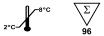


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Instructions for use

Serotonin Research ELISA ™







RUO

For research use only – Not for use in diagnostic procedures

Serotonin Research ELISA

1. Intended use and principle of the test

Ultra-sensitive Enzyme Immunoassay for the quantitative determination of Serotonin. Flexible test system for various biological sample types and volumes.

Serotonin is acylated and detected by the subsequent competitive ELISA kit, which uses the microtiter plate format. Antigens are bound to the solid phase of the microtiter plate. The acylated standards, controls and samples and the solid phase bound analytes compete for a fixed number of antibody binding sites. After the system is in equilibrium, free antigen and free antigen-antibody complexes are removed by washing. The antibody bound to the solid phase is detected by an anti-rabbit IgG-peroxidase conjugate using TMB as a substrate. The reaction is monitored at 450 nm.

Quantification of unknown samples is achieved by comparing their absorbance with a standard curve prepared with known standard concentrations.

2. Procedural Cautions, Guidelines and Warnings

- (1) This kit is intended for professional use only. Users should have a thorough understanding of this protocol for the successful use of this kit. Only the test instruction provided with the kit is valid and has to be used to run the assay. Reliable performance will only be attained by strict and careful adherence to the instructions provided.
- (2) The principles of Good Laboratory Practice (GLP) have to be followed.
- (3) In order to reduce exposure to potentially harmful substances, wear lab coats, disposable latex gloves and protective glasses where necessary.
- (4) All kit reagents and specimens should be brought to room temperature and mixed gently but thoroughly before use. Avoid repeated freezing and thawing of reagents and specimens.
- (5) For dilution or reconstitution purposes, use deionized, distilled, or ultra-pure water.
- (6) The microplate contains snap-off strips. Unused wells must be stored at 2 °C to 8 °C in the sealed foil pouch with desiccant and used in the frame provided.
- (7) Duplicate determination of sample is highly recommended to be able to identify potential pipetting errors.
- (8) Once the test has been started, all steps should be completed without interruption. Make sure that the required reagents, materials and devices are prepared ready at the appropriate time.
- (9) Incubation times do influence the results. All wells should be handled in the same order and time intervals.
- (10) To avoid cross-contamination of reagents, use new disposable pipette tips for dispensing each reagent, sample, standard and control.
- (11) A standard curve must be established for each run.
- (12) The controls should be included in each run and fall within established confidence limits. The confidence limits are listed in the QC-Report.
- (13) Do not mix kit components with different lot numbers within a test and do not use reagents beyond expiry date as shown on the kit labels.
- (14) Avoid contact with Stop Solution containing 0.25 M H_2SO_4 . It may cause skin irritation and burns. In case of contact with eyes or skin, rinse off immediately with water.
- (15) TMB substrate has an irritant effect on skin and mucosa. In case of possible contact, wash eyes with an abundant volume of water and skin with soap and abundant water. Wash contaminated objects before reusing them.
- (16) For information on hazardous substances included in the kit please refer to Safety Data Sheet (SDS). The Safety Data Sheet for this product is made available directly on the website of the manufacturer or upon request.
- (17) Kit reagents must be regarded as hazardous waste and disposed according to national regulations.

3. Storage and stability

Store the unopened reagents at 2 - 8 °C until expiration date. Do not use components beyond the expiry date indicated on the kit labels. Once opened the reagents are stable for 1 month when stored at 2 - 8 °C. Once the resealable pouch has been opened, care should be taken to close it tightly with desiccant again.

