation

1. Sample

1) To prepare cell suspension, re-suspend cells in small amount of cell culture medium or neutral buffer after centrifugation.

- * Fig. 1 is rough standard for concentrations of cell suspension. If the suspension looks to have enough conc. (approx. 4×10^3 cells/ μL), it is unnecessary to carry out centrifugation and re-suspension.
- * When the cell sample is fixated, centrifuge it and discard the fixative. After washing the cells with PBS, re-suspend with mediums or buffers.
Re-fix the smear preparation with an appropriate fixative, after solidifying the sample on slideglass.
- * When the cell suspension contains cryoprotectant like “Cell Banker” or Glycerol, wash the cells and re-suspend with mediums or buffers.

2) Prepare $3\mu\text{L}$ of cell suspension per one slideglass.

< Table-1 Volume of mixture (cell suspension and solutionI) required for each number of slide >

		Number of slideglass				
		one	two	three	five	ten
Use as mixture	Cell suspension	3 μL	6 μL	9 μL	15 μL	30 μL
	Solution I	2 μL	4 μL	6 μL	10 μL	20 μL

*Use $5\mu\text{L}$ of solutionII per one slideglass

2. Reagents and Others

1) Thaw reagents (solutionI, II) of the Smear Gell kit in hands, just before to use and keep them at room temperature.

- * SolutionI and II are able to repeat only 2 ~ 3 times FREEZE-THAW cycles.

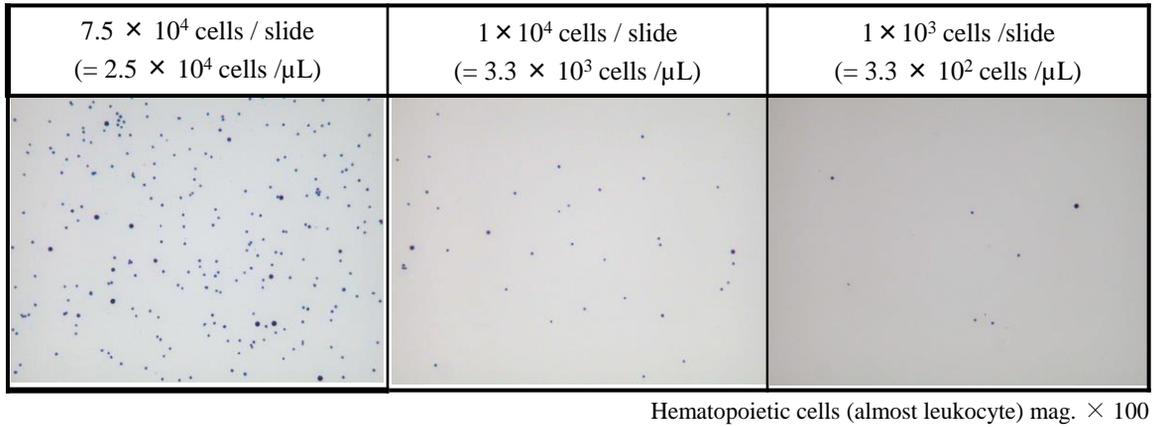
2) Prepare the APS coated slideglasses (MATSUNAMI).

- * If you use slideglasses of kit attachment, return them to room temperature before use.

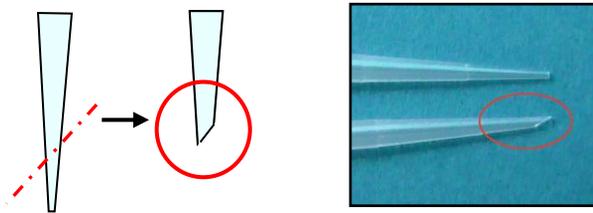
3) Prepare an appropriate fixative.

- * When you fix smear preparations using this kit on slideglasses, soak them whole in the fixative.
- * Choose appropriate fixative suitable for your staining method. DO NOT USE ACETONE for this kit.

