pluriSelect

Protocol for the use of PBMC 24+ Spin Medium pre-filled pluriMate® Tubes

pluriMate® - Specification

	pluriMate - 2 ml, pre-filled	pluriMate - 15 ml, pre-filled	pluriMate - 50 ml, pre-filled
Order No. 50 pcs.	44-09302-10	44-09315-10	44-09350-10
Order No. 100 pcs.	44-09302-11	44-09315-11	44-09350-11
Order No. 500 pcs.	44- 09302-15	44-09315-15	44-09350-15

Product Description The pluriMate® centrifugation tubes pre-filled with PBMC 24+ Spin Medium® can be used for an optimal separation peripheral blood mononuclear cells (PBMCs) in high yield from >6h whole blood and buffy coat by a simple centrifugation procedure. The key feature of pluriMate® is the porous sponge. This barrier prevents you from time-consuming and laborious overlaying of the sample material. Anticoagulated blood or buffy coat can simply be poured directly from the blood sampling tube into the pluriMate® tube. The porous barrier prevents mixture of the sample material with the separation medium. When separation is complete, the barrier prevents recontamination of the enriched cell fraction during harvest.

Pre-filled with PBMC 24+ Spin Medium® (Catalog 60-00093-10)

Enrichment of Peripheral Blood Mononuclear Cells (PBMC)

Age of blood older than 6 hours

Directions for the use of the pluriMate® Tube

1. Check that recommended medium, blood sample, density gradient medium and centrifuge are all at room temperature.

Preparation of the pluriMate® Tube

2. Centrifuge at 1000 x g for 10 sec. and leave 3 - 5 mm supernatent on top.

Add Sample Material

3. Fill in sample material on top of sponge (Fig. a).
Note: To reduce platelet contamination you can add pluriSpin® PLT Depletion (Order No. 19-00002-31)

	pluriMate® 2 ml	pluriMate® 15 ml	pluriMate® 50 ml
Sample material vol.	0.25 - 1 ml	2 - 7 ml	5 - 30 ml

Spin

4. Centrifuge for 15 minutes at 800 x g at room temperature with in a swing bukket rotor and the **brake on.**

Collect

5. Remove plasma by pipetting until white cell layer (Fig. d).

6. Collect cells in the white layer in a fresh tube (Fig. e).

Wash

7. Fill up reaction tube with wash buffer.

8. Spin down cells 10 minutes with 300 x g (no or small brake) at 4°C.

9. Pour out supernatant, leave the reaction tube on the table for 20 sec. Wash buffer excess will run down from the tube wall and collect at the bottom.

10. Aspirate most of the liquid above the pellet. The liquid will look foggy, these are mostly platelets – aspiration will improve washing result.

11. Reconstitute pellet with 1 ml of wash buffer by carefully pipetting.

12. Repeat steps 7 to 10.

13. Reconstitute pellet at your desired volume.

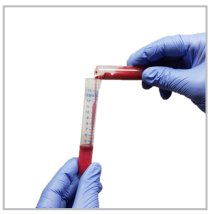
Fig. a - Fill in sample material


Fig. b - Before centrifugation

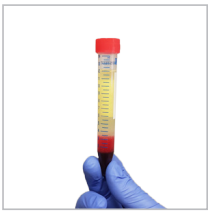
Fig. c - After centrifugation


Fig. d - Remove plasma

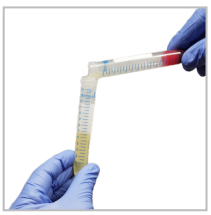
Fig. e - Collect cells

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