4. Materials

4.1 Contents of the kit

Standards and Controls - Concentrated*

Cat. no.	Component	Colour/Cap		ntration nmol/l		I tration pmol/sample vol.	Volume/ Vial
BA R-8901	STANDARD A	white	0	0	0	0	4 ml
BA R-8902	STANDARD B	light yellow	0.015	0.085	1.5	8.5	4 ml
BA R-8903	STANDARD C	orange	0.05	0.28	5	28.4	4 ml
BA R-8904	STANDARD D	dark blue	0.15	0.85	15	85	4 ml
BA R-8905	STANDARD E	light grey	0.5	2.8	50	284	4 ml
BA R-8906	STANDARD F	black	2.5	14	250	1 418	4 ml
BA R-8951	CONTROL 1 light green Refer to QC-Report for expected value and 4 ml						
BA R-8952	CONTROL 2 dark red acceptable range! 4 ml						
Conversion:	Serotonin (ng/ml) x 5.67 = Serotonin (nmol/l) Serotonin (pg/sample volume) x 5.67 = Serotonin (pmol/sample volume)						
Content:						ed quantity of se	
*Standards and controls have to be diluted 1+1000 prior to use (please see chapter 6.1 for further information). The shown concentrations apply to diluted standards and controls (1+1000) when 100 μl of undiluted sample is used. Please refer to chapter 7 for the use of a correction factor in case of using diluted samples and/or less sample volume.							
BA E-5934	ACYL-PLATE	Acylation	Plate - F	Ready to	use		
Content:	1 x 96 well plate, pre-coated with acylation reagent						
BA E-5937	STABILIZER	Stabilizer	- Ready	to use			
Content:	Stabilizing agent, 10% solution						
Volume:	1 x 4 ml/vial, purple cap						
BA E-5941	DIL-CONC 20x Dilution Concentrate – Concentrated 20x						
Content:	TRIS buffer with 1% stabilizing agent and a non-mercury preservative						
Volume:	1 x 50 ml/vial, white cap						
BA D-0090	FOILS	Adhesive	Foil - Re	eady to us	se		
Content:	Adhesive Foils in a resealable pouch						
Volume:	1 x 4 foils						
BA E-0030	WASH-CONC 50>	Wash Buff	er Conc	entrate ·	- Concentrated	50x	
Content:		non-ionic dete					
Volume:		l, light purple o	-	. ,	5		
BA E-0040	CONJUGATE	Enzyme C	oniugat	: e - Read	v to use		
Content:		-			with peroxidase	2	
Volume:	1 x 12 ml/via	l, red cap			·		
BA E-0055	SUBSTRATE	Substrate	- Readv	to use			
Content:			-		lbenzidine, sub	strate buffer and	l hydrogen
	peroxide		, j	· · · ,	· · · · , · · ·		, <u>5</u> -
Volume:	1 x 12 ml/via	l, black cap					
BA E-0080	STOP-SOLN	Stop Solu	tion - Re	eady to u	se		
Content:	0.25 M sulfu	ric acid					
Volume:	1 x 12 ml/via	al, light grey ca	р				
Hazards identification:							
		corrosive to m	otals				

H290 May be corrosive to metals. H314 Causes severe skin burns and eye damage.

BA E-5910	SER-AS	Serotonin Antiserum - Ready to use	
Content:	Rabbit anti-serotonin antibody, blue coloured		
Volume:	1 x 3 ml/vial, blue cap		
BA E-5911	ACYL-BUFF	Acylation Buffer - Ready to use	
Content:	TRIS buffer wit	h non-mercury preservative	
Volume:	1 x 4 ml/vial, y	ellow cap	
BA E-0931	III SER 5-HIAA	Serotonin Microtiter Strips - Ready to use	

Content: 1 x 96 well (12x8) antigen precoated microwell plate in a resealable pouch with desiccant

4.2 Additional materials and equipment required but not provided in the kit

- Calibrated precision pipettes to dispense volumes between 1 100 µl; 10 ml
- Microtiter plate washing device (manual, semi-automated or automated)
- ELISA reader capable of reading absorbance at 450 nm and if possible 620 650 nm
- Plate shaker (shaking amplitude 3 mm; approx. 600 rpm)
- Absorbent material (paper towel)
- Water (deionized, distilled, or ultra-pure)
- Vortex mixer

5. Sample collection and storage

In general this assay is dedicated for any biological sample such as serum, urine, platelets, platelet-poor plasma, tissue homogenates, dialysates and other samples.

Storage: up to 6 hours at 2 – 8 °C, for longer periods (up to 6 months) at -20 °C. Avoid exposure to direct sunlight.

Serum

Collect blood by venipuncture (Monovette[™] or Vacuette[™] for serum), allow to clot, and separate serum by centrifugation according to manufacturer's instructions at room temperature. Do not centrifuge before complete clotting has occurred. Patients receiving anticoagulant therapy may require increased clotting time. Haemolytic and especially lipemic samples should not be used for the assay.

Urine

Spontaneous or 24-hour urine, collected in a bottle containing 10 – 15 ml of 6 M HCl, should be used. Determine the total volume of urine excreted during a period of 24 h for calculation of the results.

Platelets

More than 98 percent of the circulating serotonin is located in the platelets and is released during blood clotting. Blood must be collected according to manufacturer's instructions by venipuncture in plastic tubes (Monovette[™]) or Vacuette[™]) containing EDTA or Citrate as anticoagulant.

To obtain platelet-rich plasma (PRP) the samples are centrifuged for 10 minutes at room temperature (200 x g). Transfer the supernatant to another tube and count the platelets.

