S-Nitrosylated Protein Detection Assay Kit
Item No. 10006518
Materials Supplied

Kit will arrive packaged as a -20°C kit, for best results, remove components and store as stated below.

<table>
<thead>
<tr>
<th>Item Number</th>
<th>Item</th>
<th>Quantity/Size</th>
<th>Storage</th>
</tr>
</thead>
<tbody>
<tr>
<td>10006519</td>
<td>S-Nitrosylation Wash Buffer (10X)</td>
<td>1 vial/25 ml</td>
<td>Room Temperature</td>
</tr>
<tr>
<td>10006520</td>
<td>S-Nitrosylation Buffer A</td>
<td>1 vial/30 ml</td>
<td>Room Temperature</td>
</tr>
<tr>
<td>10006521</td>
<td>S-Nitrosylation Buffer B (5X)</td>
<td>1 vial/60 ml</td>
<td>Room Temperature</td>
</tr>
<tr>
<td>10007142</td>
<td>S-Nitrosylation Blocking Reagent</td>
<td>3 vials</td>
<td>-20°C</td>
</tr>
<tr>
<td>10006523</td>
<td>S-Nitrosylation Labeling Reagent</td>
<td>3 vials</td>
<td>-20°C</td>
</tr>
<tr>
<td>10006522</td>
<td>S-Nitrosylation Reducing Reagent</td>
<td>3 vials</td>
<td>Room Temperature</td>
</tr>
<tr>
<td>10006524</td>
<td>S-Nitrosylation Detection Reagent I (HRP)</td>
<td>1 vial</td>
<td>-20°C</td>
</tr>
<tr>
<td>10006525</td>
<td>S-Nitrosylation Detection Reagent II (Fluorescein)</td>
<td>1 vial</td>
<td>-20°C</td>
</tr>
</tbody>
</table>

If any of the items listed above are damaged or missing, please contact our Customer Service department at (800) 364-9897 or (734) 971-3335. We cannot accept any returns without prior authorization.

WARNING: This product is for laboratory research use only: not for administration to humans. Not for human or veterinary diagnostic or therapeutic use.
Precautions

Please read these instructions carefully before beginning this assay. For research use only. Not for human or diagnostic use.

If You Have Problems

Technical Service Contact Information

Phone: 888-526-5351 (USA and Canada only) or 734-975-3888
Fax: 734-971-3641
Email: techserv@caymanchem.com
Hours: M-F 8:00 AM to 5:30 PM EST

In order for our staff to assist you quickly and efficiently, please be ready to supply the lot number of the kit (found on the outside of the box).

Storage and Stability

This kit will perform as specified if stored and used before the expiration date indicated on the outside of the box. Once this kit has been opened, store the vials of Reducing Reagent, Wash Buffer, Buffer A, and Buffer B at room temperature. Store the remaining components at -20°C.

Materials Needed But Not Supplied

1. Acetone, *HAZARDOUS SOLVENT*
2. Dimethylformamide (DMF), *HAZARDOUS SOLVENT*
3. Pipettes, 1.5 ml microcentrifuge and 15 ml tubes (polypropylene)
4. 10 ml graduated cylinders (2)
5. A Benchtop Centrifuge and Microcentrifuge (at 4°C)
6. Equipment for performing transblotting

Background

Nitric oxide (NO) is produced by three distinct isoforms of nitric oxide synthase and functions as a key signaling molecule in physiology and pathophysiology.\(^1,2\) NO transduces its effects by reacting either directly with heme and non-heme centers of proteins or indirectly *via* further oxidation to various reactive nitrogen species (RNS).\(^3,4\) Nitrosylation refers to the binding of an NO group to a transition metal, as typified in the activation of soluble guanylate cyclase, or to a thiol (SH) group of protein cysteine residues resulting in the formation of an S-NO moiety. This latter reaction, termed S-nitrosylation, is mediated by reactive nitrogen species of higher oxidation states of NO, such as NO\(_2\) and N\(_2\)O\(_3\).\(^5,6\) S-nitrosylation is a reversible, and seemingly specific, post-translational modification that regulates the activity of a large number of targets, including metabolic, structural, cytoskeletal, and signaling proteins.\(^7\)

About This Assay

Cayman's S-Nitrosylated Protein Detection Assay employs a modification of the Jaffrey *et al.* "Biotin-switch" method to allow for the direct visualization of S-nitrosylated proteins in whole cells or tissues, as well as by western blot analysis.\(^5,9\) Using this method, free SH groups are first blocked (an addition of 125.1 amu per residue) and any S-NO bonds present in the sample are then cleaved. Biotinylation of the newly formed SH groups (an addition of 523.6 amu per residue) provides the basis for visualization using streptavidin-based colorimetric or fluorescence detection.
1. **S-Nitrosylation Wash Buffer (10X) - (Item No. 10006519)**
   Bring to room temperature. Dilute the contents of the Wash Buffer vial (25 ml) to a final volume of 250 ml with purified water. This should be done before starting the assay(s). Store this buffer at room temperature.

