

Manual

# IDKmonitor<sup>®</sup> Ustekinumab free ADA ELISA

For the determination of free human antibodies against ustekinumab (e.g. Stelara®) in EDTA plasma and serum

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#### 1. INTENDED USE

This enzyme-linked immunosorbent assay (ELISA) kit is intended for the determination of free anti-drug antibodies (ADA) against the therapeutic antibody ustekinumab (e.g. Stelara<sup>®</sup>) in EDTA plasma and serum. For research use only. Not for use in diagnostic procedures.

## 2. INTRODUCTION

Ustekinumab is a monoclonal, human therapeutic antibody targeting p40, the common subunit of interleukin 12 and 23 (IL-12/23). The *IDK* monitor<sup>®</sup> ustekinumab free ADA ELISA for the detection of antibodies against ustekinumab (e. g. Stelara<sup>®</sup>) measures free anti- ustekinumab antibodies.

## 3. MATERIAL SUPPLIED

Cat. No.	Label	Kit components	Quantity
KR9666	PLATE	Microtiter plate, pre-coated with (F(ab) <sub>2</sub> )	12 x 8 wells
KR0001.C.100	WASHBUF	Wash buffer concentrate, 10x	1 x 100 ml
KR9666	CONJ	ONJ Conjugate concentrate (therapy antibody), peroxidase-labelled	
KR9666	CTRL NEG	Negative control, lyophilised (see specification for range)	4 x 1 vial
KR9666	CTRL POS	Positive control, lyophilised (see specification for range)	4 x 1 vial
KR9666	CTRL CUT OFF	Cut-off control, lyophilised	4 x 1 vial
KR9666	SAMPLEBUF	Sample dilution buffer, ready-to-use	1 x 30 ml
KR0002.15	SUB	Substrate (tetramethylbenzidine), ready-to-use	1 x 15 ml
KR0003.15	STOP	Stop solution, ready-to-use	1 x 15 ml

For reorders of single components, use the catalogue number followed by the label as product number.

#### 4. MATERIAL REQUIRED BUT NOT SUPPLIED

- Ultrapure water\*
- Calibrated precision pipettors and 10–1000 µl single-use tips
- Foil to cover the microtiter plate
- · Horizontal microtiter plate shaker
- · Multi-channel pipets or repeater pipets
- Centrifuge
- Vortex
- Standard single-use laboratory glass or plastic vials, cups, etc.
- Microtiter plate reader (required filters see chapter 7) \* Immundiagnostik AG recommends the use of ultrapure water (water type 1; ISO 3696), which is free of undissolved and colloidal ions and organic molecules (free of particles > 0.2  $\mu$ m) with an electrical conductivity of 0.055  $\mu$ S/cm at 25 °C ( $\geq$ 18.2 M $\Omega$ cm).

## 5. PREPARATION AND STORAGE OF REAGENTS

- To run the assay more than once, ensure that reagents are stored at conditions stated on the label. **Prepare only the appropriate amount necessary for each run.** The kit can be used up to 4 times within the expiry date stated on the label.
- Reagents with a volume less than  $100\,\mu l$  should be centrifuged before use to avoid loss of volume.
- Preparation of the wash buffer: The wash buffer concentrate (WASHBUF) has to be diluted with ultrapure water 1:10 before use (100 ml WASHBUF + 900 ml ultrapure water), mix well. Crystals could occur due to high salt concentration in the concentrate. Before dilution, the crystals have to be redissolved at room temperature or in a water bath at 37 °C. The WASHBUF is stable at 2–8 °C until the expiry date stated on the label. Wash buffer (1:10 diluted WASHBUF) can be stored in a closed flask at 2–8 °C for 1 month.
- The lyophilised controls (CTRL NEG, CTRL POS and CTRL CUT-OFF) are stable at 2–8 °C until the expiry date stated on the label. Reconstitution details are given in the specification data sheet. Controls (reconstituted CTRL CUT-OFF, CTRL POS and CTRL NEG) are not stable and cannot be stored.
- Preparation of the conjugate: Before use, the conjugate concentrate (CONJ) has to be diluted 1:101 in wash buffer (100 µl CONJ + 10 ml wash buffer). The CONJ is stable at 2–8 °C until the expiry date stated on the label. Conjugate (1:101 diluted CONJ) is not stable and cannot be stored.

• All other test reagents are ready-to-use. Test reagents are stable until the expiry date (see label) when stored at **2–8°C**.

#### 6. SPECIMEN COLLECTION AND PREPARATION

#### EDTA plasma and serum

EDTA plasma or serum samples must be diluted 1:10 before performing the assay,

e.g. **25 µl** sample + **225 µl** SAMPLEBUF (sample buffer), mix well.