The platelet pellet is obtained by adding 800 μ l of physiological saline to 200 μ l of PRP (containing between 350,000 – 500,000 platelets/ μ l) and centrifugation (4,500 x g, 10 minutes at 4 °C). The supernatant is then discarded.

200 μ l of water (deionized, distilled, or ultra-pure) is added to the pellet and mixed thoroughly on a vortex mixer. This suspension can be stored frozen for several weeks at < -20 °C.

After thawing of the frozen samples, centrifuge at 10,000 x g for 2 minutes at room temperature.

To protect Serotonin against oxidative degradation the samples should contain 1% Stabilizer REF BA E-5937.

6. Test procedure

Allow all reagents and samples to reach room temperature and mix thoroughly by gentle inversion before use. Duplicate determinations are recommended. It is recommended to number the strips of the microwell plate before usage to avoid any mix-up.

6.1 Sample preparation

The Serotonin Research ELISA is a flexible high sensitive test system for various biological sample types and sizes. It is not possible to give a general advice how to prepare the samples. However, the following basics should help the researcher to fit the protocol to his specific needs.

- Using poorly diluted or undiluted samples might lead to incorrect results due to a matrix effect. Therefore it is advisable to perform a linearity experiment prior to the test. Make different dilutions of a sample with the included diluent (e.g. 1:1, 1:4, 1:10, 1:20 and so on), spike each dilution with the same known concentration and check the recovery. If the samples are found consistently correct and no matrix effect is detected, samples can be used undiluted.
- If a matrix effect is detected, samples should be diluted with the included diluent prior to the test. It is also possible to dilute the standards with the sample matrix instead of diluent, in order to create the same matrix conditions for standards and samples. Currently the following buffers/solvents are evaluated for use: Ringer Buffer, PBS and 0.9% NaCl. If another substance is used, please check the compatibility by a Proof of Principle prior to the measurement. Prepare a stock solution of serotonin or use standard F. Add small amounts (to change the native sample matrix as little as possible) of the stock solution or standard F to the sample matrix and check the recovery. Please take the correction factor into account (see chapter 7).
- If the expected sample concentrations are higher than the standard range (see chapter 4.1) samples should be diluted accordingly with the included diluent. Please take the correction factor into account (see chapter 7).
- The measuring range and sensitivity of this test are defined by the correction factor, which is calculated by sample volume and dilution (see chapter 7). If the expected concentrations are unknown, please test different dilutions and amounts of sample volume, to make sure that the samples will fall into the measuring range of this assay.
- Serotonin decomposes fast in acidic solution (< pH 3) and at higher temperatures.
- When acidic sample solutions are used, protect serotonin by keeping the temperature low (2 -8 °C). Use pre-cooled buffers and materials. Adjust the pH to (6 7.4) as soon as possible.
- A pH 7 8.5 during acylation (see step 6.3.4) is mandatory.
- To protect serotonin against oxidative degradation add Stabilizer REF BA E-5937 (refer to chapter 5).

If you need any support in establishing a protocol for your specific purposes, do not hesitate to contact the manufacturer directly!

6.2 Preparation of reagents

Diluent

Dilute the 50 ml Dilution Concentrate with water (deionized, distilled, or ultra-pure) to a final volume of 1000 ml.

The Diluent (Diluted Dilution Concentrate) contains 1% Stabilizer REF BA E-5937. Storage: 1 month at 2 – 8 °C

Wash Buffer

Dilute the 20 ml Wash Buffer Concentrate with water (deionized, distilled, or ultra-pure) to a final volume of 1000 ml.

Storage: 1 month at 2 – 8 °C

Standards and controls

The standards and controls have to be diluted <u>freshly</u> 1 + 1000 with Diluent or buffer^{*1}, for example: 10μ standard + 10 ml Diluent or buffer^{*1}.

 $*^{1}$ The buffer used for the respective experiment, enriched with 1% Stabilizer REF BA E-5937.

Serotonin Microtiter Strips

In rare cases residues of the blocking and stabilizing reagent can be seen in the wells as small, white dots or lines. These residues do not influence the quality of the product.

6.3 Acylation

Pipette 100 μl of diluted standards, diluted controls, and 1 – 100 μl of samples into the respective wells of the Acylation Plate^{*2}.

2. Add **Diluent** or buffer* (refer to 6.1) to the wells containing the samples to a **final volume** of **100 μl**.

3. Add **25 µl Acylation Buffer** to all tubes.

4. Acylate for **30 min** at **RT** (20 – 25 °C) on a **shaker** (approx. 600 rpm).

100 µl of the acylated standards, controls and samples are needed for the subsequent ELISA

*²⁾ The wells of the Acylation Plate are covered by plastic bars which have to be removed prior to use.