2. **S-Nitrosylation Buffer A - (Item No. 10006520)**
   Bring to room temperature. Buffer A is ready to use as supplied. Store this buffer at room temperature.

3. **S-Nitrosylation Buffer B (5X) - (Item No. 10006521)**
   Bring to room temperature. Dilute the contents of the Buffer B vial (60 ml) to a final volume of 300 ml with purified water. Store this buffer at room temperature.

4. **S-Nitrosylation Blocking Reagent - (Item No. 10007142)**
   Supplied as a crystalline solid. Three vials are provided to allow up to three separate experiments. Store the Blocking Reagent as supplied at -20°C. Use fresh Blocking Reagent for each experiment. Storage and later use of a reconstituted solution is not advised.

5. **S-Nitrosylation Labeling Reagent - (Item No. 10006523)**
   Supplied as a lyophilized powder. Three vials are provided to allow for up to three separate experiments. Store the Labeling Reagent as supplied at -20°C. Use fresh Reducing Reagent for each experiment. Storage and later use of the reconstituted reagent is not advised.

6. **S-Nitrosylation Reducing Reagent - (Item No. 10006522)**
   Supplied as a lyophilized powder. Three vials are provided to allow up to three separate experiments. Store the Reducing Reagent as supplied at room temperature. Use fresh Reducing Reagent for each experiment. Storage and later use of the reconstituted reagent is not advised.

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**Figure 1. Diagram of the three-step process of protein nitrosothiol (S-NO) derivitization with biotin.** Protein free thiols are derivitized with a blocking agent. S-Nitrosothiols are reduced to yield free thiol(s) which are covalently labeled with maleimide-biotin. Subsequent detection by avidin-coupled reagents can be used to localize the biotinylated protein(s).
7. **S-Nitrosylation Detection Reagent I (HRP) - (Item No. 10006524)**

Supplied as a lyophilized powder. Reconstitute the contents of the vial with 400 μl of distilled water and mix gently to dissolve the powder. Use at a 1:75 dilution for blotting overlay experiments or tissue sections. *Store the reconstituted reagent at 4°C for up to six months.*

8. **S-Nitrosylation Detection Reagent II (Fluorescein) - (Item No. 10006525)**

Supplied as a lyophilized powder. Prepare with 1 ml of Buffer B containing 0.1% Triton X-100 and use at a 1:40 dilution. *Store the reconstituted reagent at 4°C for up to six months.*

### Sample Preparation

Sufficient reagents are supplied for three experiments with up to 20 samples per experiment.

<table>
<thead>
<tr>
<th>Assay Summary</th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>Cell Washing, Lysis, and Blocking</td>
<td>2-3 hours</td>
</tr>
<tr>
<td>Indirect S-NO Labeling</td>
<td>2.5 hours</td>
</tr>
<tr>
<td>Total Time</td>
<td>4.5-5.5 hours</td>
</tr>
</tbody>
</table>

**NOTE:** Steps 1-12 of this procedure are performed under indirect light. *This assay is recommended for preparation of up to 20 cell pellets (each ≤0.1 g wet weight).*

1. **Substrate-dependent cells:** Wash adherent cultured-cells with three successive aliquots of cold Wash Buffer using a fraction of the last wash to scrape and transfer the cells into a 50 ml conical tube. Generate a cell pellet by centrifugation (500 x g, five minutes) and carefully remove any remaining wash buffer above the cell pellet.

   Cell pellet from suspension culture or isolated from tissue: Add Wash Buffer and resuspend the pellet. Invert the tube to mix and generate a cell pellet by centrifugation (500 x g, five minutes). Decant the supernatant and resuspend cells in another aliquot of Wash buffer and generate a cell pellet as before. Repeat the process once more.