For testing in duplicates, pipet **2 x 100 µl** of each diluted sample.

#### Sample storage

Freshly collected EDTA plasma or serum can be stored for one day at room temperature  $(15-30 \degree C)$  or for longer storage at  $-20 \degree C$ .

## 7. ASSAY PROCEDURE

#### Principle of the test

This enzyme immunoassay is a sandwich assay for the determination of free antibodies against ustekinumab (e.g. Stelara<sup>®</sup>). In a first incubation step, the free anti-therapeutic antibodies from the sample are bound to the ustekinumab F(ab)<sub>2</sub> fragments coated on the plate. To remove all unbound substances, a washing step is carried out. In a further incubation step, peroxidase-labelled ustekinumab is added. After another washing step, to remove all unbound substances, the solid phase is incubated with the substrate, tetramethylbenzidine (TMB). An acidic stop solution is then added. The colour converts to yellow. The absorbance of the colour compound is determined photometrically. The intensity of the colour is directly proportional to the amount of bound ADAs (here: anti-ustekinumab antibodies) from the sample. The results are evaluated by a cut-off control.

#### Test procedure

Bring all reagents and samples to room temperature (15–30 °C) and mix well.

Mark the positions of controls/samples on a protocol sheet.

Take as many microtiter strips as needed from the kit. Store unused strips together with the desiccant bag in the closed aluminium packaging at 2-8 °C. Strips are stable until expiry date stated on the label.

For automated ELISA processors, the given protocol may need to be adjusted according to the specific features of the respective automated platform. For further details please contact your supplier or Immundiagnostik AG.

We recommend to carry out the tests in duplicate.

1.	<b>Before use</b> , wash the wells <b>5 times</b> with <b>250 µl wash buffer</b> . After the final washing step, remove residual wash buffer by firmly tapping the plate on absorbent paper.	
2.	Add each <b>100 µl controls/diluted samples</b> into the respective wells.	
3.	Cover the strips and incubate for <b>1 hour</b> at room temperature (15–30 °C) on a <b>horizontal shaker</b> at 550 rpm with an orbit of 2 mm.	
4.	Discard the content of each well and wash <b>5 times</b> with <b>250 µl wash buffer</b> . After the final washing step, remove residual wash buffer by firmly tapping the plate on absorbent paper.	
5.	Add <b>100 µl conjugate</b> (diluted CONJ) into each well.	
6.	Cover the strips and incubate for <b>1 hour</b> at room temperature (15–30 °C) on a <b>horizontal shaker</b> at 550 rpm with an orbit of 2 mm.	
7.	Discard the content of each well and wash <b>5 times</b> with <b>250 µl wash buffer</b> . After the final washing step, remove residual wash buffer by firmly tapping the plate on absorbent paper.	
8.	Add <b>100 µl substrate</b> (SUB) into each well.	
9.	Incubate for <b>10–20 minutes</b> * at room temperature (15–30°C) in the <b>dark</b> .	
10.	Add <b>100 µl stop solution</b> (STOP) and mix well.	
11.	Determine <b>absorption immediately</b> with an ELISA reader at <b>450 nm</b> against 620 nm (or 690 nm) as a reference. If no reference wavelength is available, read only at 450 nm. If the extinction of the highest standard exceeds the range of the photometer, absorption must be measured immediately at <b>405 nm</b> against 620 nm as a reference.	

\* The intensity of the colour change is temperature sensitive. We recommend observing the colour change and stopping the reaction upon good differentiation.

## 8. RESULTS

The analysis of the results is done using the cut-off control. Samples with a higher optical density (OD) as the OD of the cut-off control are positive. Samples with an OD lower than the OD of the cut-off control are negative.

#### Cut-off = 10 AU/ml = OD of cut-off control

For the calculation of the sample concentrations, linear regression using a linear ordinate and abscissa is recommended.

The plausibility of the pairs of values should be examined before the automatic evaluation of the results. If this option is not available with the used program, a control of the paired values should be done manually.

#### Sample calculation for a positive sample

average sample OD	0.735
average cut-off contro OD	0.065= 10 AU/ml
Concentration of the sample	$\frac{0.735 \text{ x } 10 \text{ AU/ml}}{0.065} = 113 \text{ AU/ml}$

## 9. LIMITATIONS

The lower limit of the measurement range is the LoB.

LoB see chapter "Performance Characteristics".

Samples with concentrations lower than the measurement range cannot be clearly quantified.

## **10. QUALITY CONTROL**

Immundiagnostik AG recommends the use of external controls for internal quality control, if possible.

Provided control samples should be analysed with each run. Results, generated from the analysis of control samples, should be evaluated for acceptability using appropriate statistical methods. The results for the samples may not be valid, if within the same assay one or more values of the quality control sample are outside the acceptable limits.