< Fig. 1 Rough standard for concentration of cell suspension >

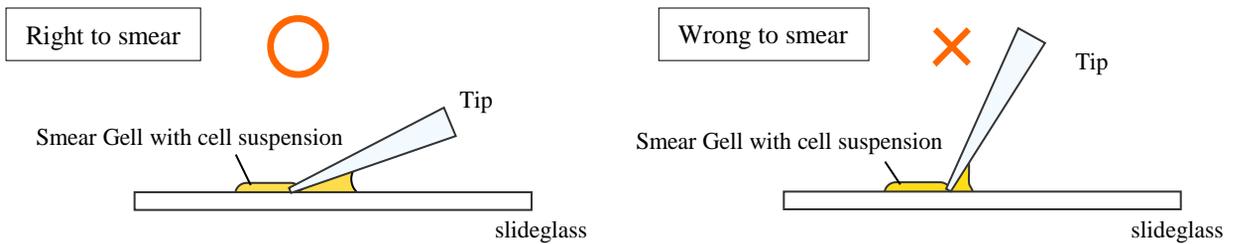


< Fig. 2 Tip >

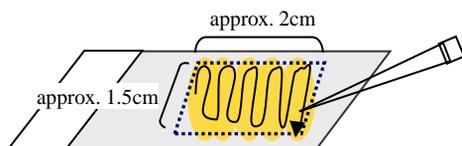


< Fig. 3 Point of smearing >

《SIDE VIEW》



《OVERHEAD VIEW》

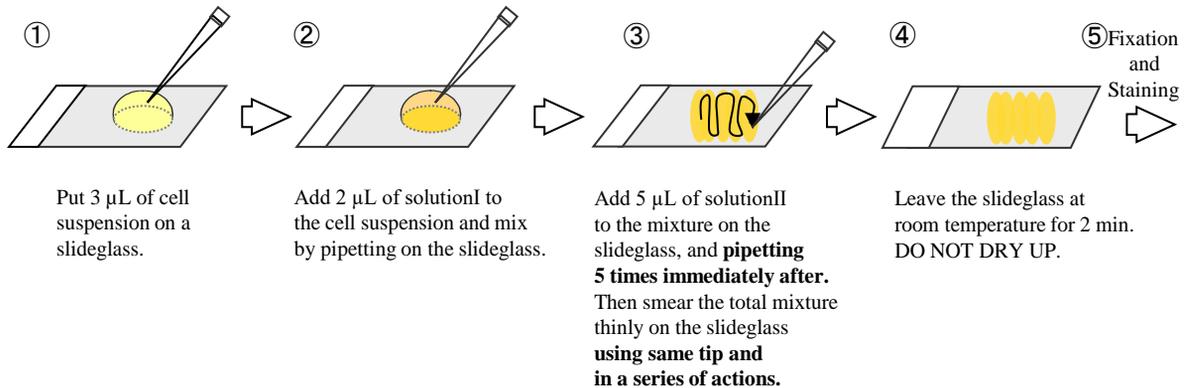


It is suitable for smearing at appropriate thickness to make the tip shuttle about 5 times in approx. 2cm \times 1.5cm area.
DO NOT CARRY OUT RE-SMEARING.
 Finish smearing in one action.