2. Carefully remove any residual Wash Buffer with a pipette. Keep all samples on ice.

3. Dissolve the contents of 1 vial of Blocking Reagent with 100 μl DMF and then add 900 μl of Buffer A to the vial. Close the vial and mix well by inversion. Add 1 ml of the reconstituted reagent to 9 ml Buffer A and mix well; this is now **Buffer A containing Blocking Reagent**.

4. To each cell pellet add 5 volumes (up to 0.5 ml total volume) of **Buffer A containing Blocking Reagent** and tap the side of each tube to resuspend the cells. Transfer the samples to prelabeled microcentrifuge tubes. Place the samples on a shaker or rocker (with gentle agitation) for 30 minutes at 4°C.

5. Generate clarified lysates by centrifugation of the samples for 10 minutes at 4°C with a tabletop microcentrifuge.

6. Transfer the supernatants into separate cold, pre-labeled 15 ml conical polypropylene tubes and add 4 volumes of ice-cold acetone to each sample.

7. Mix each sample thoroughly by inversion followed by incubation at -20°C for one hour.

8. Remove the samples from -20°C and produce a compact protein pellet by centrifugation at 3,000 x g for 10 minutes at 4°C. (Proceed to step 9 during centrifugation.)

9. During centrifugation, reconstitute the contents of 1 vial of S-Nitrosylation Reducing Reagent with 1 ml of Buffer B. Also, dissolve the contents of 1 vial of S-Nitrosylation Labeling Reagent with 100 μl of DMF followed by 900 μl of Buffer B, mixing well. When each reagent is fully dissolved, add 0.5 ml of the Reducing Reagent and 0.5 ml of the Labeling Reagent into a graduated cylinder containing 4 ml of Buffer B. This is called **Buffer B containing Reducing and Labeling Reagents**. Mix it well by inversion.

10. Remove the samples from the centrifuge and decant the supernatants from each tube (containing primarily acetone) into the appropriate organic solvent waste container. Set the protein pellet-containing tubes on ice. Remove residual acetone from pellets with a pipette.

11. Resuspend each protein pellet with 0.5 ml of **Buffer B containing Reducing and Labeling Reagents**.
Control Sample: Contribution of signal from endogenously biotinylated proteins can be observed by resuspension of control cell pellets in Buffer B alone.

12. Incubate these samples for one hour at room temperature.
13. Add 4 volumes (2 ml) of ice-cold acetone to each sample, mixing each thoroughly by inversion followed by incubation at -20°C for one hour.
14. Remove the samples from -20°C and produce a compact protein pellet by centrifugation at 3,000 x g for 10 minutes at 4°C.
15. Decant the acetone-containing supernatant into a solvent waste container and set the protein pellet samples on ice.
16. Resuspend each of the protein pellets in a minimal volume (100-200 μl suggested) of cold Wash Buffer. Reserve a small volume of each for protein determination and store the rest at -20°C for analysis by western blotting, immunoprecipitation of a specific protein, or anti-biotin immunoprecipitation for concentration of nitrosylated proteins.

**Avidin Overlay/Western Blotting Notes:**
1. Prepare protein samples for SDS-PAGE as is done for western blotting; add Laemmli buffer, mix, and incubate samples in a boiling water bath for five minutes followed by five minutes on ice prior to loading. The biotin label is covalently bound.
2. Do not use milk products for blocking unoccupied surfaces of the transfer-membranes because the endogenous biotin content may cause high background signal or no signal from experimental samples when probing with the avidin detection reagents. Solutions of 2% bovine serum albumin (BSA) in PBS or TBS buffers are acceptable for blocking membranes and dilution of Detection Reagent I (HRP) in this assay.
3. Reconstitute the contents of the S-Nitrosylation Detection Reagent I (HRP) vial with 400 μl of distilled water. Use at a 1:75 dilution for overlay experiment (similar to western blotting). Develop the membrane(s) with Enhanced Chemiluminescence (ECL) Reagents or other peroxidase compatible substrate. Alternatively, S-Nitrosylation Detection Reagent II (Fluorescein) may be used in place of Detection Reagent I (HRP) if the appropriate detection equipment is available.
Materials Needed But Not Supplied (Cells)

1. 6-, 12-, 24-, or 96-well plates for cell culture
2. Humidified chamber
3. Paraformaldehyde
4. Dimethylformamide (DMF), A HAZARDOUS SOLVENT
5. Triton X-100
6. Confocal microscope with fluorescence source
7. AquaPoly/Mount (optional)
8. Nuclear counterstain (optional); for example, Propidium Iodide (PI) or 4’6-Diamidino-2-phenylindole (DAPI), Caution: these chemicals are probable MUTAGENS

Performing the Assay (Cells)

NOTE: Perform steps 2-11 of this procedure are done under indirect light at room temperature unless otherwise stated.