#### Reference range

We recommend each laboratory to establish its own reference range.

## **11. PERFORMANCE CHARACTERISTICS**

#### Accuracy – Precision

#### Repeatability (Intra-Assay); n=20

The repeatability was assessed with 2 serum samples under constant parameters (same operator, measurement system, day and kit lot).

Sample	Mean value [AU/ml]	<b>CV</b> [%]
1	25.94	3.7
2	25.92	2.2

#### Reproducibility (Inter-Assay); n=21

The reproducibility was assessed with 2 serum samples under varying parameters (different operators, measurement systems, days and kit lots).

Sample	Mean value [AU/ml]	<b>CV</b> [%]
1	25.74	8.4
2	17.07	6.2

## Analytical sensitivity

The following values have been estimated based on the concentrations of the standards without considering possibly used sample dilution factors.

Limit of blank, LoB

4.75 AU/ml

## Analytical specificity

The specificity of the antibody was tested by measuring the cross-reactivity against against the following therapeutic antibodies: anti-adalimumab, anti-infliximab, anti-etanercept, anti-golimumab and anti-vedolizumab. No cross-reactivity was observed.

## **12. PRECAUTIONS**

- For research use only.
- Human materials used in kit components were tested and found to be negative for HIV, Hepatitis B and Hepatitis C. However, for safety reasons, all kit components should be treated as potentially infectious.

- Kit reagents contain sodium azide or ProClin as bactericides. Sodium azide and ProClin are harmful to health and the environment. Substrates for enzymatic color reactions can also cause skin and/or respiratory irritation. Any contact with the substances should be avoided. Further safety information can be found in the safety data sheet, which is available from Immundiagnostik AG on request.
- The 10x Wash buffer concentrate (WASHBUF) contains surfactants which may cause severe eye irritation in case of eye contact. **Warning:** Causes serious eye irritation. **IF IN EYES:** Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing. If eye irritation persists: get medical Advice/attention.
- The stop solution consists of diluted sulphuric acid, a strong acid. Although diluted, it still must be handled with care. It can cause burns and should be handled with gloves, eye protection, and appropriate protective clothing. Any spill should be wiped out immediately with copious quantities of water. Do not breath vapour and avoid inhalation.

## **13. TECHNICAL HINTS**

- Do not interchange different lot numbers of any kit component within the same assay. Furthermore we recommend not to assemble wells of different microtiter plates for analysis, even if they are of the same batch as wells from already opened microtiter plates are exposed to different conditions as sealed ones.
- Control samples should be analysed with each run.
- Reagents should not be used beyond the expiration date stated on kit label.
- Substrate solution should remain colourless until use.
- To ensure accurate results, proper adhesion of plate sealers during incubation steps is necessary.
- Avoid foaming when mixing reagents.
- Do not mix plugs and caps from different reagents.
- The assay should always be performed according to the enclosed manual.

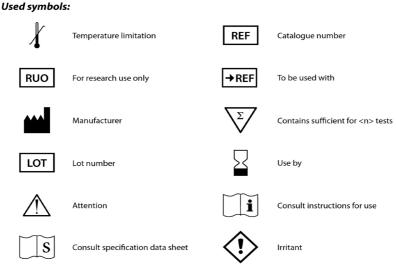
## 14. GENERAL NOTES ON THE TEST AND TEST PROCEDURE

• Quality control guidelines should be followed.

- IDK monitor<sup>®</sup> is a trademark of Immundiagnostik AG.
- Incubation time, incubation temperature and pipetting volumes of the components are defined by the producer. Any variation of the test procedure, which is not coordinated with the producer, may influence the results of the test. Immundiagnostik AG can therefore not be held responsible for any damage resulting from wrong use.
- Warranty claims and complaints in respect of deficiencies must be logged within 14 days after receipt of the product. The product should be send to Immundiagnostik AG along with a written complaint.

## 15. REFERENCES

- 1. Scherl EJ, Kumar S, Warren RU. Review of the safety and efficacy of ustekinumab. *Therapeutic Advances in Gastroenterology*. 2010;**3**(5):321–8.
- Chiu HY, Chu TW, Cheng YP, Tsai TF. The association between clinical response to ustekinumab and immunogenicity to ustekinumab and prior adalimumab. *PLoS ONE*. 2015;**10**(11):1–10.
- Ordás I, Mould DR, Feagan BG, Sandborn WJ. Anti-TNF monoclonal antibodies in inflammatory bowel disease: pharmacokinetics-based dosing paradigms. *Clinical pharmacology and therapeutics*. 2012/02/24. 2012 Apr;**91**(4):635–46.



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