§ 2. Method of preparing smear preparation using Smear Gell

【Preparing only one slideglass at once】

< Schematic >



【Protocol】

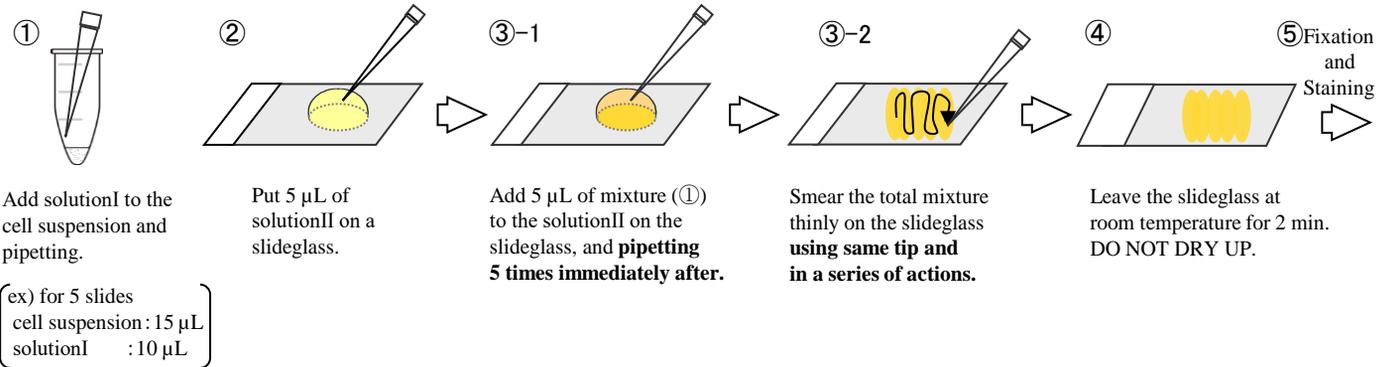
1. Put 3 μL of cell suspension on a slideglass.
2. Add 2 μL of solution I to the cell suspension and mix by pipetting on same slideglass.
3. Add 5 μL of solution II to the mixture on same slideglass, and pipetting 5 times immediately after, and furthermore smear the total mixture thinly with same tip, in a series of actions.

It is easier to smear that the tip is slanted like Fig. 3.

- * [Do not pipetting more than 5 times when you mix “mixture A” and solution II, and start smearing no sooner after finish 5 times pipetting.](#)
 - * If you feel difficulty in smearing with normal tips, cut the head of tips like Fig. 2 and try to use it in Step 3.
4. Leave the slideglass at room temperature for 2 minutes to solidify the gel. **AVOID DRYING.**
 - * Do not leave more than 2 minutes.
 5. Soak the slideglass in an appropriate fixative which suitable for your objective staining method.
 - * About 15 ~ 30 minutes fixation is enough.

【Preparing some slideglasses at once】

< Schematic >



【Protocol】

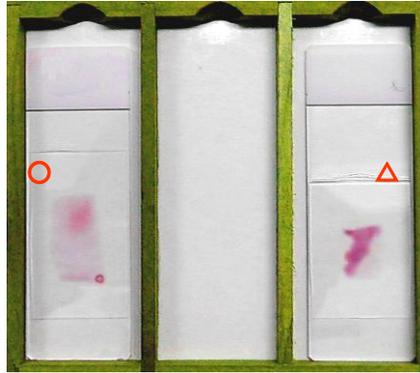
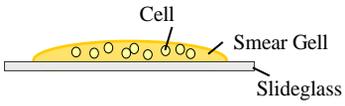
1. Add solution I to the cell suspension and pipetting well. (→ mixture A)
 - * See Table-1
2. Put 5 μL of solution II on a slideglass.
3. Add 5 μL of mixture A to solution II on the slideglass, and pipetting 5 times immediately after, and furthermore smear the total mixture thinly with same tip, in a series of actions.

It is easier to smear that the tip is slanted like Fig. 3.

 - * [Do not pipetting more than 5 times when you mix “mixture A” and solution II, and start smearing no sooner after finish 5 times pipetting.](#)
 - * If you feel difficulty in smearing with normal tips, cut the head of tips like Fig. 2 and try to use it in Step 3.
4. Leave the slideglass at room temperature for 2 minutes to solidify the gel. **AVOID DRYING.**
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5. Soak the slideglass in an appropriate fixative which suitable for your objective staining method.
 - * About 15 ~ 30 minutes fixation is enough.

<Fig. 4 Example of smear preparation using Smear Gell >

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- * Uniformly spreaded and appropriate thick Smear Gell containing cells.
 - * Cells were almost monolayer.
 - * It is easy to observe by a microscope.



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- * Smear Gell is likely to unstick due to its inequality.
 - * Cells were multi-layer.
 - * It seems difficult to observe by a microscope.