1. Prepare enough Wash Buffer containing 4% paraformaldehyde to fix the cells but not more than needed to allow for the washing steps and separate experiments.
2. Remove media from culture wells and rinse each briefly with Wash Buffer three times.
3. Fix the cells by incubation with Wash Buffer containing 4% paraformaldehyde for 20 minutes.
4. Wash cells 3 x 5 minutes with Wash Buffer. Let the last wash remain on the cells and proceed to step 5.
5. Prepare the buffer for step 6 immediately before use. To make Buffer B containing Blocking Reagent and 0.1% Triton X-100, dissolve the contents of one vial of blocking reagent with 100 μl of DMF; and then add 900 μl of Buffer B. Transfer the dissolved Blocking reagent into a 20 ml graduated cylinder, add 10 μl Triton X-100 and bring to 10 ml final volume with Buffer B. Mix this buffer well by inversions.
6. Remove Wash Buffer from the cells, and incubate cells with Buffer B containing Blocking Reagent and 0.1% Triton X-100 for 30 minutes at 4°C.
7. Wash cells 3 x 5 minutes with Buffer B containing 0.1% Triton X-100. Let the last wash remain on the cells and proceed to step 8.
8. Prepare the Reducing/Labeling buffer immediately before use by reconstituting 1 vial of S-Nitrosylation Reducing Reagent with 1 ml of Buffer B containing 0.1% Triton X-100. Also, dissolve the contents of 1 vial of S-Nitrosylation Labeling Reagent with 100 μl of dimethylformamide (DMF) followed by 900 μl of Buffer B containing 0.1% Triton X-100, mixing well. When each reagent is fully dissolved, transfer all of the Reducing Reagent and all of the Labeling Reagent into a graduated cylinder containing 8 ml of Buffer B containing 0.1% Triton X-100. This is called Buffer B containing 0.1% Triton X-100, Reducing and Labeling Reagents. Mix it well by inversion. Remove the Wash Buffer from the cells and add enough Buffer B containing 0.1% Triton X-100, Reducing and Labeling Reagents to cover the cells. Incubate for one hour at room temperature.
9. Wash cells 3 x 5 minutes with Buffer B containing 0.1% Triton X-100.
10. Remove the last wash and incubate the cells with Buffer B containing 0.1% Triton X-100 and 1:40 dilution of Detection Reagent II (Fluorescein) for one hour in a humidified chamber.
11. Wash cells 3 x 5 minutes with Buffer B containing 0.1% Triton X-100. The cells may be observed by fluorescence microscopy (λ_{EX} = 490 nm, λ_{EM} = 520).

Optional technique: Stain nuclei with propidium iodide (10 µg/ml solution) for 30 minutes in a humidified chamber. Wash away excess propidium iodide with Buffer B containing 0.1% Triton X-100 and examine by fluorescence microscopy.
Detection of S-Nitrosylated Proteins

Sufficient reagents are supplied for three separate experiments of up to 25 samples.

### Assay Summary and Timeframe

<table>
<thead>
<tr>
<th>Step</th>
<th>Time Frame</th>
</tr>
</thead>
<tbody>
<tr>
<td>Tissue Preparation</td>
<td>up to 2 hours</td>
</tr>
<tr>
<td>Wash/Fix Sections</td>
<td>45 minutes</td>
</tr>
<tr>
<td>Block/Label S-Nitrosylation</td>
<td>90 minutes</td>
</tr>
<tr>
<td>Detection Reagent Incubation</td>
<td>75 minutes (or overnight)</td>
</tr>
<tr>
<td>Counterstain/Wash</td>
<td>60 minutes</td>
</tr>
<tr>
<td>Total Time</td>
<td>&lt;5 hours</td>
</tr>
</tbody>
</table>