6.4 Serotonin ELISA

- **1.** Pipette **100 μl** of the **acylated standards, controls and samples** into the appropriate wells of the **Serotonin/5-HIAA Microtiter Strips.**
- 2. Pipette 25 µl of the Serotonin Antiserum into all wells.
- 3. Cover plate with Adhesive Foil and incubate for 15 20 h at 2 8 °C.
- Remove the foil. Discard or aspirate the content of the wells. Wash the plate 3 x by adding 300 μl of Wash Buffer, discarding the content and blotting dry each time by tapping the inverted plate on absorbent material.
- 5. Pipette 100 µl of the Enzyme Conjugate into all wells.
- 6. Incubate for **30 min** at **RT** (20 25 °C) on a **shaker** (approx. 600 rpm).
- Discard or aspirate the content of the wells. Wash the plate 3 x by adding 300 µl of Wash Buffer, discarding the content and blotting dry each time by tapping the inverted plate on absorbent material.
- 8. Pipette 100 μl of the Substrate into all wells and incubate for 20 30 min at RT (20 25 °C) on a shaker (approx. 600 rpm). Avoid exposure to direct sunlight!
- **9.** Add **100** µl of the **Stop Solution** to each well and shake the microtiter plate to ensure a homogeneous distribution of the solution.
- **10. Read** the absorbance of the solution in the wells within 10 minutes, using a microplate reader set to **450 nm** (if possible a reference wavelength between 620 nm and 650 nm is recommended).

7. Calculation of results

The calibration curve from which the concentrations of the samples can be read off, is obtained by plotting the absorbance readings (calculate the mean absorbance) measured for the standards (linear, y-axis) against the corresponding standard concentrations (logarithmic, x-axis).

The use of a non-linear regression for curve fitting (e.g. spline, 4- parameter, akima) is recommended.



This assay is a competitive assay. This means: the OD-values are decreasing with increasing concentrations of the analyte. OD-values found below the standard curve correspond to high concentrations of the analyte in the sample and have to be reported as being positive.

The concentrations of the samples taken from the standard curve have to be multiplied by a correction factor:

	100 µl (volume of standards)
Correction factor =	sample volume (µl)

Three different examples are shown to illustrate the calculation of possible correction factors:

Example 1)

10 μ l of the undiluted sample are acylated and the concentration taken from the standard curve is 0.02 ng/ml serotonin.

Correction factor = 100/10 = 10Final concentration of the sample = 0.02 ng/ml x 10 = 0.2 ng/ml serotonin

Example 2)

100 μ l of the 1:100 prediluted sample are acylated and the concentration taken from the standard curve is 0.02 ng/ml serotonin. Correction factor = 100

Final concentration of the sample = $0.02 \text{ ng/ml} \times 100 = 2.0 \text{ ng/ml}$ serotonin

Example 3)

10 µl of the 1:100 prediluted sample are acylated and the concentration taken from the standard curve is 0.02 ng/ml serotonin. Correction factor = $100 \times (100/10) = 1000$

Final concentration of the sample = $0.02 \text{ ng/ml} \times 1000 = 20 \text{ ng/ml}$ serotonin

Conversion

Serotonin (pg/ml) x 5.67 = Serotonin (pmol/l) Serotonin (pg/sample volume) x 5.67 = Serotonin (pmol/sample volume)

7.1 Quality control

The confidence limits of the kit controls are listed in the QC-Report.

8. Assay characteristics

8.1 Sensitivity

	Serotonin		
Sensitivity	0.005 ng/ml x C*		

C* = correction factor (refer to 7.)

8.2 Specificity

Compound	Cross-reactivity (%)		
Serotonin	100		
Tryptamine	0.19		
Melatonin	0.03		
5-Hydroxyindole acetic acid	<0.002		
Phenylalanine	<0.002		
Histidine	<0.002		
Tyramine	<0.002		
5-Hydroxytryptophan	<0.002		

For literature or any other information please contact your local supplier.

The liability of the manufacturer shall be limited to the replacement of defective products. The manufacturer takes no liability for any damages or expenses arising directly or indirectly from the use of this product.

Symbols:

+2 +2	Storage temperature	~~~	Manufacturer	Σ	Contains sufficient for <n> tests</n>
52	Expiry date	LOT	Batch code	I V D	For in-vitro diagnostic use only!
i	Consult instructions for use	CONT	Content	CE	CE labelled
Â	Caution	REF	Catalogue number	RUO	For research use only!