**Materials Needed But Not Supplied (Tissue)**

1. Embedded frozen tissue samples
2. Tissue and histology preparation equipment (Tissue cassette, Embedding Solution, liquid nitrogen, cryostat, microscope slides)
3. Paraformaldehyde
4. Triton X-100
5. Dimethylformamide (DMF), *Caution: HAZARDOUS SOLVENT*
6. Confocal microscope with fluorescence source
7. PAP Pen (optional)
8. Nuclear counterstain (optional); for example, Propidium Iodide (PI) or 4′,6-Diamidino-2-phenylindole (DAPI), *Caution: these chemicals are probable MUTAGENS*

**General Information**

- Perform all steps of this procedure under indirect light only.
- Preparation of fresh tissue sections is highly recommended due to the transient nature of the S-nitrosothiol moiety. However, if tissue sections have been prepared previously, skip ahead to step 3.
Performing the Assay (Tissue)

1. Infuse or rinse organ tissues with cold Wash Buffer for 10 minutes.

2. Use your standard protocol to perform tissue embedding, freezing, section cutting, and slide fixation.

3. Wash sections 3 x 5 minutes with Wash Buffer. Circumscribe tissues with PAP Pen if desired.

4. Fix sections in Wash Buffer containing 4% paraformaldehyde for 30 minutes at room temperature.

5. Wash sections 3 x 5 minutes with Wash Buffer containing 1% Triton X-100.

6. Incubate sections with Wash Buffer containing Blocking Reagent and 1% Triton X-100 for 30 minutes at 4°C.

7. Wash sections 3 x 5 minutes with Wash Buffer containing 1% Triton X-100.

8. Incubate sections in Buffer B alone (as the endogenous biotinylated protein control) or Buffer B containing 1% Triton X-100, Reducing and Labeling Reagents for one hour at room temperature.

   If not already done, prepare the Buffer for step 10 during this time: resuspend the contents of one vial of S-Nitrosylation Detection Reagent II (Fluorescein) with 1 ml of Buffer B containing 1% Triton X-100. Once dissolved keep this reagent on ice.

9. Wash the sections 3 x 5 minutes with Buffer B containing 1% Triton X-100.

10. Use the S-Nitrosylation Detection Reagent II (Fluorescein) at a 1:40 dilution in Buffer B containing 1% Triton X-100. Incubate the sections for one hour at room temperature or overnight at 4°C (wrapped in foil or other dark container).

11. Wash the sections 3 x 5 minutes with Buffer B containing 1% Triton X-100.

   Optional: Counterstain the sections (nuclei) with DAPI or PI as suggested by the product manufacturer. Wash away excess counterstain with Buffer B containing 1% Triton X-100.

12. View stained sections by confocal fluorescence microscopy ($\lambda_{\text{EX}} = 490$ nm, $\lambda_{\text{EM}} = 520$).
Troubleshooting

<table>
<thead>
<tr>
<th>Problem</th>
<th>Possible Causes</th>
<th>Recommended Solutions</th>
</tr>
</thead>
<tbody>
<tr>
<td>High background</td>
<td>A. Insufficient blocking of free thiols</td>
<td>A. Increase the incubation time of the blocking step</td>
</tr>
<tr>
<td></td>
<td>B. Excessive detection reagent for the test</td>
<td>B. Increase the dilution factor of the detection reagent to 1:1,000-1:2,000</td>
</tr>
<tr>
<td>Nothing detected</td>
<td>A. Low levels of endogenous S-NO may not be detected</td>
<td>A. Comparison of results to positive and negative controls will provide evidence for further investigation into the source of the problem</td>
</tr>
<tr>
<td></td>
<td>B. Poor protein recovery</td>
<td>B. More protein may need to be loaded in gels</td>
</tr>
</tbody>
</table>

References


Related Products

- Aconitase Fluorometric Assay Kit - Item No. 700600
- Ascorbate Assay Kit - Item No. 700420
- Nitrate/Nitrite Colorimetric Assay Kit (LDH Method) - Item No. 760871
- Nitrate/Nitrite Fluorometric Assay Kit - Item No. 780051
- Protein Carbonyl Colorimetric Assay Kit - Item No. 10005020
- Superoxide Dismutase Assay Kit - Item No. 706002

For a complete list of related products please visit: www.caymanchem.com/catalog/10006518

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Said refund or replacement is conditioned on Buyer giving written notice to Cayman within thirty (30) days after arrival of the material at its destination. Failure of Buyer to give said notice within thirty (30) days shall constitute a waiver by Buyer of all claims hereunder with respect to said material.

For further details, please refer to our Warranty and Limitation of Remedy located on our website and in our catalog.